

RESEARCH

Open Access



Screening and identification of key biomarkers associated with endometriosis using bioinformatics and next-generation sequencing data analysis

Basavaraj Vastrad¹ and Chanabasayya Vastrad^{2*}

Abstract

Background Endometriosis is a common cause of endometrial-type mucosa outside the uterine cavity with symptoms such as painful periods, chronic pelvic pain, pain with intercourse and infertility. However, the early diagnosis of endometriosis is still restricted. The purpose of this investigation is to identify and validate the key biomarkers of endometriosis.

Methods Next-generation sequencing dataset GSE243039 was obtained from the Gene Expression Omnibus database, and differentially expressed genes (DEGs) between endometriosis and normal control samples were identified. After screening of DEGs, gene ontology (GO) and REACTOME pathway enrichment analyses were performed. Furthermore, a protein–protein interaction (PPI) network was constructed and modules were analyzed using the Human Integrated Protein–Protein Interaction rEference database and Cytoscape software, and hub genes were identified. Subsequently, a network between miRNAs and hub genes, and network between TFs and hub genes were constructed using the miRNet and NetworkAnalyst tool, and possible key miRNAs and TFs were predicted. Finally, receiver operating characteristic curve analysis was used to validate the hub genes.

Results A total of 958 DEGs, including 479 upregulated genes and 479 downregulated genes, were screened between endometriosis and normal control samples. GO and REACTOME pathway enrichment analyses of the 958 DEGs showed that they were mainly involved in multicellular organismal process, developmental process, signaling by GPCR and muscle contraction. Further analysis of the PPI network and modules identified 10 hub genes, including *vcam1*, *snca*, *prkcb*, *adrb2*, *foxq1*, *mdfi*, *actb12*, *prkd1*, *dapk1* and *actc1*. Possible target miRNAs, including *hsa-mir-3143* and *hsa-mir-2110*, and target TFs, including *tcf3* (transcription factor 3) and *clock* (clock circadian regulator), were predicted by constructing a miRNA-hub gene regulatory network and TF-hub gene regulatory network.

Conclusions This investigation used bioinformatics techniques to explore the potential and novel biomarkers. These biomarkers might provide new ideas and methods for the early diagnosis, treatment and monitoring of endometriosis.

Keywords Bioinformatics, Biomarkers, Differentially expressed genes, Endometriosis, Hub genes, Next-generation sequencing, Signaling pathways

*Correspondence:

Chanabasayya Vastrad
channu.vastrad@gmail.com

Full list of author information is available at the end of the article



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

Background

Endometriosis is one of the most important chronic inflammatory diseases and has become the key cause of serious reproductive and general health condition [1]. Endometriosis is characterized by the presence of endometrial-type mucosa outside the uterine cavity [2]. The outbreak of their first period occurs through menopause, disregarding of ethnic origin or social status [2]. The clinical incidence of endometriosis is high, and its main features include dysmenorrhea, dyspareunia, chronic pelvic pain, irregular uterine bleeding and infertility [3], which places a great burden on the economy of health and reduces quality of life in worldwide; 10% of women of reproductive age are diagnosed with endometriosis each year [4]. These patients might have various complications including gynecological cancer (ovarian, endometrial and cervical cancers) [5], polycystic ovary syndrome [6], cardiovascular diseases [7], obesity [8], gestational diabetes mellitus [9], diabetes mellitus [10] and hypertension [11]. Endometriosis is recognized to be vulnerable on sex hormone estrogen, which rise the inflammation, growth and pain linked with the disease [6]. Studies have revealed that the progression of endometriosis is related to genetic risk factors [12] as well as environmental factors [13]. Because of this disorder complex pathogenesis, it is mainly treated by gynecological surgery [14], oral contraceptives [15], progestins [16], nonsteroidal anti-inflammatory drugs [17] and gonadotropin-releasing hormone agonists [18]. But these treatments have not been effective for longer period. Therefore, it is urgent to find specific molecular biomarkers for early assessment of the prognosis of patients with endometriosis, so as to further advancing the treatment and prognosis of patients. In current years, molecular biomarkers were demonstrated highly useful as clinical tools for endometriosis diagnosis and treatment [19].

The underlying complex molecular mechanisms in endometriosis pose a special challenge to daily clinical practice. Studies have found that genes including *cyr61* [20], *esr2* and *cyp19a1* [21], *hoxa10* [22], *foxd3* [23], *lox11* and *htra1* [24] can be used as an important marker for early diagnosis, prognosis and treatment of endometriosis. Studies have shown that signaling pathways including AKT and ERK signaling pathways [25], Wnt/ β -catenin signaling pathway [26], PI3K-Akt-mTOR and MAPK signaling pathways [27], notch signaling pathway [28] and MAPK/ERK signal pathway [29] are involved in the progression of endometriosis. Taken together, current evidence suggests that the genes and signaling pathways are closely related to the progression of endometriosis.

The mechanisms of endometriosis at the molecular level are essentially for treating the disease. With the wide application of next-generation sequencing (NGS)

technology, endometriosis-related genes have been widely identified, which is a key step in exploring the complex pathology of endometriosis and finding drugs that combat the illness. Numerous NGS data of gene expression have been published in public databases such as NCBI Gene Expression Omnibus (GEO) [<https://www.ncbi.nlm.nih.gov/geo/>] [30] during the past few years, and they are being increasingly used in bioinformatics and NGS data analysis to explore target genes or proteins associated in various diseases [31, 32].

Bioinformatics and network analysis of NGS data are an effective way to explore biomarkers in the pathogenesis of various diseases. Therefore, this investigation aimed to use bioinformatics analysis to identify hub genes and molecular pathways involved in endometriosis, to identify key diagnostic or therapeutic biomarkers. We obtained DEGs between endometriosis and normal control samples from GSE243039, a gene expression profile downloaded from the GEO database. Immediately after, we performed gene ontology (GO) and REACTOME pathway enrichment analysis on these DEGs. By constructing PPI networks, we screened for the significant modules and hub genes. We constructed miRNA-hub gene regulatory network and TF-hub gene regulatory network, and we screened for the miRNAs, TFs and hub genes. To validate that these hub genes can serve as molecular markers of endometriosis, we determined hub genes by using receiver operating characteristic curve (ROC) analysis. This investigation might offer better insight into potential molecular mechanisms to explore preventive and therapeutic strategies for endometriosis.

Methods

Next-generation sequencing (NGS) data source

The NGS dataset GSE243039 was obtained from the GEO database (Accession Date: 11/09/2023). The GSE243039 dataset included 20 endometriosis samples and 20 normal control samples. The platform used was the GPL24676 Illumina NovaSeq 6000 (Homo sapiens).

Identification of DEGs

The limma R/Bioconductor software package [33] was used to perform the identification of DEGs between endometriosis samples and normal control samples. We adjusted *p*-value to correct the false positive error caused by the multiple tests and determined it by the Benjamini and Hochberg method [34], which is the common tools to minimize the false discovery rate. The cutoff criteria were $|\log_{2}FC| > 1.304$ (log₂ fold change) for upregulated genes, $|\log_{2}FC| > 1.304$ (log₂ fold change) < -1.2644 for downregulated genes and a *adj.P.Val* < 0.05 . Thereafter, we used R packages “ggplot2” and “gplot” to show the

Table 1 The statistical metrics for key differentially expressed genes (DEGs)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
PLPPR4	6.76335	6.20E-15	2.24E-11	12.15991	Up	Phospholipid phosphatase related 4
PCSK9	5.07288	5.03E-12	2.78E-09	9.713276	Up	Proprotein convertase subtilisin/kexin type 9
ADAMTS19	4.976936	1.00E-14	2.25E-11	11.97438	Up	ADAM metalloproteinase with thrombospondin type 1 motif 19
KIF26A	4.329184	9.75E-08	4.69E-06	6.504061	Up	Kinesin family member 26A
GRIA4	4.296838	2.87E-08	1.91E-06	6.886288	Up	Glutamate ionotropic receptor AMPA-type subunit 4
CNTN4	4.108296	6.99E-08	3.70E-06	6.608031	Up	Contactin 4
APCDD1	4.052813	4.17E-06	9.09E-05	5.335862	Up	APC downregulated 1
SEMA3A	4.047205	3.35E-07	1.28E-05	6.120174	Up	Semaphorin 3A
TMEM26	4.037968	1.59E-12	1.10E-09	10.11548	Up	Transmembrane protein 26
GRIN2A	4.022587	2.01E-08	1.44E-06	6.998121	Up	Glutamate ionotropic receptor NMDA-type subunit 2A
LAMC3	3.98438	7.26E-06	0.000141	5.162268	Up	Laminin subunit gamma 3
L1CAM	3.939758	1.85E-06	4.86E-05	5.589465	Up	L1 cell adhesion molecule
CBLN2	3.905839	1.06E-07	5.03E-06	6.477874	Up	Cerebellin 2 precursor
HPSE2	3.800586	2.29E-07	9.22E-06	6.238608	Up	Heparanase 2 (inactive)
CILP	3.794436	0.000108	0.001167	4.301899	Up	Cartilage intermediate layer protein
SDK1	3.794238	3.57E-07	1.34E-05	6.100051	Up	Sidekick cell adhesion molecule 1
ROBO2	3.793291	1.94E-09	2.38E-07	7.73845	Up	Roundabout guidance receptor 2
LPAR3	3.77025	2.07E-08	1.47E-06	6.988144	Up	Lysophosphatidic acid receptor 3
GALNT17	3.764892	1.28E-06	3.70E-05	5.703322	Up	Polypeptide N-acetylgalactosaminyltransferase 17
PALMD	3.746961	1.97E-06	5.08E-05	5.569086	Up	Palmelphin
JAKMIP2	3.740977	2.39E-09	2.79E-07	7.671792	Up	Janus kinase and microtubule interacting protein 2
SLC35F3	3.733243	4.47E-09	4.41E-07	7.472579	Up	Solute carrier family 35 member F3
CACNA1G	3.652448	1.35E-07	6.07E-06	6.403646	Up	Calcium voltage-gated channel subunit alpha1 G
AJAP1	3.630833	9.51E-06	0.000175	5.077432	Up	Adherens junctions-associated protein 1
RORB	3.592522	0.000505	0.003859	3.789292	Up	RAR-related orphan receptor B
HSD17B2	3.586894	5.44E-06	0.000112	5.252487	Up	Hydroxysteroid 17-beta dehydrogenase 2
PCDH19	3.462986	8.50E-08	4.28E-06	6.54683	Up	Protocadherin 19
S100A4	3.361421	7.83E-15	2.24E-11	12.06958	Up	S100 calcium binding protein A4
NLGN4X	3.346885	4.69E-05	0.000609	4.570634	Up	Neuroigin 4 X-linked
VCAM1	3.305329	0.000134	0.001387	4.230696	Up	Vascular cell adhesion molecule 1
STRA6	3.252045	0.0002	0.001881	4.098793	Up	Signaling receptor and transporter of retinol STRA6
COL6A6	3.23629	1.92E-08	1.39E-06	7.011592	Up	Collagen type VI alpha 6 chain
SIX2	3.233347	0.000242	0.002174	4.035541	Up	SIX homeobox 2
CLEC3B	3.219362	6.86E-08	3.64E-06	6.613763	Up	C-type lectin domain family 3 member B
ST8SIA4	3.210139	2.46E-06	6.00E-05	5.500511	Up	ST8 alpha-N-acetyl-neuraminide alpha-2,8-sialyltransferase 4
MMP3	3.201862	3.29E-05	0.000461	4.684818	Up	Matrix metalloproteinase 3
PLCH1	3.175137	0.000139	0.001428	4.217957	Up	Phospholipase C eta 1
CABCOCO1	3.141923	7.85E-12	3.96E-09	9.56007	Up	Ciliary-associated calcium binding coiled-coil 1
DPP6	3.141142	2.99E-06	6.98E-05	5.439696	Up	Dipeptidyl peptidase-like 6
COL28A1	3.064911	2.00E-05	0.000311	4.842314	Up	Collagen type XXVIII alpha 1 chain
COCH	3.05674	9.67E-06	0.000178	5.071932	Up	Cochlin
EFNA5	3.05662	5.11E-08	2.96E-06	6.705933	Up	Ephrin A5
GRP	3.056161	1.13E-08	9.17E-07	7.178938	Up	Gastrin releasing peptide

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
AVPR1A	3.052514	0.000105	0.001144	4.309966	Up	Arginine vasopressin receptor 1A
ADGRL3	3.042869	5.00E-06	0.000105	5.278659	Up	Adhesion G protein-coupled receptor L3
EPHA6	3.029344	1.31E-09	1.78E-07	7.865147	Up	EPH receptor A6
CNTN5	3.004363	4.15E-08	2.53E-06	6.770742	Up	Contactin 5
TUBA3E	3.003025	1.70E-05	0.000275	4.894684	Up	Tubulin alpha 3e
RGS7	2.977539	3.87E-10	7.39E-08	8.259352	Up	Regulator of G protein signaling 7
AQP8	2.973723	2.61E-09	2.99E-07	7.643696	Up	Aquaporin 8
WNT6	2.964101	1.91E-06	4.98E-05	5.57843	Up	Wnt family member 6
FABP4	2.95997	7.51E-08	3.90E-06	6.585421	Up	Fatty acid binding protein 4
PSG1	2.955976	0.000331	0.002783	3.931445	Up	Pregnancy-specific beta-1-glycoprotein 1
ANO1	2.949958	6.03E-08	3.33E-06	6.65401	Up	Anoctamin 1
SOX6	2.938951	2.83E-10	6.16E-08	8.361581	Up	SRY-box transcription factor 6
NDP	2.91885	0.000663	0.004802	3.696355	Up	Norrincystine knot growth factor NDP
ERVMER34-1	2.886936	3.13E-06	7.21E-05	5.425421	Up	Endogenous retrovirus group MER34 member 1, envelope
PREX2	2.876737	2.03E-06	5.18E-05	5.560168	Up	Phosphatidylinositol-3,4,5-trisphosphate-dependent Rac exchange factor 2
TFAP2C	2.864072	0.00077	0.005349	3.644851	Up	Transcription factor AP-2 gamma
NTRK1	2.855604	0.00041	0.003283	3.859591	Up	Neurotrophic receptor tyrosine kinase 1
CNTN1	2.853904	3.97E-05	0.000531	4.624175	Up	Contactin 1
GALNT13	2.842063	7.03E-05	0.000834	4.440753	Up	Polypeptide N-acetylgalactosaminyltransferase 13
SYT9	2.839815	3.44E-06	7.81E-05	5.395487	Up	Synaptotagmin 9
FGF10	2.839506	8.05E-10	1.29E-07	8.021417	Up	Fibroblast growth factor 10
RAMP3	2.832349	2.64E-06	6.33E-05	5.478637	Up	Receptor activity modifying protein 3
ALDH1L1	2.799	1.51E-06	4.19E-05	5.651506	Up	Aldehyde dehydrogenase 1 family member L1
PLA2G7	2.794211	1.73E-07	7.33E-06	6.326221	Up	Phospholipase A2 group VII
MMP12	2.792426	8.64E-05	0.000983	4.373945	Up	Matrix metalloproteinase 12
FAIM2	2.790032	1.17E-09	1.62E-07	7.902043	Up	Fas apoptotic inhibitory molecule 2
SEMA6A	2.781374	8.57E-10	1.34E-07	8.001094	Up	Semaphorin 6A
ADCY1	2.747518	0.000609	0.004472	3.725191	Up	Adenylatecyclase 1
MDGA2	2.743823	4.06E-07	1.50E-05	6.060738	Up	MAM domain containing glycosylphosphatidylinositol anchor 2
COL14A1	2.740007	1.47E-09	1.93E-07	7.827594	Up	Collagen type XIV alpha 1 chain
LCP1	2.739277	2.89E-06	6.80E-05	5.450181	Up	Lymphocyte cytosolic protein 1
MMP27	2.734557	8.62E-07	2.69E-05	5.826621	Up	Matrix metalloproteinase 27
LRP1B	2.727044	6.97E-07	2.30E-05	5.892855	Up	LDL receptor-related protein 1B
LY6K	2.697355	1.17E-05	0.000206	5.012773	Up	Lymphocyte antigen 6 family member K
PIP5K1B	2.685746	3.33E-09	3.59E-07	7.566542	Up	Phosphatidylinositol-4-phosphate 5-kinase type 1 beta
ZNF726	2.675392	2.65E-08	1.81E-06	6.911657	Up	Zinc finger protein 726
ADRA1D	2.663313	5.86E-07	2.03E-05	5.946796	Up	Adrenoceptor alpha 1D
SNCA	2.646582	0.002632	0.013959	3.210533	Up	Synuclein alpha
ST6GALNAC3	2.636022	1.03E-07	4.92E-06	6.486685	Up	ST6 N-acetylgalactosaminide alpha-2,6-sialyltransferase 3
TMEM52B	2.627669	1.79E-05	0.000287	4.877499	Up	Transmembrane protein 52B
TMEM37	2.616893	0.000162	0.001595	4.169509	Up	Transmembrane protein 37
CLIC2	2.612803	2.04E-09	2.45E-07	7.722159	Up	Chloride intracellular channel 2
NAALAD2	2.599709	9.33E-08	4.57E-06	6.517728	Up	N-acetylated alpha-linked acidic dipeptidase 2
LMTK3	2.580405	1.71E-07	7.29E-06	6.32841	Up	Lemur tyrosine kinase 3

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
STAR	2.573317	1.36E-05	0.000232	4.965242	Up	Steroidogenic acute regulatory protein
ACP3	2.532701	8.27E-06	0.000158	5.121323	Up	Acid phosphatase 3
IL31RA	2.523753	1.20E-09	1.64E-07	7.893868	Up	Interleukin 31 receptor A
APOD	2.520315	2.01E-05	0.000312	4.840658	Up	Apolipoprotein D
ANO4	2.518658	0.002076	0.011589	3.296307	Up	Anoctamin 4
LRATD1	2.518289	5.35E-11	1.62E-08	8.911076	Up	LRAT domain containing 1
INSRR	2.495802	0.000526	0.003994	3.774937	Up	Insulin receptor-related receptor
LAG3	2.494584	3.41E-05	0.000473	4.672734	Up	Lymphocyte activating 3
SOX18	2.481299	3.15E-06	7.25E-05	5.423675	Up	SRY-box transcription factor 18
F2RL2	2.478309	2.13E-06	5.38E-05	5.545123	Up	Coagulation factor II thrombin receptor-like 2
TMEM132B	2.476601	1.49E-05	0.00025	4.934886	Up	Transmembrane protein 132B
CCR1	2.473974	1.88E-05	0.000297	4.862981	Up	C-C motif chemokine receptor 1
NCKAP5	2.473105	0.000694	0.004962	3.680433	Up	NCK-associated protein 5
C2orf88	2.472711	2.18E-10	5.04E-08	8.446715	Up	Chromosome 2 open reading frame 88
ICA1	2.471974	1.03E-06	3.11E-05	5.77108	Up	Islet cell autoantigen 1
CST1	2.470675	0.000727	0.00514	3.664434	Up	Cystatin SN
NKAIN1	2.460596	0.001948	0.011011	3.319084	Up	Sodium/potassium transporting ATPase interacting 1
IP6K3	2.457598	0.000285	0.002469	3.981865	Up	Inositol hexakisphosphate kinase 3
RETREG1	2.457043	1.48E-06	4.10E-05	5.659324	Up	Reticulophagy regulator 1
CYP39A1	2.447117	1.10E-06	3.28E-05	5.75072	Up	Cytochrome P450 family 39 subfamily A member 1
S100A3	2.444216	9.86E-15	2.25E-11	11.98095	Up	S100 calcium binding protein A3
ISM1	2.438578	2.07E-05	0.000319	4.831004	Up	Isthmin 1
CHST1	2.432231	2.51E-06	6.10E-05	5.493885	Up	Carbohydrate sulfotransferase 1
ATCAY	2.424636	1.58E-06	4.30E-05	5.63875	Up	ATCAY kinesin light chain interacting caytaxin
PREX1	2.422015	7.44E-07	2.40E-05	5.872563	Up	Phosphatidylinositol-3,4,5-trisphosphate-dependent Rac exchange factor 1
CARD16	2.415338	6.30E-06	0.000126	5.206741	Up	Caspase recruitment domain family member 16
ADRA2C	2.405717	0.001481	0.008888	3.41651	Up	Adrenoceptor alpha 2C
FLT1	2.40555	4.08E-05	0.000544	4.615809	Up	Fms-related receptor tyrosine kinase 1
FABP5	2.404621	2.66E-08	1.81E-06	6.910337	Up	Fatty acid binding protein 5
PRDM1	2.398435	0.003852	0.018662	3.070723	Up	PR/SET domain 1
TRPC3	2.393228	1.51E-12	1.10E-09	10.13214	Up	Transient receptor potential cation channel subfamily C member 3
TUNAR	2.38405	1.98E-06	5.08E-05	5.568021	Up	TCL1 upstream neural differentiation-associated RNA
IGSF3	2.379319	1.48E-09	1.93E-07	7.825786	Up	Immunoglobulin superfamily member 3
THSD7A	2.376559	0.00529	0.024026	2.951945	Up	Thrombospondin type 1 domain containing 7A
ADAMTS17	2.37201	1.42E-06	3.98E-05	5.672089	Up	ADAM metallopeptidase with thrombospondin type 1 motif 17
DKK2	2.371245	1.51E-06	4.17E-05	5.653171	Up	Dickkopf WNT signaling pathway inhibitor 2
PROK1	2.361571	0.012248	0.045333	2.625366	Up	Prokineticin 1
SCTR	2.355346	2.76E-06	6.58E-05	5.464606	Up	Secretin receptor
FOXL2NB	2.341574	0.001967	0.0111	3.315561	Up	FOXL2 neighbor
WNT16	2.336909	0.002059	0.011516	3.299158	Up	Wnt family member 16
ZNF804A	2.332815	0.000402	0.003239	3.865913	Up	Zinc finger protein 804A
RNF157	2.321389	0.000916	0.006147	3.584773	Up	Ring finger protein 157

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
FILNC1	2.313159	2.61E-10	5.75E-08	8.387101	Up	FOXO-induced long non-coding RNA 1
COL5A3	2.302619	3.26E-08	2.11E-06	6.846013	Up	Collagen type V alpha 3 chain
F13A1	2.302086	3.45E-05	0.000476	4.669571	Up	Coagulation factor XIII A chain
SH3GL2	2.296048	7.55E-10	1.23E-07	8.042216	Up	SH3 domain containing GRB2-like 2, endophilin A1
PRKCB	2.295018	0.0001	0.001103	4.326301	Up	Protein kinase C beta
DHCR24	2.292674	1.76E-12	1.18E-09	10.07839	Up	24-dehydrocholesterol reductase
GPR20	2.290641	1.69E-06	4.53E-05	5.617085	Up	G protein-coupled receptor 20
PDE3B	2.290631	5.46E-08	3.11E-06	6.68473	Up	Phosphodiesterase 3B
CHST2	2.281982	1.57E-07	6.76E-06	6.355455	Up	Carbohydrate sulfotransferase 2
BMP4	2.278367	9.72E-13	7.87E-10	10.28843	Up	Bone morphogenetic protein 4
IL1RL1	2.271743	0.013247	0.048129	2.59379	Up	Interleukin 1 receptor-like 1
MYPN	2.259711	9.70E-06	0.000178	5.071057	Up	Myopalladin
HS3ST5	2.254222	3.44E-07	1.31E-05	6.111664	Up	Heparan sulfate-glucosamine 3-sulfotransferase 5
PLCG2	2.246402	2.57E-13	3.06E-10	10.76548	Up	Phospholipase C gamma 2
KCNH5	2.245021	8.24E-07	2.60E-05	5.840638	Up	Potassium voltage-gated channel subfamily H member 5
PRL	2.243593	0.01139	0.042843	2.654416	Up	Prolactin
FOXQ1	2.24188	0.001101	0.007076	3.520892	Up	Forkhead box Q1
ALPP	2.240977	5.86E-08	3.29E-06	6.662879	Up	Alkaline phosphatase, placental
SEMA6D	2.239721	0.000164	0.001613	4.164838	Up	Semaphorin 6D
NEFL	2.235525	0.000173	0.001684	4.147159	Up	Neurofilament light chain
WNT5A	2.234375	0.000779	0.005402	3.640615	Up	Wnt family member 5A
MACC1	2.218788	3.88E-05	0.00052	4.631753	Up	MET transcriptional regulator MACC1
TRPC6	2.214357	0.000918	0.006155	3.583976	Up	Transient receptor potential cation channel subfamily C member 6
MEOX1	2.213199	0.002772	0.014517	3.191701	Up	Mesenchyme homeobox 1
DOCK4	2.198954	2.50E-06	6.07E-05	5.495826	Up	Dedicator of cytokinesis 4
HTR1B	2.174988	0.001557	0.009248	3.398983	Up	5-Hydroxytryptamine receptor 1B
PRAME	2.156065	1.78E-08	1.31E-06	7.035603	Up	PRAME nuclear receptor transcriptional regulator
THSD7B	2.152357	6.08E-08	3.33E-06	6.651165	Up	Thrombospondin type 1 domain containing 7B
TIMP3	2.150985	2.16E-05	0.000331	4.818136	Up	TIMP metalloproteinase inhibitor 3
SLCO4A1	2.150798	1.71E-06	4.55E-05	5.613756	Up	Solute carrier organic anion transporter family member 4A1
EGFL6	2.146157	2.05E-07	8.48E-06	6.272113	Up	EGF-like domain multiple 6
SERPINB2	2.144838	0.000271	0.002375	3.997832	Up	Serpin family B member 2
CYP4B1	2.142878	0.000609	0.004473	3.725041	Up	Cytochrome P450 family 4 subfamily B member 1
COL7A1	2.141664	3.70E-05	0.000501	4.64651	Up	Collagen type VII alpha 1 chain
FRZB	2.138613	5.90E-09	5.48E-07	7.38458	Up	Frizzled-related protein
PSG3	2.122902	0.000211	0.001955	4.081503	Up	Pregnancy-specific beta-1-glycoprotein 3
VWC2	2.119857	0.000176	0.001702	4.142014	Up	Von Willebrand factor C domain containing 2
CFD	2.113436	0.000635	0.004631	3.710733	Up	Complement factor D
CRACD	2.113112	3.13E-05	0.000443	4.700251	Up	Capping protein inhibiting regulator of actin dynamics
LANCL3	2.108051	2.95E-06	6.92E-05	5.443294	Up	LanC-like family member 3
RAB6C	2.105403	3.52E-10	6.81E-08	8.290269	Up	RAB6C, member RAS oncogene family
NLRP2	2.101201	0.001176	0.007429	3.497735	Up	NLR family pyrin domain containing 2

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
CPE	2.098622	1.22E-07	5.60E-06	6.435386	Up	Carboxypeptidase E
ADAMTS9	2.097533	8.34E-06	0.000158	5.118628	Up	ADAM metallopeptidase with thrombospondin type 1 motif 9
SAMD13	2.094561	4.87E-08	2.86E-06	6.720529	Up	Sterile alpha motif domain containing 13
GPBAR1	2.089881	0.000529	0.004012	3.773206	Up	G protein-coupled bile acid receptor 1
DNAJC6	2.089319	1.38E-05	0.000235	4.960018	Up	DnaJ heat shock protein family (Hsp40) member C6
NPW	2.089175	1.98E-08	1.43E-06	7.001975	Up	Neuropeptide W
XK	2.08758	2.05E-06	5.23E-05	5.55645	Up	X-linked Kx blood group antigen, Kell and VPS13A binding protein
EREG	2.078451	0.0007	0.004995	3.677405	Up	Epiregulin
GABRA2	2.07472	0.000126	0.00132	4.249985	Up	Gamma-aminobutyric acid type A receptor subunit alpha 2
TMCC3	2.069775	2.54E-06	6.15E-05	5.490304	Up	Transmembrane and coiled-coil domain family 3
NDNF	2.06	0.000881	0.005955	3.598115	Up	Neuron-derived neurotrophic factor
DCHS1	2.055197	9.01E-06	0.000169	5.094397	Up	Dachshous cadherin-related 1
CNTN6	2.043284	2.55E-06	6.17E-05	5.488599	Up	Contactin 6
MEGF10	2.040746	6.01E-05	0.000737	4.491082	Up	Multiple EGF-like domains 10
MEI4	2.023279	2.05E-06	5.21E-05	5.557795	Up	Meiotic double-stranded break formation protein 4
SNAP25	1.995428	0.002541	0.013588	3.223379	Up	Synaptosome-associated protein 25
CARD9	1.992794	3.15E-09	3.46E-07	7.584141	Up	Caspase recruitment domain family member 9
ELMO1	1.991602	0.001782	0.010277	3.350853	Up	Engulfment and cell motility 1
SYT1	1.990573	0.000161	0.001593	4.170655	Up	Synaptotagmin 1
NETO1	1.982914	0.007261	0.030523	2.831024	Up	Neuropilin and tolloid-like 1
CD24	1.974298	9.23E-05	0.001036	4.352364	Up	CD24 molecule
SLC26A7	1.966152	0.000434	0.00343	3.84067	Up	Solute carrier family 26 member 7
ZBTB7C	1.9647	1.56E-05	0.000257	4.922019	Up	Zinc finger and BTB domain containing 7C
KIAA1210	1.957336	0.007147	0.030161	2.837133	Up	KIAA1210
LHCGR	1.957307	7.93E-05	0.000917	4.401541	Up	Luteinizing hormone/choriogonadotropin receptor
CLSTN2	1.95386	3.67E-07	1.37E-05	6.092016	Up	Calsyntenin 2
SCD	1.948808	2.97E-10	6.26E-08	8.345503	Up	Stearoyl-CoA desaturase
DTX1	1.943827	0.000125	0.001307	4.254768	Up	Deltex E3 ubiquitin ligase 1
COL24A1	1.939722	0.000211	0.001955	4.081409	Up	Collagen type XXIV alpha 1 chain
SYTL5	1.939593	7.69E-05	0.000894	4.411666	Up	Synaptotagmin-like 5
PDGFB	1.938244	0.002996	0.015394	3.16336	Up	Platelet-derived growth factor subunit B
ZNF608	1.936373	0.000242	0.002174	4.035469	Up	Zinc finger protein 608
RASL11B	1.935406	0.00167	0.009794	3.374029	Up	RAS-like family 11 member B
LSAMP	1.933966	0.00022	0.00202	4.068241	Up	Limbic system-associated membrane protein
WFDC1	1.928958	3.44E-05	0.000476	4.669817	Up	WAP four-disulfide core domain 1
MMP7	1.928028	0.008602	0.034662	2.765233	Up	Matrix metallopeptidase 7
PIK3CG	1.927056	2.85E-05	0.000413	4.730577	Up	Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit gamma
ACSM5	1.925101	0.000743	0.005206	3.65709	Up	acyl-CoA synthetase medium chain family member 5
DGCR6	1.920838	1.73E-06	4.59E-05	5.609382	Up	DiGeorge syndrome critical region gene 6
CYYR1	1.919394	5.50E-05	0.000692	4.519664	Up	Cysteine and tyrosine-rich 1
EFHB	1.919312	0.000592	0.004376	3.734642	Up	EF-hand domain family member B

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
MMRN1	1.917217	0.006887	0.029355	2.851399	Up	Multimerin 1
APOL4	1.916085	0.000867	0.005878	3.603849	Up	Apolipoprotein L4
LDLR	1.915435	8.92E-10	1.37E-07	7.988438	Up	Low-density lipoprotein receptor
SOAT2	1.912507	0.000132	0.001367	4.235818	Up	Sterol O-acyltransferase 2
RGCC	1.911303	2.83E-07	1.11E-05	6.172257	Up	Regulator of cell cycle
CD4	1.910165	0.000151	0.001516	4.191808	Up	CD4 molecule
FOXL2	1.904983	0.003288	0.016488	3.129183	Up	Forkhead box L2
MYZAP	1.89753	0.000551	0.004128	3.759569	Up	Myocardial zonulaadherens protein
AKNAD1	1.896549	1.37E-07	6.12E-06	6.398722	Up	AKNA domain containing 1
AIF1L	1.894951	2.32E-06	5.77E-05	5.518188	Up	Allograft inflammatory factor 1-like
DPF3	1.890886	4.46E-05	0.000582	4.587222	Up	Double PHD fingers 3
LRRC9	1.888336	1.71E-06	4.54E-05	5.614477	Up	Leucine-rich repeat containing 9
GPR183	1.887312	0.000178	0.001718	4.137994	Up	G protein-coupled receptor 183
GPAT3	1.885466	8.31E-05	0.000953	4.386524	Up	Glycerol-3-phosphate acyltransferase 3
CYP2S1	1.876127	0.000157	0.00156	4.179408	Up	Cytochrome P450 family 2 subfamily S member 1
TRPA1	1.874787	2.39E-05	0.000361	4.785926	Up	Transient receptor potential cation channel subfamily A member 1
PTGER4	1.871058	7.25E-08	3.81E-06	6.596382	Up	Prostaglandin E receptor 4
GUCY1A2	1.87003	0.002926	0.015117	3.1719	Up	Guanylatecyclase 1 soluble subunit alpha 2
EPHA5	1.869065	0.00017	0.001659	4.153086	Up	EPH receptor A5
F2RL3	1.867425	2.40E-06	5.91E-05	5.507821	Up	F2R-like thrombin or trypsin receptor 3
DOC2B	1.865626	0.003349	0.01673	3.122408	Up	Double C2 domain beta
KCNMB4	1.863178	1.01E-11	4.66E-09	9.475144	Up	Potassium calcium-activated channel sub-family M regulatory beta subunit 4
C1QL1	1.861516	6.16E-07	2.11E-05	5.930949	Up	Complement C1q-like 1
TOX	1.860898	0.008698	0.034956	2.760886	Up	Thymocyte selection-associated high-mobility group box
ADAMTS5	1.858577	2.85E-06	6.74E-05	5.454439	Up	ADAM metalloproteinase with thrombospondin type 1 motif 5
CST4	1.854793	0.000981	0.006477	3.561072	Up	Cystatin S
DENND2A	1.849173	1.49E-05	0.000249	4.936687	Up	DENN domain containing 2A
RSP03	1.838526	0.003545	0.017522	3.101462	Up	R-spondin 3
WNK2	1.834822	0.004199	0.020035	3.03863	Up	WNK lysine-deficient protein kinase 2
CD163L1	1.832977	0.000434	0.003431	3.840479	Up	CD163 molecule-like 1
MYO7A	1.832212	1.08E-06	3.24E-05	5.755675	Up	Myosin VIIA
LAMB4	1.830346	0.000255	0.002255	4.018242	Up	Laminin subunit beta 4
C1orf115	1.829301	0.001783	0.010279	3.350666	Up	Chromosome 1 open reading frame 115
KRT17	1.826069	0.007054	0.029863	2.842174	Up	Keratin 17
SLCO2A1	1.824825	0.011356	0.042777	2.655589	Up	Solute carrier organic anion transporter family member 2A1
ADGRB3	1.824244	0.011341	0.042738	2.656129	Up	Adhesion G protein-coupled receptor B3
AP1M2	1.81959	8.53E-07	2.67E-05	5.829938	Up	Adaptor-related protein complex 1 subunit mu 2
KRT23	1.815768	0.006914	0.02944	2.849908	Up	Keratin 23
CACNA2D3	1.812351	0.000448	0.003511	3.829647	Up	Calcium voltage-gated channel auxiliary subunit alpha2delta 3
CNNM1	1.81155	2.43E-08	1.69E-06	6.93773	Up	Cyclin and CBS domain divalent metal cation transport mediator 1
MAP2K6	1.806928	2.08E-05	0.00032	4.829723	Up	Mitogen-activated protein kinase kinase 6
ADAMTS1	1.799871	9.71E-05	0.001078	4.33595	Up	ADAM metalloproteinase with thrombospondin type 1 motif 1

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
PDE4B	1.797337	3.32E-05	0.000465	4.681741	Up	Phosphodiesterase 4B
LONRF2	1.796485	0.000406	0.003259	3.862741	Up	LON peptidase N-terminal domain and ring finger 2
PRODH	1.796094	0.000202	0.001894	4.095604	Up	Proline dehydrogenase 1
NES	1.795876	2.81E-08	1.88E-06	6.89227	Up	Nestin
PPM1K-DT	1.790804	0.000253	0.002235	4.021614	Up	PPM1K divergent transcript
SH2D2A	1.784919	0.002087	0.011636	3.294392	Up	SH2 domain containing 2A
FLVCR1-DT	1.76383	2.84E-06	6.72E-05	5.4554	Up	FLVCR1 divergent transcript
IL2RB	1.762693	1.59E-07	6.84E-06	6.351329	Up	Interleukin 2 receptor subunit beta
TH	1.759115	0.000387	0.003149	3.878918	Up	Tyrosine hydroxylase
TMEM119	1.757054	0.000451	0.00353	3.827295	Up	Transmembrane protein 119
IRX6	1.754264	5.32E-05	0.000672	4.530268	Up	Iroquois homeobox 6
PSG9	1.74561	0.000334	0.002807	3.928326	Up	Pregnancy-specific beta-1-glycoprotein 9
GSG1	1.745508	0.000363	0.002994	3.900141	Up	Germ cell associated 1
RIPOR3	1.744933	0.002066	0.011541	3.298045	Up	RIPOR family member 3
PADI2	1.744161	0.008207	0.033553	2.783561	Up	Peptidyl arginine deiminase 2
MUC12	1.743784	1.10E-07	5.12E-06	6.467978	Up	Mucin 12, cell surface associated
CACNA1D	1.742432	9.20E-06	0.000171	5.087788	Up	Calcium voltage-gated channel subunit alpha1 D
LRFN5	1.741517	0.004516	0.021256	3.011478	Up	Leucine-rich repeat and fibronectin type III domain containing 5
MMP8	1.741159	0.000474	0.003672	3.810273	Up	Matrix metalloproteinase 8
KDR	1.735916	0.000728	0.005141	3.663967	Up	Kinase insert domain receptor
SHISA9	1.727647	0.000861	0.005851	3.606041	Up	Shisa family member 9
CNFN	1.725304	8.97E-06	0.000168	5.095699	Up	Cornifelin
TMEM179	1.720603	0.001031	0.006725	3.543782	Up	Transmembrane protein 179
KRT36	1.707213	5.96E-08	3.31E-06	6.657399	Up	Keratin 36
ADRB2	1.707123	0.012557	0.046242	2.615362	Up	Adrenoceptor beta 2
NAP1L3	1.704244	6.36E-06	0.000127	5.203771	Up	Nucleosome assembly protein 1-like 3
ACKR3	1.704237	0.001281	0.00795	3.467756	Up	Atypical chemokine receptor 3
P2RX6	1.701605	6.86E-08	3.64E-06	6.613663	Up	Purinergic receptor P2X 6
RGL3	1.698453	1.70E-07	7.24E-06	6.331144	Up	Ral guanine nucleotide dissociation stimulator-like 3
OLFML2A	1.697944	0.000135	0.001392	4.228681	Up	Olfactomedin-like 2A
CNGA1	1.689869	9.69E-05	0.001077	4.336457	Up	Cyclic nucleotide gated channel subunit alpha 1
ZNF385D	1.68789	0.000248	0.002208	4.027498	Up	Zinc finger protein 385D
CDHR3	1.685844	6.74E-09	6.16E-07	7.342318	Up	Cadherin-related family member 3
GJA3	1.682527	8.54E-05	0.000976	4.377758	Up	Gap junction protein alpha 3
MGAT3	1.682319	2.46E-06	6.00E-05	5.500659	Up	Beta-1,4-mannosyl-glycoprotein 4-beta-N-acetylglucosaminyltransferase
COL21A1	1.680805	0.001948	0.011011	3.319004	Up	Collagen type XXI alpha 1 chain
MSMO1	1.674252	8.37E-10	1.34E-07	8.008722	Up	Methylsterolmonooxygenase 1
ATP6V0D2	1.673953	0.001215	0.007617	3.486263	Up	ATPase H+ transporting V0 subunit d2
TNFRSF9	1.672028	1.68E-05	0.000274	4.897474	Up	TNF receptor superfamily member 9
PTPRC	1.670463	0.00115	0.007314	3.505535	Up	Protein tyrosine phosphatase receptor type C
APOBEC3B	1.669855	0.000729	0.005142	3.663748	Up	Apolipoprotein B mRNA editing enzyme catalytic subunit 3B
SEMA3D	1.665302	0.000472	0.003657	3.812069	Up	Semaphorin 3D
ANK1	1.660532	0.000278	0.002421	3.989632	Up	Ankyrin 1
DIRAS2	1.659017	0.012554	0.04624	2.615447	Up	DIRAS family GTPase 2

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
B4GALNT2	1.658134	0.000614	0.004498	3.722662	Up	Beta-1,4-N-acetyl-galactosaminyltransferase 2
CCDC144A	1.657617	0.006513	0.02811	2.872803	Up	Coiled-coil domain containing 144A
ZNF474	1.655783	1.24E-05	0.000215	4.993492	Up	Zinc finger protein 474
MARK1	1.653507	0.000873	0.005913	3.601391	Up	Microtubule affinity regulating kinase 1
PITX1	1.652387	0.003109	0.015818	3.149696	Up	Paired-like homeodomain 1
EYA1	1.650534	0.000544	0.004096	3.763743	Up	EYA transcriptional coactivator and phosphatase 1
TXK	1.639055	1.06E-06	3.18E-05	5.762193	Up	TXK tyrosine kinase
KL	1.638649	0.000243	0.002178	4.034235	Up	Klotho
PCARE	1.635391	0.000312	0.002658	3.951005	Up	Photoreceptor cilium actin regulator
PRECSIT	1.633825	0.002252	0.012361	3.267025	Up	p53-regulated carcinoma-associated Stat3 activating long intergenic non-protein coding transcript
ST8SIA1	1.633014	0.000571	0.004251	3.747217	Up	ST8 alpha-N-acetyl-neuraminide alpha-2,8-sialyltransferase 1
PLAU	1.631288	5.97E-07	2.07E-05	5.940868	Up	Plasminogen activator, urokinase
EPHA7	1.62913	0.005243	0.023861	2.955306	Up	EPH receptor A7
ZNF365	1.624049	0.000217	0.002005	4.071338	Up	Zinc finger protein 365
PIK3R3	1.62296	0.001303	0.008059	3.461725	Up	Phosphoinositide-3-kinase regulatory subunit 3
GALNT14	1.618549	0.002579	0.013742	3.21792	Up	Polypeptide N-acetylgalactosaminyltransferase 14
ADGRL4	1.618549	0.000762	0.005307	3.64825	Up	Adhesion G protein-coupled receptor L4
GJB2	1.618325	0.007075	0.029917	2.841034	Up	Gap junction protein beta 2
SYN3	1.618277	0.001316	0.00812	3.458311	Up	Synapsin III
CCDC141	1.614116	0.000716	0.005087	3.669619	Up	Coiled-coil domain containing 141
HECW2	1.612933	0.000636	0.004637	3.710178	Up	HECT, C2 and WW domain containing E3 ubiquitin protein ligase 2
SOX8	1.609757	8.04E-05	0.000927	4.39731	Up	SRY-box transcription factor 8
CCND2	1.609603	5.94E-07	2.06E-05	5.942393	Up	Cyclin D2
ATP1B2	1.609317	3.49E-05	0.00048	4.665774	Up	ATPase Na ⁺ /K ⁺ transporting subunit beta 2
RTN4R	1.60759	2.91E-09	3.25E-07	7.608653	Up	Reticulon 4 receptor
HMGCS1	1.606762	2.67E-09	3.04E-07	7.636955	Up	3-Hydroxy-3-methylglutaryl-CoA synthase 1
PTGS1	1.601971	2.41E-05	0.000363	4.782843	Up	Prostaglandin-endoperoxide synthase 1
CRABP2	1.599948	3.53E-07	1.33E-05	6.10377	Up	Cellular retinoic acid binding protein 2
KRTAP2-3	1.597381	0.000586	0.004334	3.738465	Up	Keratin-associated protein 2-3
PCOTH	1.596003	0.000192	0.001826	4.11299	Up	Prostate and testis expressed opposite C1QTNF9B and MIPEP
NIIPB2	1.593271	0.003013	0.015455	3.161236	Up	Nuclear pore complex interacting protein family member B2
HECW1	1.58806	0.00442	0.020879	3.019552	Up	HECT, C2 and WW domain containing E3 ubiquitin protein ligase 1
PTPRD	1.587373	2.59E-12	1.59E-09	9.943946	Up	Protein tyrosine phosphatase receptor type D
ZNF114	1.585505	1.17E-06	3.42E-05	5.732094	Up	Zinc finger protein 114
MCTP1	1.58428	0.000679	0.004886	3.688218	Up	Multiple C2 and transmembrane domain containing 1
PCDH9	1.582512	9.60E-07	2.94E-05	5.793167	Up	Protocadherin 9
MASP1	1.58163	0.000666	0.004822	3.694404	Up	MBL-associated serine protease 1
NALF1	1.578515	0.005281	0.024012	2.95261	Up	NALCN channel auxiliary factor 1
TOGARAM2	1.576442	2.67E-07	1.05E-05	6.191049	Up	TOG array regulator of axonemal microtubules 2

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
ST6GALNAC2	1.576212	9.42E-05	0.001053	4.345596	Up	ST6 N-acetylgalactosaminide alpha-2,6-sialyltransferase 2
LNCOG	1.57379	8.65E-06	0.000163	5.106912	Up	lncRNA osteogenesis associated
QPRT	1.572637	1.33E-07	6.02E-06	6.408497	Up	Quinolate phosphoribosyltransferase
INSIG1	1.570033	3.38E-07	1.29E-05	6.117312	Up	Insulin-induced gene 1
TRIM9	1.56912	0.010349	0.0399	2.692488	Up	Tripartite motif containing 9
BMP8A	1.568731	0.000269	0.002359	4.000743	Up	Bone morphogenetic protein 8a
DYNLT5	1.560446	1.07E-07	5.03E-06	6.475716	Up	Dynein light chain Tctex-type family member 5
SORCS1	1.558679	0.008327	0.033886	2.777893	Up	Sortilin-related VPS10 domain containing receptor 1
IMPA2	1.557806	1.84E-05	0.000292	4.869246	Up	Inositol monophosphatase 2
MACC1-DT	1.554741	0.000276	0.002403	3.992735	Up	MACC1 divergent transcript
CHRM2	1.550458	0.00899	0.035819	2.747955	Up	Cholinergic receptor muscarinic 2
HKDC1	1.549584	6.00E-05	0.000736	4.491685	Up	Hexokinase domain containing 1
EPCAM	1.547502	0.002885	0.014961	3.177114	Up	Epithelial cell adhesion molecule
WNT9A	1.535805	0.00015	0.001506	4.194741	Up	Wnt family member 9A
UCA1	1.535128	0.012638	0.046442	2.612777	Up	Urothelial cancer associated 1
PPM1H	1.532532	0.010359	0.03992	2.692121	Up	Protein phosphatase, Mg2+/Mn2+ dependent 1H
PRXL2A	1.531991	0.006859	0.02927	2.852957	Up	Peroxioredoxin-like 2A
KRT37	1.531948	2.39E-05	0.000361	4.786092	Up	Keratin 37
FSIP2	1.531	0.0005	0.003829	3.792398	Up	Fibrous sheath interacting protein 2
TRIM55	1.530497	0.000422	0.003361	3.849581	Up	Tripartite motif containing 55
GRM4	1.529606	7.85E-08	4.01E-06	6.57182	Up	Glutamate metabotropic receptor 4
IRX2	1.527283	0.002829	0.014726	3.184301	Up	Iroquois homeobox 2
SEMA3B	1.523654	1.97E-08	1.42E-06	7.004789	Up	Semaphorin 3B
TSPAN12	1.521905	3.93E-08	2.44E-06	6.787408	Up	Tetraspanin 12
SNTB1	1.519848	7.66E-13	7.58E-10	10.37296	Up	Syntrophin beta 1
BATF2	1.517422	9.31E-05	0.001041	4.349725	Up	Basic leucine zipper ATF-like transcription factor 2
SIGLEC1	1.51605	2.89E-05	0.000416	4.725468	Up	Sialic acid binding Ig-like lectin 1
GCOM1	1.509491	0.002311	0.012631	3.257672	Up	GCOM1, MYZAP-POLR2M combined locus
SLC37A2	1.509308	2.96E-06	6.92E-05	5.442942	Up	Solute carrier family 37 member 2
CLDN23	1.509024	0.003664	0.017942	3.089276	Up	Claudin 23
KRT34	1.506734	0.000158	0.001571	4.176501	Up	Keratin 34
MBOAT1	1.503727	3.60E-06	8.08E-05	5.381682	Up	Membrane-bound O-acyltransferase domain containing 1
UCN2	1.501515	0.001732	0.010056	3.361125	Up	Urocortin 2
CRTAC1	1.501044	0.000227	0.002071	4.056501	Up	Cartilage acidic protein 1
ADGRG6	1.500022	0.000505	0.003859	3.789317	Up	Adhesion G protein-coupled receptor G6
TNFRSF18	1.499245	0.000345	0.00288	3.917721	Up	TNF receptor superfamily member 18
ACAT2	1.498762	5.90E-09	5.48E-07	7.384612	Up	Acetyl-CoA acetyltransferase 2
CYP2J2	1.496024	2.63E-05	0.000389	4.755929	Up	Cytochrome P450 family 2 subfamily J member 2
CLEC14A	1.495564	0.00037	0.003041	3.893644	Up	C-type lectin domain containing 14A
CYP1A1	1.491253	0.00975	0.038182	2.716075	Up	Cytochrome P450 family 1 subfamily A member 1
NEFM	1.490799	0.001644	0.009686	3.379503	Up	Neurofilament medium chain
RUBCNL	1.489824	3.29E-05	0.000462	4.684258	Up	Rubicon-like autophagy enhancer
CDO1	1.486353	7.93E-05	0.000917	4.401608	Up	Cysteine dioxygenase type 1

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
H2AC20	1.485079	0.000107	0.00116	4.304163	Up	H2A clustered histone 20
ASTN2	1.483569	2.17E-09	2.56E-07	7.70268	Up	Astrotactin 2
NTN1	1.479365	0.011399	0.042871	2.654087	Up	Netrin 1
P3H2	1.479255	3.81E-05	0.000512	4.637609	Up	Prolyl 3-hydroxylase 2
FBXW10	1.477472	2.41E-05	0.000362	4.784075	Up	F-box and WD repeat domain containing 10
GRIP2	1.474979	7.91E-07	2.52E-05	5.853548	Up	Glutamate receptor interacting protein 2
ATP1A2	1.473718	0.010376	0.03997	2.691488	Up	ATPase Na ⁺ /K ⁺ transporting subunit alpha 2
STAC2	1.469529	0.004907	0.022687	2.980292	Up	SH3 and cysteine-rich domain 2
ARPP21	1.46692	0.001824	0.010466	3.342611	Up	cAMP-regulated phosphoprotein 21
PDGFD	1.46332	4.95E-05	0.000633	4.553809	Up	Platelet-derived growth factor D
AFF3	1.461558	0.000108	0.001167	4.301496	Up	ALF transcription elongation factor 3
ADGRG1	1.453238	0.000199	0.001869	4.101306	Up	Adhesion G protein-coupled receptor G1
HS3ST1	1.446114	0.003511	0.017392	3.105029	Up	Heparan sulfate-glucosamine 3-sulfotransferase 1
PNPLA3	1.441511	1.37E-06	3.89E-05	5.681647	Up	Patatin-like phospholipase domain containing 3
TNFRSF19	1.440266	0.002116	0.011764	3.289401	Up	TNF receptor superfamily member 19
CCDC188	1.433143	1.51E-05	0.000251	4.932387	Up	Coiled-coil domain containing 188
CLDN3	1.432576	0.00114	0.00727	3.508451	Up	Claudin 3
MSTN	1.43247	0.000702	0.005005	3.676601	Up	Myostatin
TMC1	1.431978	0.008233	0.033618	2.782322	Up	Transmembrane channel-like 1
CCDC85A	1.427842	0.003586	0.017671	3.097223	Up	Coiled-coil domain containing 85A
VPREB3	1.425686	1.32E-05	0.000226	4.973789	Up	V-set pre-B cell surrogate light chain 3
ERMP1	1.425535	5.89E-08	3.30E-06	6.661374	Up	Endoplasmic reticulum metalloproteinase 1
LEPR	1.418868	0.000184	0.001768	4.126541	Up	Leptin receptor
FADS1	1.418126	9.06E-10	1.37E-07	7.983374	Up	Fatty acid desaturase 1
SDK2	1.416107	0.002872	0.014912	3.178806	Up	Sidekick cell adhesion molecule 2
DMRTA2	1.413035	0.000639	0.004655	3.708625	Up	DMRT-like family A2
CEROX1	1.410648	0.00155	0.009217	3.400537	Up	Cytoplasmic endogenous regulator of oxidative phosphorylation 1
RTKN2	1.409168	0.00655	0.028242	2.870656	Up	Rhotekin 2
CD248	1.408077	4.55E-08	2.72E-06	6.741817	Up	CD248 molecule
IL15	1.40691	0.004192	0.020011	3.039324	Up	Interleukin 15
STK32B	1.406641	0.000785	0.005428	3.63813	Up	Serine/threonine kinase 32B
GSTT2B	1.406073	2.50E-05	0.000374	4.771519	Up	Glutathione S-transferase theta 2B
CPED1	1.403905	1.89E-07	7.85E-06	6.298593	Up	Cadherin-like and PC-esterase domain containing 1
KREMEN1	1.403521	7.96E-07	2.53E-05	5.851322	Up	Kringle containing transmembrane protein 1
CACNA1H	1.402919	0.005026	0.023107	2.971315	Up	Calcium voltage-gated channel subunit alpha1 H
BMP2	1.402331	0.002908	0.015056	3.174247	Up	Bone morphogenetic protein 2
PCDHB15	1.401583	1.31E-07	5.97E-06	6.412115	Up	Protocadherin beta 15
GIMAP2	1.400683	0.000251	0.002227	4.023704	Up	GTPase, IMAP family member 2
GLUD2	1.400362	1.17E-05	0.000206	5.011695	Up	Glutamate dehydrogenase 2
FAM169A	1.399373	4.26E-05	0.000563	4.601522	Up	Family with sequence similarity 169 member A
PCOLCE	1.398122	1.53E-08	1.16E-06	7.084186	Up	Procollagen C-endopeptidase enhancer
LPAR4	1.391463	0.00028	0.002431	3.987885	Up	Lysophosphatidic acid receptor 4
LAMA5	1.390713	1.56E-06	4.30E-05	5.6425	Up	Laminin subunit alpha 5
SYT7	1.389911	0.000605	0.004453	3.727472	Up	Synaptotagmin 7

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
SLC2A8	1.387939	1.41E-05	0.000238	4.953929	Up	Solute carrier family 2 member 8
NTNG1	1.387735	0.005839	0.025811	2.91451	Up	Netrin G1
NELL2	1.387296	0.00387	0.018726	3.069057	Up	Neural EGFL-like 2
TCIM	1.386703	4.85E-05	0.000624	4.560422	Up	Transcriptional and immune response regulator
OGFRL1	1.385353	2.05E-05	0.000317	4.834026	Up	Opioid growth factor receptor-like 1
CRYBG1	1.382787	0.00034	0.002847	3.922792	Up	Crystallin beta-gamma domain containing 1
TAS2R1	1.382739	0.00365	0.017882	3.090739	Up	Taste 2 receptor member 1
ZBTB46	1.380412	6.99E-07	2.31E-05	5.891925	Up	Zinc finger and BTB domain containing 46
PAK3	1.377753	0.003696	0.018063	3.086095	Up	p21 (RAC1) activated kinase 3
KRT19	1.377283	2.40E-05	0.000361	4.785049	Up	Keratin 19
FABP3	1.376442	2.80E-05	0.000408	4.735269	Up	Fatty acid binding protein 3
IDI1	1.37204	3.14E-10	6.40E-08	8.326735	Up	Isopentenyl-diphosphate delta isomerase 1
ROS1	1.371129	0.002688	0.01419	3.202863	Up	ROS proto-oncogene 1, receptor tyrosine kinase
ATRNL1	1.370951	1.82E-05	0.00029	4.871979	Up	Attractin-like 1
MME	1.370733	0.002971	0.0153	3.16643	Up	Membrane metalloendopeptidase
APOE	1.369649	0.006638	0.028528	2.865549	Up	Apolipoprotein E
SFMBT2	1.369082	0.001173	0.007418	3.498669	Up	Scm-like with four mbt domains 2
CUBN	1.367232	5.16E-05	0.000656	4.540303	Up	Cubilin
ACSL5	1.366293	2.81E-08	1.88E-06	6.892859	Up	Acyl-CoA synthetase long chain family member 5
SIPA1L2	1.365666	0.001365	0.008339	3.445393	Up	Signal-induced proliferation associated 1-like 2
PTCH1	1.361587	1.34E-09	1.80E-07	7.857233	Up	Patched 1
OSR2	1.360422	0.005582	0.02499	2.931623	Up	Odd-skipped-related transcription factor 2
ITPKA	1.359673	1.02E-06	3.08E-05	5.775409	Up	Inositol-trisphosphate 3-kinase A
RAB7B	1.359256	1.56E-07	6.74E-06	6.358232	Up	RAB7B, member RAS oncogene family
RBM20	1.353082	0.004207	0.02006	3.037929	Up	RNA binding motif protein 20
CASP1	1.343594	1.22E-05	0.000213	4.999578	Up	Caspase 1
PDE9A	1.341431	0.002844	0.014796	3.182322	Up	Phosphodiesterase 9A
PTPRN2	1.340227	1.74E-05	0.000281	4.887206	Up	Protein tyrosine phosphatase receptor type N2
MGLL	1.33916	9.37E-11	2.50E-08	8.724848	Up	Monoglyceride lipase
EFNB2	1.336805	0.000908	0.006098	3.587838	Up	Ephrin B2
GYPE	1.335653	0.000109	0.001173	4.298897	Up	Glycophorin E (MNS blood group)
STAMBPL1	1.335123	0.000565	0.00422	3.750655	Up	STAM binding protein-like 1
HLA-C	1.331822	9.95E-10	1.47E-07	7.953065	Up	Major histocompatibility complex, class I, C
NID1	1.330925	7.14E-07	2.35E-05	5.885025	Up	Nidogen 1
ABCG2	1.328504	0.000181	0.001743	4.132914	Up	ATP binding cassette subfamily G member 2 (Junior blood group)
XCR1	1.327928	0.003631	0.017807	3.092625	Up	X-C motif chemokine receptor 1
HMGCR	1.327709	6.18E-10	1.05E-07	8.107146	Up	3-Hydroxy-3-methylglutaryl-CoA reductase
ACE	1.32685	2.56E-06	6.17E-05	5.487931	Up	Angiotensin I converting enzyme
GRASLND	1.326126	0.000509	0.003885	3.78638	Up	Glycosaminoglycan regulatory-associated long non-coding RNA
PPFIA2	1.323272	0.001732	0.010057	3.361016	Up	PTPRF interacting protein alpha 2
ENTREP1	1.322912	0.004413	0.020852	3.02012	Up	Endosomal transmembrane epsin interactor 1
RNF17	1.32049	0.01074	0.041023	2.677805	Up	Ring finger protein 17
CORT	1.318854	0.000189	0.0018	4.118604	Up	Cortistatin

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
BFSP1	1.3175	3.77E-12	2.19E-09	9.813358	Up	Beaded filament structural protein 1
PGR	1.317315	0.000265	0.002329	4.005569	Up	Progesterone receptor
RENBP	1.316502	0.000375	0.003067	3.889321	Up	Renin binding protein
WLS	1.313999	7.40E-07	2.40E-05	5.8742	Up	Wnt ligand secretion mediator
RIMS1	1.313761	0.00186	0.010619	3.335646	Up	Regulating synaptic membrane exocytosis 1
KLK3	1.31258	0.006761	0.028959	2.858486	Up	Kallikrein-related peptidase 3
MCOLN2	1.311284	0.004696	0.021935	2.996798	Up	Mucolipin TRP cation channel 2
ST3GAL5	1.310723	1.91E-06	4.98E-05	5.578951	Up	ST3 beta-galactoside alpha-2,3-sialyltransferase 5
STMN2	1.310233	0.008348	0.033934	2.776924	Up	Stathmin 2
ARSL	1.307945	0.000358	0.00296	3.905428	Up	Arylsulfatase L
GREM2	1.306505	0.004675	0.021874	2.99854	Up	Gremlin 2, DAN family BMP antagonist
C1QTNF7	1.304475	0.001677	0.009816	3.37246	Up	C1q and TNF related 7
PTGIS	-5.23674	3.67E-08	2.30E-06	-6.80923	Down	Prostaglandin I2 synthase
SFRP4	-5.07827	6.79E-11	1.84E-08	-8.83175	Down	Secreted frizzled-related protein 4
MFAP5	-4.50412	0.00014	0.001432	-4.21667	Down	Microfibril-associated protein 5
BMP6	-4.46815	1.78E-10	4.45E-08	-8.51213	Down	Bone morphogenetic protein 6
CDH6	-4.39935	1.13E-09	1.60E-07	-7.91281	Down	Cadherin 6
TSPAN2	-4.37263	3.01E-10	6.26E-08	-8.34126	Down	Tetraspanin 2
PDE1C	-4.30317	1.98E-06	5.09E-05	-5.56744	Down	Phosphodiesterase 1C
GPC6	-4.22114	2.48E-10	5.61E-08	-8.40467	Down	Glypican 6
PIEZO2	-4.10461	4.39E-08	2.65E-06	-6.75297	Down	Piezo-type mechanosensitive ion channel component 2
PKP2	-4.02356	3.01E-10	6.26E-08	-8.34141	Down	Plakophilin 2
RARRES1	-3.93563	5.18E-17	5.87E-13	-14.1022	Down	Retinoic acid receptor responder 1
MDFI	-3.75278	2.44E-10	5.58E-08	-8.40984	Down	MyoD family inhibitor
NALCN	-3.73823	2.85E-09	3.22E-07	-7.61577	Down	Sodium leak channel, non-selective
EMB	-3.71037	2.54E-07	1.00E-05	-6.20599	Down	Embigin
PDLIM3	-3.69103	0.00088	0.005951	-3.59883	Down	PDZ and LIM domain 3
MGAM	-3.5935	4.99E-11	1.53E-08	-8.93413	Down	Maltase-glucoamylase
COL11A1	-3.56036	2.26E-06	5.64E-05	-5.52742	Down	Collagen type XI alpha 1 chain
RERG	-3.556	0.000342	0.002863	-3.92067	Down	RAS-like estrogen-regulated growth inhibitor
GDNF	-3.5355	6.42E-11	1.80E-08	-8.85003	Down	Glial cell-derived neurotrophic factor
CHRD2	-3.51084	2.03E-07	8.40E-06	-6.27594	Down	Chordin-like 2
FOXL1	-3.49556	2.80E-08	1.88E-06	-6.89356	Down	Forkhead box L1
PLPP4	-3.43759	7.45E-18	1.69E-13	-14.9443	Down	Phospholipid phosphatase 4
C5orf46	-3.32884	0.000143	0.001451	-4.2099	Down	Chromosome 5 open reading frame 46
ANXA8	-3.32023	8.43E-12	4.16E-09	-9.53561	Down	Annexin A8
FIBIN	-3.29706	1.84E-09	2.27E-07	-7.75512	Down	Fin bud initiation factor homolog
LMO3	-3.28827	1.01E-05	0.000184	-5.05759	Down	LIM domain only 3
CABLES1	-3.2806	1.65E-10	4.20E-08	-8.53847	Down	Cdk5 and Abl enzyme substrate 1
ITGBL1	-3.26694	1.22E-07	5.60E-06	-6.43497	Down	Integrin subunit beta-like 1
NFASC	-3.25827	4.61E-08	2.74E-06	-6.7377	Down	Neurofascin
FGFR2	-3.25145	6.44E-07	2.19E-05	-5.91735	Down	Fibroblast growth factor receptor 2
ANXA8L1	-3.24754	4.18E-11	1.37E-08	-8.99366	Down	Annexin A8-like 1
KCNJ12	-3.23099	1.52E-09	1.97E-07	-7.81704	Down	Potassium inwardly rectifying channel subfamily J member 12
NPY4R	-3.17277	6.04E-07	2.08E-05	-5.93721	Down	Neuropeptide Y receptor Y4
GALNT12	-3.16958	2.96E-10	6.26E-08	-8.34597	Down	Polypeptide N-acetylgalactosaminyltransferase 12

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
TMEM215	-3.15136	0.000603	0.004439	-3.72871	Down	Transmembrane protein 215
IL33	-3.14699	3.03E-09	3.35E-07	-7.59651	Down	Interleukin 33
GAL3ST3	-3.07934	1.32E-11	5.77E-09	-9.38217	Down	Galactose-3-O-sulfotransferase 3
PLCB4	-3.07703	6.02E-10	1.04E-07	-8.11536	Down	Phospholipase C beta 4
NGF	-3.05568	2.83E-07	1.11E-05	-6.17229	Down	Nerve growth factor
PAPPA2	-3.02516	6.76E-06	0.000134	-5.18461	Down	Pappalysin 2
CD200	-3.00614	7.69E-09	6.92E-07	-7.30053	Down	CD200 molecule
BST2	-2.99884	8.75E-06	0.000165	-5.10331	Down	Bone marrow stromal cell antigen 2
TENM2	-2.99726	7.57E-05	0.000885	-4.41665	Down	Teneurin transmembrane protein 2
ARHGEF4	-2.97645	3.76E-09	3.95E-07	-7.52717	Down	Rho guanine nucleotide exchange factor 4
SEL1L3	-2.97399	3.91E-06	8.64E-05	-5.35617	Down	SEL1L family member 3
TBX18	-2.93338	5.69E-06	0.000116	-5.23831	Down	T-box transcription factor 18
MN1	-2.92877	4.16E-05	0.000552	-4.6095	Down	MN1 proto-oncogene, transcriptional regulator
FAM110C	-2.92689	0.00014	0.00143	-4.21713	Down	Family with sequence similarity 110 member C
HAPLN1	-2.92354	3.12E-06	7.21E-05	-5.42614	Down	Hyaluronan and proteoglycan link protein 1
KIAA1549L	-2.91526	7.15E-07	2.35E-05	-5.88491	Down	KIAA1549-like
GDF6	-2.89617	0.003217	0.016242	-3.13726	Down	Growth differentiation factor 6
NPY4R2	-2.88311	5.68E-07	1.98E-05	-5.95614	Down	Neuropeptide Y receptor Y4-2
FOXC2	-2.85881	5.18E-06	0.000108	-5.26771	Down	Forkhead box C2
ERG	-2.84212	0.000503	0.003853	-3.79014	Down	ETS transcription factor ERG
COMP	-2.84085	0.001248	0.00778	-3.47699	Down	Cartilage oligomeric matrix protein
CECR2	-2.83971	6.76E-08	3.61E-06	-6.61824	Down	CECR2 histone acetyl-lysine reader
FST	-2.83619	3.30E-10	6.56E-08	-8.31108	Down	Follistatin
GOLGA8M	-2.82467	7.78E-09	6.97E-07	-7.29701	Down	Golgin A8 family member M
FMO2	-2.82238	2.18E-07	8.84E-06	-6.25419	Down	Flavin containing dimethylaniline monooxygenase 2
AK4	-2.81954	3.84E-05	0.000516	-4.63504	Down	Adenylate kinase 4
SORBS2	-2.8058	0.000102	0.001116	-4.32079	Down	Sorbin and SH3 domain containing 2
CACNG7	-2.79309	1.93E-06	5.01E-05	-5.5752	Down	Calcium voltage-gated channel auxiliary subunit gamma 7
ZMAT1	-2.78465	3.16E-10	6.40E-08	-8.32498	Down	Zinc finger matrin-type 1
BEND7	-2.77362	2.41E-15	1.36E-11	-12.5293	Down	BEN domain containing 7
TPD52L1	-2.76596	5.49E-06	0.000113	-5.2496	Down	TPD52-like 1
TCF23	-2.75207	0.007372	0.030866	-2.82513	Down	Transcription factor 23
FGF14	-2.74619	1.20E-07	5.53E-06	-6.4404	Down	Fibroblast growth factor 14
ITGA7	-2.73939	1.02E-08	8.46E-07	-7.2129	Down	Integrin subunit alpha 7
PLN	-2.7368	0.000946	0.0063	-3.57356	Down	Phospholamban
EFEMP1	-2.73301	2.81E-06	6.66E-05	-5.45894	Down	EGF containing fibulin extracellular matrix protein 1
GATA6	-2.71505	9.68E-09	8.19E-07	-7.22789	Down	GATA binding protein 6
CXADR	-2.67583	1.39E-06	3.93E-05	-5.67788	Down	CXADR Ig-like cell adhesion molecule
BHLHE41	-2.67205	5.13E-08	2.96E-06	-6.70473	Down	Basic helix-loop-helix family member e41
ACAN	-2.65447	0.00106	0.006881	-3.53408	Down	Aggrecan
MYH1	-2.65008	0.003197	0.016158	-3.1395	Down	Myosin heavy chain 1
LRRN2	-2.6404	0.000231	0.002093	-4.05193	Down	Leucine-rich repeat neuronal 2
CSGALNACT1	-2.63549	6.22E-08	3.40E-06	-6.6444	Down	Chondroitin sulfate N-acetylgalactosaminyltransferase 1
TCF21	-2.63196	0.004002	0.019265	-3.05653	Down	Transcription factor 21

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
CACNG6	-2.62787	2.46E-06	6.00E-05	-5.5005	Down	Calcium voltage-gated channel auxiliary subunit gamma 6
PTGS2	-2.61561	0.000993	0.006533	-3.5567	Down	Prostaglandin-endoperoxide synthase 2
MTSS1	-2.5969	3.28E-08	2.12E-06	-6.84404	Down	MTSS I-BAR domain containing 1
LRRC32	-2.5596	9.10E-12	4.29E-09	-9.50977	Down	Leucine-rich repeat containing 32
APCDD1L	-2.55771	1.78E-06	4.70E-05	-5.6015	Down	APC downregulated 1-like
GDF10	-2.52375	8.52E-06	0.000161	-5.11174	Down	Growth differentiation factor 10
APCDD1L-DT	-2.52144	0.000121	0.00128	-4.26316	Down	APCDD1L divergent transcript
CPNE5	-2.50055	3.14E-05	0.000445	-4.69907	Down	Copine 5
RASSF9	-2.48511	0.000533	0.004036	-3.77084	Down	Ras association domain family member 9
TNFAIP8L3	-2.46545	4.02E-05	0.000537	-4.61998	Down	TNF alpha-induced protein 8-like 3
KCNN3	-2.46136	4.68E-07	1.69E-05	-6.01633	Down	Potassium calcium-activated channel sub-family N member 3
DHRS3	-2.45497	1.30E-08	1.02E-06	-7.13549	Down	Dehydrogenase/reductase 3
HMCN1	-2.44784	3.66E-05	0.000497	-4.64995	Down	Hemicentin 1
DACT1	-2.43049	9.85E-09	8.27E-07	-7.22242	Down	Disheveled binding antagonist of beta catenin 1
SPINT2	-2.42485	6.05E-08	3.33E-06	-6.65307	Down	Serine peptidase inhibitor, Kunitz type 2
NKX3-1	-2.41903	4.50E-10	8.23E-08	-8.20991	Down	NK3 homeobox 1
CMKLR2	-2.41878	2.94E-05	0.000421	-4.72009	Down	Chemerin chemokine-like receptor 2
CHN2	-2.41303	0.00019	0.001815	-4.11517	Down	Chimerin 2
HOXC8	-2.41049	0.001204	0.007561	-3.48945	Down	Homeobox C8
DGAT2	-2.3977	1.91E-06	4.98E-05	-5.57954	Down	Diacylglycerol O-acyltransferase 2
PITX2	-2.38971	0.000544	0.004096	-3.76375	Down	Paired-like homeodomain 2
MEGF6	-2.38799	1.44E-06	4.03E-05	-5.66645	Down	Multiple EGF-like domains 6
TCEAL2	-2.38755	3.09E-06	7.16E-05	-5.42892	Down	Transcription elongation factor A-like 2
SLC2A5	-2.38396	4.28E-05	0.000564	-4.60045	Down	Solute carrier family 2 member 5
AKR1C3	-2.37252	8.03E-05	0.000927	-4.39752	Down	Aldo-ketoreductase family 1 member C3
TNFSF10	-2.36958	0.000237	0.00214	-4.04219	Down	TNF superfamily member 10
SCGB3A2	-2.35953	4.24E-05	0.000561	-4.60308	Down	Secretoglobulin family 3A member 2
BDNF	-2.35866	1.27E-09	1.73E-07	-7.87543	Down	Brain-derived neurotrophic factor
KRT7	-2.35066	7.32E-06	0.000142	-5.15962	Down	Keratin 7
ADAMTSL1	-2.34915	0.000201	0.001888	-4.09725	Down	ADAMTS-like 1
RGS4	-2.34753	0.001356	0.008304	-3.44775	Down	Regulator of G protein signaling 4
KCNE4	-2.34479	1.96E-06	5.06E-05	-5.57154	Down	Potassium voltage-gated channel subfamily E regulatory subunit 4
SMIM43	-2.34033	9.83E-07	3.00E-05	-5.78594	Down	Small integral membrane protein 43
SBSPPON	-2.33756	1.88E-05	0.000297	-4.86278	Down	Somatomedin B and thrombospondin type 1 domain containing
RAPGEF5	-2.33731	0.000947	0.006305	-3.57308	Down	Rap guanine nucleotide exchange factor 5
NDRG2	-2.33365	3.13E-10	6.40E-08	-8.3278	Down	NDRG family member 2
IGFN1	-2.33352	0.00021	0.001948	-4.0831	Down	Immunoglobulin-like and fibronectin type III domain containing 1
THEM5	-2.31898	2.19E-06	5.50E-05	-5.53726	Down	Thioesterase superfamily member 5
SCG2	-2.31212	0.000444	0.003496	-3.83241	Down	Secretogranin II
MT1F	-2.29146	5.10E-10	9.17E-08	-8.16938	Down	Metallothionein 1F
EPHA3	-2.27945	0.010466	0.040236	-2.68805	Down	EPH receptor A3
PLA2G5	-2.27384	0.001836	0.010509	-3.34033	Down	Phospholipase A2 group V
OLAH	-2.27268	1.47E-05	0.000246	-4.94049	Down	Oleoyl-ACP hydrolase
EYA2	-2.27257	8.10E-05	0.000934	-4.39458	Down	EYA transcriptional coactivator and phosphatase 2

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
GMNC	-2.26975	6.58E-06	0.000131	-5.19309	Down	Geminin coiled-coil domain containing
PCDHA12	-2.26677	1.73E-09	2.19E-07	-7.77547	Down	Protocadherin alpha 12
KCNJ2	-2.26148	1.99E-05	0.00031	-4.84451	Down	Potassium inwardly rectifying channel subfamily J member 2
EVA1A	-2.2484	2.10E-06	5.32E-05	-5.54978	Down	Eva-1 homolog A, regulator of programmed cell death
INHBA	-2.24796	3.67E-09	3.89E-07	-7.53531	Down	Inhibin subunit beta A
ALPL	-2.2474	0.000123	0.001295	-4.25866	Down	Alkaline phosphatase, biomineralization associated
C10orf90	-2.24435	8.67E-08	4.35E-06	-6.5408	Down	Chromosome 10 open reading frame 90
LXN	-2.2373	1.21E-16	9.11E-13	-13.7459	Down	Latexin
MECOM	-2.23059	3.12E-06	7.21E-05	-5.42619	Down	MDS1 and EVI1 complex locus
SGK1	-2.22874	7.04E-14	1.14E-10	-11.2391	Down	Serum/glucocorticoid-regulated kinase 1
CSTA	-2.22764	1.34E-05	0.000228	-4.97019	Down	Cystatin A
ATP10A	-2.20744	4.70E-07	1.69E-05	-6.01505	Down	ATPase phospholipid transporting 10A (putative)
RAP1GAP	-2.20507	4.84E-09	4.73E-07	-7.44742	Down	RAP1 GTPase activating protein
ULBP1	-2.20094	1.57E-06	4.30E-05	-5.64097	Down	UL16 binding protein 1
HOXC9	-2.19532	0.001264	0.007861	-3.47228	Down	Homeobox C9
PLEKHB1	-2.17758	2.38E-07	9.51E-06	-6.22619	Down	Pleckstrin homology domain containing B1
FHIP1A	-2.17607	3.08E-05	0.000437	-4.70586	Down	FHF complex subunit HOOK interacting protein 1A
VEPH1	-2.17448	7.23E-05	0.000852	-4.43137	Down	Ventricular zone expressed PH domain containing 1
PRR15	-2.17303	0.000208	0.001932	-4.08688	Down	Proline-rich 15
SLC2A12	-2.16601	2.10E-08	1.49E-06	-6.98398	Down	Solute carrier family 2 member 12
RGS7BP	-2.16065	0.00015	0.001506	-4.19413	Down	Regulator of G protein signaling 7 binding protein
COL4A4	-2.15631	0.000804	0.005533	-3.63009	Down	Collagen type IV alpha 4 chain
TMEM252	-2.1552	0.001179	0.007441	-3.49684	Down	transmembrane protein 252
TFPI2	-2.15359	3.66E-07	1.37E-05	-6.09249	Down	Tissue factor pathway inhibitor 2
MYH3	-2.14535	7.26E-05	0.000855	-4.43002	Down	Myosin heavy chain 3
DIO3	-2.14233	0.000107	0.001161	-4.30373	Down	Iodothyronine deiodinase 3
DYSF	-2.14228	1.95E-05	0.000304	-4.85115	Down	Dysferlin
EPB41L3	-2.14036	0.013569	0.049032	-2.58408	Down	Erythrocyte membrane protein band 4.1-like 3
TLR3	-2.14034	0.000506	0.003864	-3.78855	Down	Toll-like receptor 3
ANGPTL1	-2.13393	0.005354	0.024253	-2.9474	Down	Angiopoietin-like 1
SPTLC3	-2.13272	0.000242	0.002174	-4.03589	Down	Serine palmitoyltransferase long chain base subunit 3
PPP1R12B	-2.12791	6.22E-09	5.76E-07	-7.3677	Down	Protein phosphatase 1 regulatory subunit 12B
VSTM2L	-2.12654	1.64E-07	7.01E-06	-6.34319	Down	V-set and transmembrane domain containing 2-like
CELF2	-2.12104	8.02E-05	0.000926	-4.39815	Down	CUGBP Elav-like family member 2
MARCHF3	-2.1184	2.49E-07	9.90E-06	-6.21202	Down	Membrane-associated ring-CH-type finger 3
ANGPTL4	-2.11488	1.09E-05	0.000195	-5.03512	Down	Angiopoietin-like 4
SLC1A7	-2.11257	1.46E-07	6.43E-06	-6.37842	Down	Solute carrier family 1 member 7
EPHB2	-2.11095	2.06E-13	2.59E-10	-10.8454	Down	EPH receptor B2
ZBED9	-2.10312	3.24E-12	1.93E-09	-9.86565	Down	Zinc finger BED-type containing 9
FLI1	-2.09763	7.54E-09	6.81E-07	-7.30668	Down	Fli-1 proto-oncogene, ETS transcription factor

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
RASSF5	-2.09562	1.06E-07	5.03E-06	-6.47681	Down	Ras association domain family member 5
THBS1	-2.09173	1.78E-07	7.50E-06	-6.31621	Down	Thrombospondin 1
SLC66A1L	-2.08339	0.00011	0.001177	-4.29665	Down	Solute carrier family 66 member 1-like
GCH1	-2.08255	2.26E-06	5.64E-05	-5.52711	Down	GTP cyclohydrolase 1
MBP	-2.07759	6.50E-07	2.20E-05	-5.91444	Down	Myelin basic protein
PPP1R14A	-2.07597	8.56E-06	0.000162	-5.11027	Down	Protein phosphatase 1 regulatory inhibitor subunit 14A
MFAP3L	-2.0747	1.04E-06	3.12E-05	-5.76892	Down	Microfibril-associated protein 3-like
NXPH4	-2.066	2.63E-05	0.000389	-4.75555	Down	Neurexophilin 4
ID3	-2.06263	1.48E-07	6.45E-06	-6.37456	Down	Inhibitor of DNA binding 3
TNNT2	-2.06055	0.002524	0.013521	-3.22568	Down	Troponin T2, cardiac type
LIMCH1	-2.06017	4.90E-08	2.86E-06	-6.7188	Down	LIM and calponin homology domains 1
NOX4	-2.0487	1.70E-08	1.26E-06	-7.05129	Down	NADPH oxidase 4
CTRB2	-2.0478	4.57E-06	9.78E-05	-5.30672	Down	Chymotrypsinogen B2
CGREF1	-2.04395	0.001147	0.007301	-3.50646	Down	Cell growth regulator with EF-hand domain 1
LHX9	-2.04248	0.00036	0.002971	-3.90358	Down	LIM homeobox 9
S1PR5	-2.02711	1.44E-07	6.36E-06	-6.38316	Down	Sphingosine-1-phosphate receptor 5
NXPH3	-2.02511	1.51E-07	6.55E-06	-6.36811	Down	Neurexophilin 3
CACNG8	-2.02501	1.26E-08	9.95E-07	-7.14593	Down	Calcium voltage-gated channel auxiliary subunit gamma 8
PLA2G4A	-2.02297	0.0054	0.024408	-2.94417	Down	Phospholipase A2 group IVA
SERPINE2	-2.02191	5.41E-05	0.000682	-4.52496	Down	Serpin family E member 2
OR2W3	-2.01915	4.71E-05	0.00061	-4.56978	Down	Olfactory receptor family 2 subfamily W member 3
RAPGEF4	-2.01695	1.31E-07	5.97E-06	-6.41194	Down	Rap guanine nucleotide exchange factor 4
ALDH1B1	-2.01467	2.16E-07	8.79E-06	-6.25731	Down	Aldehyde dehydrogenase 1 family member B1
PRUNE2	-2.00548	7.40E-06	0.000144	-5.15609	Down	Prune homolog 2 with BCH domain
TRH	-1.99889	3.24E-07	1.25E-05	-6.13018	Down	Thyrotropin releasing hormone
TINAGL1	-1.99794	7.06E-07	2.33E-05	-5.88874	Down	Tubulointerstitial nephritis antigen-like 1
SYTL2	-1.99772	6.49E-08	3.51E-06	-6.63095	Down	Synaptotagmin-like 2
ADAP1	-1.9933	1.87E-08	1.37E-06	-7.02063	Down	ArfGAP with dual PH domains 1
SHANK2	-1.98828	2.21E-07	8.96E-06	-6.24956	Down	SH3 and multiple ankyrin repeat domains 2
IGSF9	-1.98524	3.33E-05	0.000466	-4.68027	Down	Immunoglobulin superfamily member 9
PI16	-1.98436	0.000958	0.00636	-3.56934	Down	Peptidase inhibitor 16
TLR1	-1.9714	1.04E-06	3.12E-05	-5.7691	Down	Toll-like receptor 1
STXBP6	-1.97135	0.000136	0.001404	-4.22513	Down	Syntaxin binding protein 6
CRLF1	-1.96167	0.002322	0.012654	-3.25588	Down	Cytokine receptor-like factor 1
EN1	-1.94422	0.005523	0.024788	-2.93568	Down	Engrailed homeobox 1
RSPO1	-1.94117	0.007895	0.032555	-2.79861	Down	R-spondin 1
PCSK1	-1.93847	0.012658	0.04649	-2.61212	Down	Proprotein convertase subtilisin/kexin type 1
CALB2	-1.93536	0.00306	0.015621	-3.15552	Down	Calbindin 2
TGFBI	-1.93292	8.67E-13	7.58E-10	-10.3288	Down	Transforming growth factor beta-induced
IGF2BP1	-1.93152	0.00046	0.003586	-3.82035	Down	Insulin-like growth factor 2 mRNA binding protein 1
SALL4	-1.92736	9.28E-05	0.001039	-4.35048	Down	Spalt-like transcription factor 4
EYA4	-1.91854	0.000274	0.002389	-3.99513	Down	EYA transcriptional coactivator and phosphatase 4
KLHL4	-1.91835	0.012076	0.044816	-2.63101	Down	Kelch-like family member 4
KRT18	-1.91099	2.60E-08	1.79E-06	-6.91712	Down	Keratin 18

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
ITGB1BP2	-1.90746	0.004274	0.020316	-3.03203	Down	Integrin subunit beta 1 binding protein 2
CCDC80	-1.90172	3.46E-07	1.31E-05	-6.10983	Down	Coiled-coil domain containing 80
DAPK1	-1.89532	0.000177	0.001711	-4.13957	Down	Death-associated protein kinase 1
WNK4	-1.89353	9.28E-08	4.56E-06	-6.51956	Down	WNK lysine deficient protein kinase 4
KRT13	-1.89249	0.008927	0.03562	-2.7507	Down	Keratin 13
DLX5	-1.89124	0.001255	0.007816	-3.47494	Down	Distal-less homeobox 5
FMO3	-1.88354	9.88E-05	0.001093	-4.33029	Down	Flavin containing dimethylaniline monooxygenase 3
VLDLR	-1.87827	2.11E-09	2.51E-07	-7.71161	Down	Very low density lipoprotein receptor
VDR	-1.87723	7.82E-07	2.51E-05	-5.85697	Down	Vitamin D receptor
GPC4	-1.86856	1.06E-07	5.02E-06	-6.47928	Down	Glypican 4
FAM20A	-1.86013	9.50E-08	4.63E-06	-6.51229	Down	FAM20A golgi-associated secretory pathway pseudokinase
FOXS1	-1.85885	0.002611	0.013865	-3.21349	Down	Forkhead box S1
LZTS1	-1.85517	1.62E-05	0.000265	-4.90959	Down	Leucine zipper tumor suppressor 1
RARB	-1.85254	6.68E-08	3.57E-06	-6.62213	Down	Retinoic acid receptor beta
KISS1	-1.85075	0.008338	0.033908	-2.77737	Down	KiSS-1 metastasis suppressor
GEM	-1.85038	4.00E-10	7.44E-08	-8.24801	Down	GTP binding protein overexpressed in skeletal muscle
FZD7	-1.84689	4.94E-05	0.000632	-4.55449	Down	Frizzled class receptor 7
HOXB6	-1.8464	0.000111	0.00119	-4.29191	Down	Homeobox B6
PTPRN	-1.84327	1.64E-05	0.000268	-4.90588	Down	Protein tyrosine phosphatase receptor type N
ZFH2	-1.84083	1.18E-08	9.47E-07	-7.16493	Down	Zinc finger homeobox 2
CELSR2	-1.83846	1.08E-08	8.88E-07	-7.1931	Down	Cadherin EGF LAG seven-pass G-type receptor 2
OR51E2	-1.83269	0.000424	0.003368	-3.84843	Down	Olfactory receptor family 51 subfamily E member 2
OMG	-1.82461	0.00021	0.00195	-4.0825	Down	Oligodendrocyte myelin glycoprotein
TLCD4	-1.8233	1.34E-07	6.05E-06	-6.40593	Down	TLC domain containing 4
EPHA4	-1.81892	4.29E-06	9.30E-05	-5.32687	Down	EPH receptor A4
EN2	-1.81816	1.65E-06	4.45E-05	-5.62445	Down	Engrailed homeobox 2
LRRC38	-1.81694	0.003933	0.01899	-3.06297	Down	Leucine-rich repeat containing 38
PODN	-1.81065	3.33E-06	7.59E-05	-5.4062	Down	Podocan
SCN4B	-1.80981	1.91E-06	4.98E-05	-5.57981	Down	Sodium voltage-gated channel beta subunit 4
FMO1	-1.80856	0.001182	0.007455	-3.49585	Down	Flavin containing dimethylaniline monooxygenase 1
DAW1	-1.80436	2.04E-05	0.000316	-4.83563	Down	Dynein assembly factor with WD repeats 1
SORCS2	-1.80129	0.00011	0.001185	-4.29419	Down	Sortilin-related VPS10 domain containing receptor 2
CDH8	-1.798	0.000148	0.001495	-4.19805	Down	Cadherin 8
ENC1	-1.78484	8.51E-07	2.67E-05	-5.83061	Down	Ectodermal-neural cortex 1
IFNE	-1.78476	9.46E-07	2.90E-05	-5.79788	Down	Interferon epsilon
TNNT1	-1.78213	1.64E-05	0.000268	-4.90515	Down	Troponin T1, slow skeletal type
ANKRD1	-1.7785	0.000493	0.003783	-3.79748	Down	Ankyrin repeat domain 1
TBILA	-1.77649	9.15E-06	0.00017	-5.08928	Down	TGF-beta-induced lncRNA
SGCA	-1.77282	0.009194	0.036483	-2.73918	Down	Sarcoglycan alpha
ZFPM2	-1.76616	1.92E-08	1.39E-06	-7.01159	Down	Zinc finger protein, FOG family member 2
SOX9	-1.76309	0.000564	0.004211	-3.75153	Down	SRY-box transcription factor 9
SLC22A23	-1.76019	1.74E-08	1.29E-06	-7.04295	Down	Solute carrier family 22 member 23
MGP	-1.75918	1.22E-05	0.000213	-5.00007	Down	Matrix Gla protein

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
SULF1	-1.75807	4.24E-06	9.22E-05	-5.3303	Down	Sulfatase 1
IL20RA	-1.75316	0.006167	0.02697	-2.8937	Down	Interleukin 20 receptor subunit alpha
PIPOX	-1.75171	1.39E-09	1.85E-07	-7.84567	Down	Pipelicolic acid and sarcosine oxidase
DEPP1	-1.75008	0.000166	0.001626	-4.16145	Down	DEPP autophagy regulator 1
CYP24A1	-1.74299	9.00E-06	0.000169	-5.09464	Down	Cytochrome P450 family 24 subfamily A member 1
BACH2	-1.73896	4.49E-08	2.70E-06	-6.74624	Down	BTB domain and CNC homolog 2
DNAH11	-1.73836	5.56E-06	0.000114	-5.24577	Down	Dynein axonemal heavy chain 11
PKIA	-1.7378	6.72E-07	2.25E-05	-5.90405	Down	cAMP-dependent protein kinase inhibitor alpha
ID2	-1.73319	5.43E-09	5.12E-07	-7.41108	Down	Inhibitor of DNA binding 2
MCTP2	-1.73276	0.011965	0.044491	-2.63472	Down	Multiple C2 and transmembrane domain containing 2
WSCD1	-1.72857	2.68E-05	0.000395	-4.74914	Down	WSC domain containing 1
STEAP1B	-1.72452	1.39E-06	3.93E-05	-5.67797	Down	STEAP family member 1B
TLR2	-1.72425	2.42E-06	5.95E-05	-5.50499	Down	Toll-like receptor 2
ARHGEF37	-1.7203	8.61E-09	7.56E-07	-7.26502	Down	Rho guanine nucleotide exchange factor 37
IL6	-1.71293	0.002096	0.011676	-3.29281	Down	Interleukin 6
KLHL23	-1.70426	2.22E-09	2.61E-07	-7.69491	Down	Kelch-like family member 23
NPPB	-1.70388	0.004912	0.022698	-2.97995	Down	Natriuretic peptide B
DIO3OS	-1.70365	2.75E-05	0.000402	-4.74205	Down	DIO3 opposite strand upstream RNA
OVCH2	-1.70131	0.000263	0.002311	-4.00871	Down	Ovochymase 2
FRMD5	-1.69745	1.82E-09	2.25E-07	-7.75951	Down	FERM domain containing 5
SPINK1	-1.6942	0.000938	0.006255	-3.57646	Down	Serine peptidase inhibitor Kazal type 1
GAS6	-1.69335	8.35E-07	2.63E-05	-5.83652	Down	Growth arrest specific 6
GPC3	-1.69025	0.008756	0.035122	-2.75827	Down	Glypican 3
NTRK3	-1.688	3.78E-05	0.000508	-4.64039	Down	Neurotrophic receptor tyrosine kinase 3
SLC6A15	-1.68799	0.001765	0.010206	-3.35421	Down	Solute carrier family 6 member 15
FBXO32	-1.68771	1.37E-07	6.12E-06	-6.39812	Down	F-box protein 32
CCNA1	-1.68516	0.000112	0.001194	-4.29037	Down	Cyclin A1
AMIGO2	-1.68485	2.17E-05	0.000332	-4.81716	Down	Adhesion molecule with Ig-like domain 2
F11R	-1.68051	0.000754	0.005261	-3.65216	Down	F11 receptor
HPGDS	-1.67767	0.000369	0.003033	-3.89491	Down	Hematopoietic prostaglandin D synthase
MPP2	-1.67247	3.88E-05	0.000521	-4.63125	Down	MAGUK p55 scaffold protein 2
RAB11FIP1	-1.66972	3.72E-11	1.28E-08	-9.03219	Down	RAB11 family interacting protein 1
SIGLEC15	-1.66662	0.010585	0.040571	-2.68356	Down	Sialic acid binding Ig-like lectin 15
LNCSRLR	-1.66631	6.95E-05	0.000829	-4.44413	Down	lncRNA sorafenib resistance in renal cell carcinoma associated
FOXD1	-1.66456	0.000243	0.002178	-4.03439	Down	Forkhead box D1
CCNO	-1.6641	1.93E-05	0.000302	-4.85415	Down	Cyclin O
ADAM12	-1.65893	8.34E-09	7.36E-07	-7.27483	Down	ADAM metallopeptidase domain 12
BISPR	-1.65887	1.88E-05	0.000297	-4.86248	Down	BST2 interferon stimulated positive regulator
BEGAIN	-1.65732	0.004062	0.019486	-3.05102	Down	Brain-enriched guanylate kinase associated
RASSF2	-1.65325	0.009	0.035841	-2.74754	Down	Ras association domain family member 2
CCDC110	-1.65218	0.0077	0.031935	-2.80834	Down	Coiled-coil domain containing 110
CTRB1	-1.64621	0.001089	0.007023	-3.52466	Down	Chymotrypsinogen B1
ALDH8A1	-1.64599	4.97E-05	0.000635	-4.55206	Down	Aldehyde dehydrogenase 8 family member A1
LGALS2	-1.64481	2.30E-07	9.25E-06	-6.23679	Down	Galectin 2
SEL1L2	-1.63989	0.005162	0.023628	-2.96122	Down	SEL1L2 adaptor subunit of ERAD E3 ligase
HAS3	-1.63832	0.000108	0.001167	-4.30132	Down	Hyaluronan synthase 3

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
NCAM1	-1.63777	0.002841	0.014783	-3.18274	Down	Neural cell adhesion molecule 1
LGI2	-1.62905	0.00797	0.032818	-2.79492	Down	Leucine-rich repeat LGI family member 2
ABCA9	-1.62869	0.000622	0.004553	-3.71794	Down	ATP binding cassette subfamily A member 9
CALHM5	-1.62299	6.38E-08	3.47E-06	-6.63653	Down	Calcium homeostasis modulator family member 5
MYOC	-1.62137	0.004188	0.019999	-3.03963	Down	Myocilin
ZFHX4	-1.61935	1.93E-06	5.01E-05	-5.57544	Down	Zinc finger homeobox 4
PPFIA4	-1.61786	8.60E-06	0.000162	-5.10896	Down	PTPRF interacting protein alpha 4
HAS1	-1.61211	0.002204	0.012145	-3.27468	Down	Hyaluronan synthase 1
GCSAM	-1.60977	0.000286	0.002475	-3.9807	Down	Germinal center-associated signaling and motility
HOXB7	-1.60906	0.000289	0.002504	-3.97632	Down	Homeobox B7
GALNT9	-1.60616	0.000323	0.002731	-3.93973	Down	Polypeptide N-acetylgalactosaminyltransferase 9
DUSP2	-1.60389	0.000427	0.003391	-3.84547	Down	Dual specificity phosphatase 2
USP2	-1.60125	6.08E-08	3.33E-06	-6.65132	Down	Ubiquitin specific peptidase 2
RAB3IP	-1.60019	8.70E-13	7.58E-10	-10.3278	Down	RAB3A interacting protein
ABCA10	-1.59931	0.000167	0.001636	-4.15891	Down	ATP binding cassette subfamily A member 10
KLF2	-1.59915	0.000152	0.001521	-4.19038	Down	KLF transcription factor 2
SIK1	-1.59191	0.004429	0.020911	-3.01874	Down	Salt inducible kinase 1
EEF1A2	-1.58379	0.004251	0.020235	-3.03407	Down	Eukaryotic translation elongation factor 1 alpha 2
OR2A7	-1.58343	6.96E-06	0.000137	-5.17537	Down	Olfactory receptor family 2 subfamily A member 7
SIX1	-1.58277	7.30E-06	0.000142	-5.16045	Down	SIX homeobox 1
FGF7	-1.57915	5.70E-10	1.01E-07	-8.13309	Down	Fibroblast growth factor 7
ESYT3	-1.5739	2.49E-05	0.000373	-4.77272	Down	Extended synaptotagmin 3
CCN2	-1.56954	4.32E-06	9.33E-05	-5.32471	Down	Cellular communication network factor 2
JCAD	-1.56924	3.80E-09	3.97E-07	-7.5239	Down	Junctional cadherin 5 associated
MAP3K9	-1.56819	9.01E-06	0.000169	-5.09419	Down	Mitogen-activated protein kinase kinase kinase 9
SUPT20HL1	-1.56464	0.000373	0.003056	-3.89128	Down	SUPT20H-like 1
MYOZ3	-1.56364	0.000805	0.005542	-3.62926	Down	Myozenin 3
SMIM10L2A	-1.56314	0.000163	0.001606	-4.16698	Down	Small integral membrane protein 10-like 2A
ABCA8	-1.55861	0.001604	0.009481	-3.3883	Down	ATP binding cassette subfamily A member 8
EXPH5	-1.55444	5.56E-05	0.000697	-4.51595	Down	Exophilin 5
DAB1	-1.55404	0.003521	0.017423	-3.10397	Down	DAB adaptor protein 1
PKDCC	-1.55391	7.68E-06	0.000148	-5.14443	Down	Protein kinase domain containing, cytoplasmic
PLPPR3	-1.54125	0.004043	0.019408	-3.05277	Down	Phospholipid phosphatase related 3
MYH10	-1.53969	8.54E-07	2.67E-05	-5.82944	Down	Myosin heavy chain 10
FRMPD4	-1.53684	8.55E-05	0.000977	-4.37704	Down	FERM and PDZ domain containing 4
IGFBP3	-1.53514	5.73E-05	0.000709	-4.50655	Down	Insulin-like growth factor binding protein 3
FANK1	-1.52718	8.24E-09	7.29E-07	-7.27887	Down	Fibronectin type III and ankyrin repeat domains 1
IFITM10	-1.52693	9.56E-09	8.12E-07	-7.23185	Down	Interferon-induced transmembrane protein 10
MPP7	-1.52412	2.20E-05	0.000336	-4.81221	Down	MAGUK p55 scaffold protein 7
PLXDC2	-1.52327	0.002033	0.0114	-3.30371	Down	Plexin domain containing 2
GRAMD1B	-1.51361	1.09E-14	2.25E-11	-11.9416	Down	GRAM domain containing 1B
ARHGDI3	-1.50611	0.004765	0.022182	-2.99136	Down	Rho GDP dissociation inhibitor gamma

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
TRIM63	-1.50548	0.002381	0.012902	-3.24686	Down	Tripartite motif containing 63
SLC4A4	-1.50352	3.21E-08	2.09E-06	-6.85153	Down	Solute carrier family 4 member 4
SH3RF2	-1.50251	0.011665	0.043547	-2.64489	Down	SH3 domain containing ring finger 2
HRNR	-1.49315	0.001072	0.006929	-3.53015	Down	Hornerin
SLC9C1	-1.49104	5.78E-06	0.000117	-5.23371	Down	Solute carrier family 9 member C1
HUNK	-1.48709	1.56E-05	0.000258	-4.92035	Down	Hormonally up-regulated Neu-associated kinase
CYRIA	-1.48637	0.003429	0.017064	-3.11375	Down	CYFIP-related Rac1 interactor A
LYVE1	-1.48603	4.33E-05	0.00057	-4.59653	Down	Lymphatic vessel endothelial hyaluronan receptor 1
TMEM38A	-1.48316	4.11E-09	4.20E-07	-7.49924	Down	Transmembrane protein 38A
BEND5	-1.48116	0.00269	0.014195	-3.20264	Down	BEN domain containing 5
SLAIN1	-1.48056	0.000233	0.002111	-4.04819	Down	SLAIN motif family member 1
PRKD1	-1.47989	2.42E-09	2.82E-07	-7.66775	Down	Protein kinase D1
CYP3A7	-1.47698	0.002711	0.01429	-3.19973	Down	Cytochrome P450 family 3 subfamily A member 7
TTC9	-1.4755	0.000904	0.006079	-3.58943	Down	Tetratricopeptide repeat domain 9
CHL1	-1.47355	0.003068	0.015656	-3.15461	Down	Cell adhesion molecule L1-like
CILP2	-1.47199	0.000823	0.005642	-3.62168	Down	Cartilage intermediate layer protein 2
KCNG2	-1.46863	0.002174	0.012006	-3.27964	Down	Potassium voltage-gated channel modifier subfamily G member 2
PLD5	-1.46805	0.000466	0.003624	-3.81645	Down	Phospholipase D family member 5
BGN	-1.46712	0.000467	0.00363	-3.81575	Down	Biglycan
SLC35F2	-1.4638	8.63E-10	1.34E-07	-7.99901	Down	Solute carrier family 35 member F2
EPPK1	-1.46283	0.010978	0.041646	-2.66907	Down	Epiplakin 1
SIRPB1	-1.4611	0.000175	0.001699	-4.1432	Down	Signal regulatory protein beta 1
EDA	-1.46025	0.003186	0.016118	-3.14073	Down	Ectodysplasin A
MYEF2	-1.45681	4.38E-07	1.60E-05	-6.03714	Down	Myelin expression factor 2
STEAP2	-1.4563	0.000201	0.001889	-4.09682	Down	STEAP2 metalloreductase
PARP15	-1.45533	5.85E-05	0.000721	-4.50001	Down	Poly(ADP-ribose) polymerase family member 15
ACTBL2	-1.45475	0.003908	0.018897	-3.06536	Down	Actin beta-like 2
CPEB1	-1.45421	4.20E-06	9.13E-05	-5.33383	Down	Cytoplasmic polyadenylation element binding protein 1
NTRK2	-1.45301	0.00721	0.030366	-2.83373	Down	Neurotrophic receptor tyrosine kinase 2
SYT15	-1.45278	4.46E-06	9.58E-05	-5.31449	Down	Synaptotagmin 15
HSD17B6	-1.45106	2.74E-05	0.000402	-4.74239	Down	Hydroxysteroid 17-beta dehydrogenase 6
FPR3	-1.45054	0.013294	0.048282	-2.59237	Down	Formyl peptide receptor 3
SLIT2	-1.44843	0.000778	0.005397	-3.64129	Down	Slit guidance ligand 2
IRX3	-1.448	0.008462	0.034258	-2.77161	Down	Iroquois homeobox 3
MPO	-1.44767	9.22E-06	0.000171	-5.08704	Down	Myeloperoxidase
MANEA-DT	-1.44692	7.82E-06	0.00015	-5.13889	Down	MANEA divergent transcript
COL8A2	-1.44373	0.000319	0.002698	-3.94428	Down	Collagen type VIII alpha 2 chain
ACTC1	-1.44263	0.001093	0.007041	-3.52346	Down	Actin alpha cardiac muscle 1
FLG	-1.43718	2.18E-05	0.000333	-4.81474	Down	Filaggrin
HEG1	-1.43712	9.45E-08	4.62E-06	-6.51394	Down	Heart development protein with EGF-like domains 1
NRP2	-1.43679	1.83E-11	7.17E-09	-9.27083	Down	Neuropilin 2
B3GALT5	-1.43583	0.007636	0.031726	-2.81152	Down	Beta-1,3-galactosyltransferase 5
NNMT	-1.42889	2.33E-11	8.81E-09	-9.18978	Down	Nicotinamide N-methyltransferase
CHI3L1	-1.42851	0.002587	0.013767	-3.21683	Down	Chitinase 3-like 1

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
TNIK	-1.42153	2.61E-05	0.000387	-4.75795	Down	TRAF2 and NCK interacting kinase
LMCD1	-1.42114	0.000221	0.002026	-4.06669	Down	LIM and cysteine-rich domains 1
RHD	-1.41493	0.004381	0.020742	-3.02287	Down	Rh blood group D antigen
STEAP1	-1.41	3.84E-06	8.53E-05	-5.36171	Down	STEAP family member 1
SP6	-1.40925	0.000606	0.004456	-3.72701	Down	Sp6 transcription factor
EXTL1	-1.40841	9.26E-05	0.001038	-4.35128	Down	Exostosin-like glycosyltransferase 1
RUNX1	-1.40441	1.85E-07	7.71E-06	-6.30475	Down	RUNX family transcription factor 1
NFIB	-1.40425	1.88E-06	4.94E-05	-5.5835	Down	Nuclear factor I B
APLN	-1.39953	0.010642	0.04075	-2.68144	Down	Apelin
ANKRD29	-1.39607	7.42E-07	2.40E-05	-5.87325	Down	Ankyrin repeat domain 29
SORBS1	-1.39462	0.001393	0.008472	-3.43827	Down	Sorbin and SH3 domain containing 1
EPB41	-1.39438	4.81E-07	1.72E-05	-6.00756	Down	Erythrocyte membrane protein band 4.1
GZMM	-1.38989	0.000363	0.002991	-3.90077	Down	Granzyme M
PDGFRL	-1.38791	2.18E-05	0.000333	-4.81524	Down	Platelet-derived growth factor receptor-like
ART4	-1.38626	0.001104	0.007088	-3.51966	Down	ADP-ribosyltransferase 4 (inactive) (Dombrock blood group)
ACKR2	-1.38274	0.006722	0.028841	-2.86071	Down	Atypical chemokine receptor 2
MGST1	-1.37937	2.58E-05	0.000384	-4.76127	Down	Microsomal glutathione S-transferase 1
FAM180A	-1.37911	1.42E-05	0.00024	-4.95043	Down	Family with sequence similarity 180 member A
FPR1	-1.3785	2.49E-06	6.07E-05	-5.49678	Down	Formyl peptide receptor 1
STOX2	-1.37795	0.000718	0.005098	-3.66866	Down	Storkhead box 2
ST3GAL6	-1.37756	9.44E-09	8.10E-07	-7.23598	Down	ST3 beta-galactoside alpha-2,3-sialyltransferase 6
PLP1	-1.37284	0.000225	0.002056	-4.05966	Down	Proteolipid protein 1
MYOM2	-1.37149	0.00055	0.004123	-3.76021	Down	Myomesin 2
NBPF14	-1.36956	7.92E-15	2.24E-11	-12.0651	Down	NBPF member 14
NAV2	-1.36928	5.56E-11	1.66E-08	-8.8983	Down	Neuron navigator 2
PDE7B	-1.36867	8.42E-07	2.65E-05	-5.8338	Down	Phosphodiesterase 7B
INSYN2A	-1.36767	4.47E-05	0.000583	-4.58645	Down	Inhibitory synaptic factor 2A
MYOZ1	-1.36713	0.000202	0.001894	-4.09555	Down	Myozenin 1
PMEL	-1.36522	0.006691	0.028726	-2.86246	Down	Premelanosome protein
SLC35F4	-1.3639	0.000534	0.004042	-3.76997	Down	Solute carrier family 35 member F4
COL4A3	-1.36303	0.007824	0.032335	-2.80209	Down	Collagen type IV alpha 3 chain
GAS6-DT	-1.36132	1.02E-05	0.000185	-5.05604	Down	GAS6 divergent transcript
KCNQ4	-1.36021	2.07E-06	5.26E-05	-5.55373	Down	Potassium voltage-gated channel subfamily Q member 4
SSPN	-1.35519	1.41E-06	3.95E-05	-5.67467	Down	Sarcospan
PPP1R13L	-1.35489	4.31E-05	0.000568	-4.59787	Down	Protein phosphatase 1 regulatory subunit 13-like
SEMA3C	-1.35338	1.20E-05	0.00021	-5.004	Down	Semaphorin 3C
BMAL2	-1.35187	1.74E-05	0.000281	-4.88662	Down	Basic helix-loop-helix ARNT-like 2
ZNF334	-1.35102	1.77E-09	2.22E-07	-7.76728	Down	Zinc finger protein 334
SLFN1	-1.34826	1.10E-05	0.000197	-5.03039	Down	Schlafen-like 1
FNDC1	-1.34778	0.003748	0.018272	-3.08084	Down	Fibronectin type III domain containing 1
DCLK2	-1.34715	1.44E-08	1.11E-06	-7.10315	Down	Doublecortin-like kinase 2
NOTCH2	-1.34466	1.59E-12	1.10E-09	-10.1137	Down	Notch receptor 2
DSE	-1.34245	5.19E-13	5.71E-10	-10.5116	Down	Dermatan sulfate epimerase
ZNF185	-1.33924	1.72E-09	2.18E-07	-7.77811	Down	Zinc finger protein 185 with LIM domain
IQSEC3	-1.33771	1.44E-07	6.36E-06	-6.38305	Down	IQ motif and Sec7 domain ArfGEF 3
P4HA3	-1.33558	0.00265	0.014033	-3.20809	Down	Prolyl 4-hydroxylase subunit alpha 3

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
JPH3	-1.33528	0.000135	0.001393	-4.22789	Down	Junctophilin 3
SLC38A3	-1.33466	0.000348	0.002899	-3.91474	Down	Solute carrier family 38 member 3
PTGDR	-1.33221	0.005916	0.026081	-2.90954	Down	Prostaglandin D2 receptor
TMEM255A	-1.33178	0.002105	0.011716	-3.29122	Down	Transmembrane protein 255A
PALM2AKAP2	-1.33037	0.000609	0.004471	-3.72537	Down	PALM2 and AKAP2 fusion
SEC14L6	-1.32791	0.005769	0.02558	-2.91909	Down	SEC14-like lipid binding 6
PDK4	-1.32376	0.009379	0.037081	-2.73135	Down	Pyruvate dehydrogenase kinase 4
PITPNM3	-1.32314	2.40E-06	5.91E-05	-5.50838	Down	PITPNM family member 3
GNG7	-1.32272	0.001096	0.007055	-3.52242	Down	G protein subunit gamma 7
PGF	-1.32205	0.003132	0.015905	-3.1471	Down	Placental growth factor
THBS2	-1.31978	2.70E-05	0.000396	-4.74785	Down	Thrombospondin 2
DBNDD1	-1.31563	0.000102	0.001117	-4.32009	Down	Dysbindin domain containing 1
GPR143	-1.31545	6.83E-07	2.27E-05	-5.8991	Down	G protein-coupled receptor 143
GSTO2	-1.31528	1.57E-08	1.18E-06	-7.07604	Down	Glutathione S-transferase omega 2
LRRC2	-1.31492	4.15E-06	9.08E-05	-5.33705	Down	Leucine-rich repeat containing 2
DNAH6	-1.31229	0.001233	0.007709	-3.4811	Down	Dynein axonemal heavy chain 6
ANK3	-1.30956	0.001305	0.008066	-3.46113	Down	Ankyrin 3
DES	-1.30838	0.005578	0.024982	-2.93188	Down	Desmin
KRT86	-1.30784	0.001565	0.009285	-3.39716	Down	Keratin 86
PDLIM5	-1.30783	2.97E-09	3.30E-07	-7.60251	Down	PDZ and LIM domain 5
PRDM6	-1.30782	0.000118	0.001252	-4.27173	Down	PR/SET domain 6
RPS6KA2-IT1	-1.30648	0.002511	0.013464	-3.22766	Down	RPS6KA2 intronic transcript 1
IGFBPL1	-1.30493	0.000887	0.005985	-3.59605	Down	Insulin-like growth factor binding protein-like 1
TRABD2A	-1.30183	0.008619	0.034725	-2.76445	Down	TraB domain containing 2A
HTR6	-1.30181	0.000196	0.001852	-4.10637	Down	5-Hydroxytryptamine receptor 6
NAV1	-1.30062	2.60E-14	4.67E-11	-11.6111	Down	Neuron navigator 1
HSPB7	-1.29905	0.000499	0.003828	-3.79288	Down	Heat shock protein family B (small) member 7
NPAS1	-1.29461	0.000511	0.003897	-3.78524	Down	Neuronal PAS domain protein 1
PNPLA1	-1.29452	0.001027	0.006704	-3.54508	Down	Patatin-like phospholipase domain containing 1
NFIX	-1.29301	1.81E-07	7.57E-06	-6.31162	Down	Nuclear factor I X
PIK3AP1	-1.29075	0.002458	0.013241	-3.23532	Down	Phosphoinositide-3-kinase adaptor protein 1
NRG1	-1.28956	0.001135	0.007242	-3.51	Down	Neuregulin 1
DMRT2	-1.2883	0.001355	0.008303	-3.44799	Down	Doublesex and mab-3-related transcription factor 2
CCDC71L	-1.28655	1.36E-05	0.000232	-4.96414	Down	Coiled-coil domain containing 71-like
LIF	-1.28585	0.001024	0.006695	-3.54602	Down	LIF interleukin 6 family cytokine
MCUB	-1.28452	9.52E-09	8.12E-07	-7.23327	Down	Mitochondrial calcium uniporter dominant negative subunit beta
ADH1C	-1.28217	0.005486	0.024689	-2.93817	Down	Alcohol dehydrogenase 1C (class I), gamma polypeptide
SLC16A11	-1.28094	0.001365	0.008339	-3.44544	Down	Solute carrier family 16 member 11
SIRPA	-1.28048	5.30E-09	5.06E-07	-7.41881	Down	Signal regulatory protein alpha
B3GNT7	-1.27747	5.43E-07	1.91E-05	-5.9701	Down	UDP-GlcNAc:betaGal beta-1,3-N-acetylglucosaminyltransferase 7
NUAK1	-1.27407	4.13E-05	0.00055	-4.61137	Down	NUAK family kinase 1
CHRM4	-1.27138	9.69E-05	0.001076	-4.33666	Down	Cholinergic receptor muscarinic 4
FAM131C	-1.2709	0.000197	0.001861	-4.10365	Down	Family with sequence similarity 131 member C

Table 1 (continued)

Gene symbol	logFC	p Value	adj.P.Val	tvalue	Regulation	Gene name
CACNA1I	-1.26965	0.000148	0.00149	-4.19925	Down	Calcium voltage-gated channel subunit alpha 1 I
PLK2	-1.26873	1.57E-06	4.30E-05	-5.64037	Down	Polo-like kinase 2
STX11	-1.2683	0.000107	0.001159	-4.30497	Down	Syntaxin 11
PALM	-1.26784	2.42E-05	0.000364	-4.78214	Down	Paralemmin
SPSB1	-1.26633	2.13E-07	8.72E-06	-6.26124	Down	splA/Ryanodine receptor domain and SOCS box containing 1
PFKP	-1.26587	5.77E-07	2.01E-05	-5.95129	Down	Phosphofructokinase, platelet
TMSB15B	-1.26556	0.002332	0.012706	-3.25431	Down	Thymosin beta 15B
CDKL2	-1.26526	0.010003	0.038886	-2.70594	Down	Cyclin-dependent kinase-like 2
CTAGE9	-1.2644	2.56E-06	6.17E-05	-5.48824	Down	CTAGE family member 9

DEGs with upregulated and downregulated expression in volcano plot and heatmap, respectively.

GO and pathway enrichment analyses of DEGs

GO enrichment analysis (<http://www.geneontology.org>) [35] (Accession Date: 26/02/2024) was frequently used to annotate the degree of gene function terms in DEGs, which included biological process (BP), cellular component (CC) and molecular function (MF). REACTOME (<https://reactome.org/>) (Accession Date: 26/02/2024) [36] pathway enrichment analysis was used to demonstrate enriched signaling pathways in DEGs. The g:Profiler (<http://biit.cs.ut.ee/gprofiler/>) (Accession Date: 26/02/2024) [37] was used to perform GO and REACTOME pathway enrichment analysis of DEGs. $P < 0.05$ was considered to represent statistical significance.

Construction of the PPI network and module analysis

To ensure the optimal graphical display of protein interactions of DEGs, Human Integrated Protein-Protein Interaction rEference (HIPIE) (<https://cn.string-db.org/>) [38] (Accession Date: 23/02/2024) was used to generate the PPI network. The software Cytoscape (version 3.10.1) (<http://www.cytoscape.org/>) [39] (Accession Date: 23/02/2024) was used to visualize the PPI network. The Network Analyzer in Cytoscape was utilized to calculate node degree [40], betweenness [41], stress [42] and closeness [43]. The PEWCC Cytoscape software plugin [44] was used to create modules in the PPI network of endometriosis.

Construction of the miRNA-hub gene regulatory network

The significant miRNAs were identified from miRNA-hub gene regulatory network analysis through the TarBase, miRTarBase, miRecords, miRanda (S mansoni only), miR2Disease, HMDD, PhenomiR, SM2miR,

PharmacomiR, EpimiR, starBase, TransmiR, ADmiRE and TAM 2 via miRNet database (<https://www.mirnet.ca/>) [45] (Accession Date: 26/02/2024). This network was visualized with Cytoscape [39] (Accession Date: 26/02/2024), and the significant hub genes and miRNAs were selected via the Network Analyzer plugin in Cytoscape based on the degree connectivity.

Construction of the TF-hub gene regulatory network

The significant transcription factors (TFs) were identified from TF-hub gene regulatory network analysis through the CHEA via NetworkAnalyst database (<https://www.networkanalyst.ca/>) [46] (Accession Date: 01/05/2024). This network was visualized with Cytoscape [39] (Accession Date: 01/05/2024), and the significant hub genes and TFs were selected via the Network Analyzer plugin in Cytoscape based on the degree connectivity.

Receiver operating characteristic curve (ROC) analysis

ROC analysis was performed to predict the diagnostic effectiveness of biomarkers by pROC package of R software [47]. The area under the curve (AUC) value was utilized to determine the diagnostic effectiveness in discriminating endometriosis from normal control samples.

Results

Identification of DEGs

The DEGs were screened by “limma” package ($|\logFC| > 1.304$ (log2 fold change) for upregulated genes and $|\logFC| > 1.304$ (log2 fold change) < -1.2644 for downregulated genes and $\text{adj.P.Val} < 0.05$). The GSE243039 dataset contained 958 DEGs, including 479 upregulated genes and 479 downregulated genes, and is listed in Table 1. The volcano plot (Fig. 1) was used to show the expression pattern of DEGs in endometriosis. The heatmap of the DEGs is shown in Fig. 2.

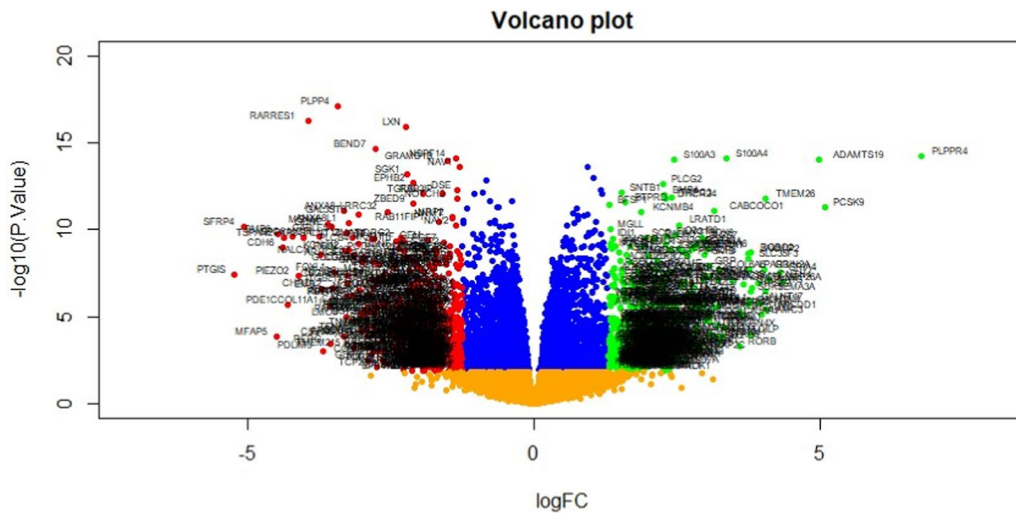


Fig. 1 Volcano plot of differentially expressed genes. Genes with a significant change of more than twofold were selected. Green dot represented upregulated significant genes and red dot represented downregulated significant genes

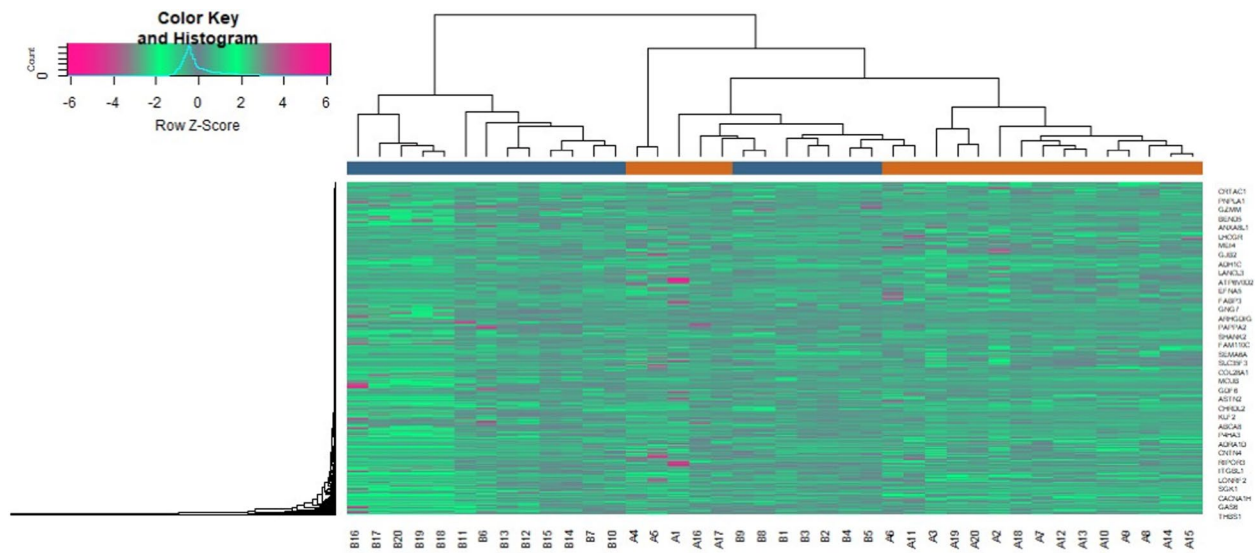


Fig. 2 Heat map of differentially expressed genes. Legend on the top left indicates log fold change of genes. (A1 – A20= Endometriosis samples; B1 – B20= Normal control samples)

GO and pathway enrichment analyses of DEGs

GO enrichment and REACTOME pathway enrichment analyses were performed on the DEGs using the g:Profiler database. GO enrichment analysis covers three aspects: BP, CC and MF (Table 2). The upregulated genes were mainly related to multicellular organismal process, regulation of biological process, membrane, extracellular region, signaling receptor binding and molecular transducer activity, while the downregulated genes were mainly involved in developmental process, biological regulation, cell periphery, cytoplasm, molecular function

regulator activity and calcium ion binding. The REACTOME pathway enrichment analysis showed that the genes upregulated genes in endometriosis were enriched in signaling by GPCR, extracellular matrix organization, muscle contraction and glycosaminoglycan metabolism (Table 3).

Construction of the PPI network and module analysis

Considering the critical role of protein interactions in protein function, we used the HIPIE database and Cytoscape software to generate PPI network once we had

Table 2 The enriched GO terms of the up- and downregulated differentially expressed genes

GO ID	CATEGORY	GO Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
Upregulated genes						
GO:0032501	BP	multicellular organismal process	8.48E-28	27.07135732	279	PLPPR4, PCSK9, KIF26A, CNTN4, APCDD1, SEMA3A, GRIN2A, LAMC3, L1CA, MCBLN2, SDK1, ROBO2, LPAR3, CACNA1G, AJAP1, RORB, HSD17B2, NLGN4, XVCAM1, STRA6, SIX2, CLEC3B, ST8SIA4, COCH, EFNA5, GRP, AVPR1A, ADGR L3, EPHA6, CNTN5, AQP8, WNT6, FABP4, PSG1, ANO1, SOX6, NDPPREX2, TFA P2C, NTRK1, CNTN1, FGF10, RAMP3, PLA2G7, MMP12, FAIM2, SEMA6A, ADC Y1, MDGA2, COL14A1, LCP1, ADRA1D, SNCA, CLIC2, AC3, IL31RA, APOD1N, SRR, LAG3, SOX18, F2RL2, CCR1, CST1, RETREG1, CYP39A1, JSM1, ATCAY, PRE X1, CARD16, ADRA2C, FLT1, FABP5, PRDM1, TRPC3, TUNAR, IGSF3, THSD7A, DKK2, PROK1, SCTR, WNT16, ZNF804A, RNF157, F13A1, SH3GL2, PRKCB, DH CR24, PDE3B, BMP4, IL1RL1, MYPN, HS3ST5, PLCG2, PRL, FOXO1, SEMA6D, N EFL, WNT5A, MEOX1, DOCK4, HTR1B, TIMP3, SERPINB2, COL7A1, FRZB, PSG 3, WWDC2, CRAGD, NLRP2, CPE, ADAMTS9, NPWX, K, EREG, GABRA2, NDNF, DC HS1, CNTN6, MEGF10, MEH4, SNAP25, SYTI, NETO1, CD24, SLC26A7, LHCGR, CLSTN2, SCD, DTX1, PDGFB, LSAMP, PIK3CG, MMRN1, LDLR, SOAT2, RGCC, C D4, FOXL2, DPF3, GPR183, TRPA1, PTGER4, EPHA5, F2RL3, KCNMB4, C1QL1, T OX, ADAMTS5, CST4, RSPO3, MYO7A, KRT17, ADGRB3, MAP2K6, ADAMTS1, PDE4B, NES, SH2D2A, TH, TMEM119, IRX6, PSG9, PADI2, CACNA1D, LRFN5, M MP8, KDR, CNFN, KRT36, ADRB2, ACKR3, P2RX6, CNGA1, GJA3, MGAT3, TNFR SF9, PTPRC, SEMA3D, MARK1, PITX1, EYAI1, TXK, KLPCARE, PLAU, EPHA7, ZN F365, PIK3R3, GJB2, CCDC141, HECW2, SOX8, CCND2, ATP1B2, RTN4R, PTGS 1, CRABP2, HECW1, PTPRD, PCDH9, INSIG1, BMP8A, CHRM2, EPCAM, WNT9 A, PRXL2A, FSIIP2, IRX2, SEMA3B, TSPAN12, SNTB1, BATF2, SIGLEC1, MBOAT1 ,UCN2, CRTAC1, ADGRG6, CYP2J2, CLEC14A, CYP11A1, ASTN2, NNTN1, GRIP2 ,ATP1A2, STAC2, PDGFD, AFF3, ADGRG1, TNFRSF19, CLDN3, MSTN, TMC1, L EPR, SDK2, DMRTA2, CD248, IL15, KREMEN1, CACNA1H, BMP2, PCDH8, H15, LA MA5, SYT7, NTNG1, NELL2, TCIM, TAS2R1, ZBTB46, PAK3, KRT19, ROS1, MME, APOE, CUBN, PTCH1, OSR2, ITPKA, RAB7B, RBM20, CASP1, PDE9A, MGLL, EFN B2, NID1, ABCG2, HMGCR, ACE, PPIA2, RNF17, BFP, SP1, PGR, RENBP, WLS, RIMS 1, KLIK3, MCOLN2, STMN2, ARSL, GREM2

Table 2 (continued)

GO ID	CATEGORY	GO Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
GO:0050789	BP	regulation of biological process	9.83E-12	11.00739244	330	PLPPR4, PCSK9, KIF26A, GRIA4, CNTN4, APCDD1, SEMA3A, GRIN2A, L1CAM, CBLN2, HPSE2, CLIP, SDK1, ROBO2, LPAR3, PALMID, CACNA1G, AJAP1, RORB, S100A4, NLGN4X, VCAM1, STRA6, SIX2, CLEC3B, MMP3, PLCH1, DPP6, COC, HLFNA5, GRR, AVPR1A, ADGRL3, EPHA6, RGS7, AQP8, WNT6, FABP4, PSG1A, NO1, SOX6, NDP, PREX2, TFAP2C, NTRK1, CNTN1, SYT9, FGF10, RAMP3, PLA2G7, MMP12, FAIM2, SEMA6A, ADCY1, COL14A1, LCP1, PIP5K1B, ZNF726, ADRA1DSNCA, CLIC2, STAR, ACP3, IL3, IRA, APOD, INSRR, LAG3, SOX18, FZRL2, CCR1, ICA1, NKAIN1, RETREG1, ISM1, ATCAY, PREX1, CARD16, ADRA2C, FLT1, FABP5, PRDM1, TRPC3, TUNAR, DKK2, PROK1, SCTR, WNT16, ZNF804A, RNF157, SH3GL2, PRKCB, DHCR24, GPR20, PDE3B, CHST2, BMP4, IL1RL1, HS3ST5, PLCG2, KCNHS, PRL, FOXQ1, SEMA6D, NEFL, WNT5A, MACC1, TRPC6, MEOX1, DOCK4, HTR1B, PRAME, TIMP3, EGF16, SERPINB2, FRZB, PSG3, VWC2, CF, D, CRACD, RAB6C, NLRP2, CPE, ADAMTS9, GPBAR1, DNAJC6, NPW, XK, EREG, GABRA2, NDNF, DCHS1, CNTN6, MEGF10, SNAP25, ELMO1, SYT1, NETO1, CD24, ZBTB7C, LHCGR, CLSTN2, SCD, DTX1, PDGF, ZNF608, RASL11B, WFDC1, MMP7, PIK3CG, EFHB, MMRN1, LDLR, SOAT2, RGCC, CD4, FOXL2, MYZAPD, PF3, GPR183, GPAT3, TRPA1, PTGER4, GUCY1A2, EPHA5, FZRL3, DOC2B, KCNMB4, C1QL1, TOX, ADAMTS5, CST4, RSP03, WNK2, C1ORF115, KRT17, ADGRB3, MAP2K6, ADAMTS1, PDE4B, PRODH, NES, SH2D2A, IL2RB, TH, TMEM119, IRX6, PSG9, PAD12, MUC12, CACNA1D, LRFN5, MMP8, KDR, SHISA9, KRT36, ADRB2, ACKR3, P2RX6, RGL3, OLFM2A, CDHR3, GJA3, MGAT3, ATP6V0D2, TNFRSF9, PTPRC, APOBEC3B, SEMA3D, ANK1, DJRAS2, B4GALNT2, MARK1, PITX1, EYA1, TXK, KL, ST8SIA1, PLAU, EPHA7, ZNF365, PIK3R3, ADGRL4, GJB2, SYN3, HECW2, SOX8, CCND2, ATP1B2, RTN4R, PTGS1, CRABP2, HECW1, PTPRD, ZNF114, MCTP1, MASP1, TOGARAM2, INSIG1, BMP8A, SORCS1, JIMPA2, CHRM2, EPCAM, WNT9A, UCA1, PRXL2A, TRIM55, GRM4, IRX2, SEMA3B, TSPAN12, BATF2, SIGLEC1, GCOM1, MBOAT1, UCN2, ADGRG6, TNFRSF18, CYP2J2, CLEC14A, CYP1A1, RUBCNL, ASTN2, NTN1, P3H2, GRIP2, ATP1A2, STAC2, PDGFD, AFF3, ADGRG1, TNFRSF19, CLDN3, MSTN, TIMC1, ERMPI1, LEPR, FADS1, DMRTA2, RTKN2, CD248, IL15, STK32B, KREMEN1, CACNA1H, BMP2, LPAR4, LAMA5, SYT7, SLC2A8, NTNG1, TCIM, TAS2RI, ZBTB46, PAK3, KRT19, FABP3, ROS1, ATRNL1, NME, APOE, SFMBT2, ACSL5, SIPA1L2, PTCH1, OSR2, ITPKA, RAB7B, RBM20, CASP1, PDE9A, PTPRN2, MGLL, EFNB2, STAMBPL1, FLA-C, CNID1, XCR1, HMGCR, ACE, PPP1A2, ENTREP1, CORT, PGR, WLS, RIMS1, KLIK3, MCO LN2, STMN2, GREM2

Table 2 (continued)

GO ID	CATEGORY	GO Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
GO:0016020	CC	membrane	4.72E-15	14.32631317	295	PLPPR4, PCSK9, GRIA4, CNTN4, APCDD1, TMEM26, GRIN2A, LAMC3, L1CAM, HPSE2, SDK1, ROBO2, LPAR3, GALNT17, PALMD, SLC3F5, CACNA1G, AJAP1, HSD17B2, PCDH19, NLGN4X, VCAM1, STRA6, ST8SIA4, PLCH1, DPP6, EFNA5, AVPR1A, ADGRL3, EPHA6, CNTN5, RGS7, AQP8, WNT6, PSG1, ANO1, ERV1, ERV1-1, PREX2, NTRK1, CNTN1, GALNT13, SYT9, FGF10, RAMP3, FAIM2, SEMA6A, DCY1, MIDGA2, LCP1, MMP27, LRP1B, L1, Y6K, PIP5K1B, ADRA1D, SNCA, ST6GALNAC3, TMEM52B, TMEM37, CLIC2, NAAALAD2, LMTK3, ACP3, IL31RA, ANO4, INSR, LAG3, F2RL2, TMEM132B, CCR1, CZORF88, ICA1, NIKAIN1, RETREG1, CYP39A1, S100A3, CHST1, ATCAY, PREX1, ADRA2C, FLT1, FABP5, TRPC3, TUNAR1, GSF3, THSD7A, SCTR, ZNF804A, SH3GL2, PRKCB, DHCR24, GPR20, PDE3B, CHST2, IL1RL1, MYPN, HSS3T5, PLCG2, KCNH5, ALP, SEMA6D, WNT5A, TRPC6, DOCK4, HTR1B, PRAME, THSD7B, LCO4A1, EGF16, SERPINB2, CYP4B1, COL7A1, FRZB, PSG3, VWC2, LANCL3, CPEGPBAR1, DNAJC6, XK, EREG, GABRA2, TMC3, DCHS1, CNTN6, MEGF10, NAP25, ELMO1, SYT1, NETO1, CD24, SLC26A7, LHCGR, CLSTN2, SCD, SYTL5, PDGFB, LSAMP, PIK3CG, CYR11, LDLR, SOAT2, CD4, MYZAP, AIF1, LGP1, R3, GPAT3, CYP251, TRPA1, PTGER4, EPHA5, F2RL3, DOC2B, KCNMB4, WNK2, CD163, L1, MYO7A, LAMB4, C, TORF115, KRT17, SLC02A1, ADGRB3, APTM2, CACNA2D3, CNNM1, PDE4B, PRODH, IL2RB, TH, TMEM119, PSG9, GSG1, MUC12, CACNA1D, LRFN5, KDR, SHISA9, CNFN, TMEM179, ADRB2, ACKR3, P2RX6, RGL3, CNGA1, CDHR3, GJA3, MGAT3, MSMO1, ATP6V0D2, TNFRSF9, PTPRC, SEMA7, A3D, ANKI, DIRAS2, B4GALNT2, ZNF474, MARK1, TXK, KL, ST8SIA1, PLA1, LEPA7, PIK3R3, GALNT14, ADGRL4, GJB2, SYN3, CCND2, ATP1B2, RTN4R, PTGS1, PTPRD, MCTP1, PCDH9, NALF1, ST6GALNAC2, INSIG1, SORCS1, CHR2, M2H, KDC1, EPCAM, GRM4, TSPAN12, SNTB1, SIGLEC1, SLC37A2, CLDN23, MBOA1, ATP1A2, STAC2, PDGFD, ADGRG1, PNPLA3, TNFRSF19, CCDC188, CLDN3, TMC1, ERMP1, LEPR, FADS1, SDK2, RTKN2, CD248, KREMEN1, CACNA1H, BMP2, PCDHB15, GIMAP2, FAMI169A, LPAR4, SYT7, SLC2A8, NTNG1, TCIM, OGFR1, TAS2R1, PAK3, KRT19, ROS1, ATRNL1, MME, APOE, CUBN, ACSL5, PTCH1, RA, B7B, CASP1, PDE9A, PTPRN2, MGLL, EFN2, GYPE, STAMBPL1, HLA-C, ABCG2, XCR1, HMGCR, ACE, PPIA2, ENTREP1, BFSP1, PGR, WLS, RIMS1, MCOLN2, ST3GAL5, STMN2, ARSL

Table 2 (continued)

GO ID	CATEGORY	GO Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
GO:0005576	CC	extracellular region	1.97E-13	12.7052702	159	PCSK9,ADAMTS19,GRIA4,CNTN4,SEMA3A,LAMC3,CBLN2,HPSE2,CILPR,OB02,S100A4,VCAM1,COL6A6,CLEC3B,MMP3,COL28A1,COCH,GRPCN,TN5,WNT16,FABP4,PSG1,ANO1,NDPERVIER34-1,CNTN1,FGF10,ALDH1L1,PLA2G7,MMP12,ADCY1,MDGA2,COL14A1,LCP1,MMP27,LY6K,SNCA,TMEM52B,ACFP3,APOD,LAG3,F2RL2,CST1,ISM1,FLT1,FABP5,THSD7A,ADAMTS17,DKK2,PROK1,WNT16,COL5A3,FLT3A1,PRKCB,BMP4,IL1RL1,PLCG2,PRL,WNT5A,TIMP3,EGFL6,SERPINB2,COL7A1,FRZB,PSG3,VWC2,CDF,CPE,ADAMTS9,NPW,EREG,NDNFNETO1,LHCGR,COL24A1,PDGFBL,LSAMPWF,DC1,MMP7,MIMRN1,APOL4,LDLR,AIF1L,F2RL3,CTQL1,ADAMTS5,CST4,R,SPO3,CD163L1,LAMB4,ADGRB3,ADAMTS1,PSG9,PADI2,MMP8,KDR,KRT36,OLFML2A,CDHR3,COL21A1,ATP6V0D2,PTPRC,SEMA3D,KL,PLAU,RTN4R,PTGS1,CRABP2,PTPRD,ZNF114,MAASP1,OPRT,BMP8A,EPCAM,WNT9A,PRXL2A,KRT37,SEMA3B,SIGLEC1,SLC37A2,KRT34,UCN2,CRTAC1,TNFRSF18,ACAT2,CYP2J2,H2AC20,NTN1,ATP1A2,PDGFD,ADGRG1,MSTN,PREB3,LEPR,CD248,IL15,GSTT2B,BMP2,PCOLCE,LAMA5,SYT7,NTNG1,NELL2,KRT19,FABP3,MME,POECUBN,HLA-C,NID1,ACE,PPFIA2,CORT,RENBP,WL,S,KLK3,ARSL,GREM2,C1QTNF7
GO:0005102	MF	signaling receptor binding	9.33E-07	6.030222484	71	PCSK9,SEMA3A,S100A4,NLGN4X,VCAM1,EFNA5,GRP,AVPR1A,WNT16,FABP4,ANO1,NDPNTRK1,FGF10,SEMA6A,LCP1,LAG3,F2RL2,ADRA2,CK2,P,ROK1,WNT16,BMP4,PRL,SEMA6D,WNT5A,WACC1,DOCK4,EGFL6,CPE,NIP,WEREG,MEGF10,SYT1,NETO1,DTX1,SYTL5,PDGFBRASL11B,PIK3CG,CD4,CTQL1,ADAMTS5,RSPO3,NES,KDR,PTPRC,SEMA3D,KL,EPHA7,PTPRD,BMP8A,WNT9A,SEMA3B,UCN2,GRIIP2,PDGFD,MSTN,IL15,BMP2,LAMA5,TCI,M,ATRNL1,APOE,PTCH1,EFNB2,HLA-C,ACE,CORT,PGR,GREM2
GO:0060089	MF	molecular transducer activity	4.32026E-05	4.3644490249	64	GRIA4,GRIN2A,L1CAM,ROBO2,LPAR3,RORB,NLGN4X,STRA6,AVPR1A,ADGRL3,EPHA6,NTRK1,RAMP3,ADRA1D,IL131RA,INSRR,LAG3,F2RL2,CCR1,ADRA2C,FLT1,SCTR,GPR20,IL1RL1,HTRI1B,GPBAR1,GABRA2,LHCGR,CD4G,PR183,PTGER4,EPHA5,F2RL3,ADGRB3,IL2RB,KDR,ADRB2,ACKR3,P2RX6,TNFRSF9,PTPRC,EPHA7,ADGRL4,RTN4R,PTPRD,SORCS1,CHRM2,GRM4,ADGRG6,TNFRSF18,ADGRG1,TNFRSF19,CLDN3,LEPR,LPAR4,OGFRL1,TA5ZRT1,ROSI,CUBN,PTCH1,PTPRN2,NID1,XCR1,PGR

Downregulated genes

Table 2 (continued)

GO ID	CATEGORY	GO Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
GO:0032502	BP	developmental process	1.56E-20	19.80631915	232	PTGIS,SFRP4,MFAP5,BMP6,CDH6,TSPAN2,PKP2,MDF1,EMB,PDLLIM3,COL11A1,GDNF,CHRD12,FOX11,PLPP4,LMO3,CABLES1,INFASC,FGFR2,TIME M215,IL33,NGF,PAPPA2,TENM2,TBX18,HAPLN1,GDF6,FOXC2,COMPCE CR2,FSTSORBS2,CACNG7,TCF23,FGF14,ITGA7,PLN,LEFEMP1,GATA6,CXADR,BHLHE41,ACAN,CSGALNACT1,TCF21,PTGS2,MTSS1,LRR32,GDF10,CPNE5,DHRS3,DACT1,SPINT2,NKX3-1,HOXC8,DGAT2,PITX2,AKR1C3,TNFSF10,BDNF,KRT7,ADAMTSL1, RGS4,SMIM43,RAPGEF5,NDRG2,SCG2,EPHA3,PLA2G5EYA2,INHBA,ALPL,MECOM,SGK1,CSTA,ATP10A,RAP1 GAP,HOXC9,PLEKHB1,SLC2A12,COL4A4,MYH3,DIO3,EPB41L3,TLR3,ANGPTL4,EPHB2,FLI1,THBS1,MBPID3,TNNT2,NOX4,LHX9,S1PR5,SERPINE2,SHANK2,IGSF9,PI16,CRLF1,EN1,PCSK1,TGFB1,IGF2BP1,SALL4,EYA4,KRT18,ITGB1BP2,WNK4,KRT13,DLX5,VLDLR,VDR,GPC4,FAM20A,FOX51,LZTS1,RARB,FZD7,HOXB6,TPRN,ZFH2,CELSR2,ORS1E2,OMG,EPHA4,EN2,DAWI,CDH8,ENC1,IFNE,TNNT1,ANKRD1,SGCA,ZFPM2,SOX9,MGP,SULF1,CYP24A1,DNAH11,ID2,TLR2,IL6,NPPB,SPINK1,GAS6,GPC3,NTRK3,CCNA1,AMIGO2,F11R,SIGLEC15,FOXD1,CCNO,ADAM12,RASSF2,NCAM1,LGI2,MYOC,HOXB7,DUSP2,USP2,KLF2,SIK1,SIX1,FGF7,CCN2,JCAD,EXP H5,DAB1,PKDCC,MYH10,IGFBP3,ARHGDI,HRNR,SLC9C1,L,YVE1,PRKD1,TTTC9,CHL1,BGN,EPPK1,EDA,MYEF2,ACTBL2,NTRK2,SLIT2,IRX3,COL8A2,ACTC1,FLG,HEG1,NRP2,NNMT,CHI3L1,TNIK,SP6,EXTL1,RUNX1,NFIB,PLN,STOX2,PLP1,MYOM2,NAV2,MYOZ1,COL4A3,KCNO4,PPP1R13L,SEMA3C,DCLK2,NOTCH2,PTGDR,PALM2,AKAP2,PGFTHB52,ANK3,DES,KRT86,PDLLIM5,PRDM6,TRABD2A,HTR6,NAV1,HSPB7,NPAS1,PNPLA1,NRGI1,DMRT2,LIF,PLK2,PALM,CDKL2

Table 2 (continued)

GO ID	CATEGORY	GO Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
GO:0065007	BP	biological regulation	3.68E-14	13.43418786	342	PTGIS,SFRP4,BMP6,CDH6,PDE1C,GPC6,PIEZO2,PKP2,RARRS1,MDF1N,ALCN,RRGG,GNFC,HRDL2,FOXJ1,PLPP4,FIBIN,LMO3,CABLE1,ITGBL1,FGFR2,KCNJ12,NPY4R,TMEM215,IL33,PLCB4,NGF,PAPPA2,CD200,BST2,ITENM2,ARHGEEF4,ITX18,MN1,FAM110C,HAPLN1,GDF6,NPY4R2,FOX2,ERG,COMPST,FMO2,AK4,SORBS2,CACNG7,TPD5ZL1,TCF23,FGF14J1,TGA7,PLN,EFEMP1,GATA6,CXADR,BHLHE41,ACAN,LRRN2,TCF21,CACN6,G6,PTGS2,MTSS1,LRR32,APCDD1L,GDF10,CPN5,RASSF9,TNFAIP8L3,DHRS3,DACT1,SPINT2,NKX3-1,CMKLR2,CHN2,HOXC8,DGAT2,PTX2,SIC2A5,AKR1C3,TNFSF10,BDNFRGS4,KCNE4,SMIM43,RAPGEF5,NDRG2,SCG2,MT1FEPA3,PLAZG5,EYA2,GMNC,KCNJ2,INHBA,ALPL,C10ORF90M,ECOM,SGK1,CSTA,ATP10A,RAP1GAPULBP1,HOXC9,PLEKHBI,VEPH1,RG57B,TFPI2,DIO3,DYSE,EPB41L3,TLR3,ANGPTL1,PPP1R12B,VSTM2L,CELF2,ANGPTL4,SLC1A7,EPHB2,ZBED9,FLI1,RASSF5,THBS1,GCH1,MBP,PPP1R14A,NXPH4,ID3,TNNT2,LIMCH1,NOX4,CGREF1,ILHX9,S1PR5,NXPH3,CACNG8,PLAZG4A,SERPINE2,OR2W3,RAPGEF4,TRH,ADAP1,SHANK2,IGSF9,PI16,TLR1,STXBP6,CRLF1,EN1,RSPO1,PCSK1,CALB2,TGFB,IGFBP1,SALL4,EYA4,KRT18,ITGB1BP2,CCDC80,DAPK1,WNK4,KRT13,DLX5,VLDLR,VDRX2,CELSR2,OR51E2,EPHA4,EN2,LRR38,PODN,SCN4B,FMO1,SORCS2,CDH8,ENCI,JFNE,TNNT1,ANKRD1,SGCA,ZFPM2,SOX9,MGPSULF1,IL20RA,DEPPI,CYP24A1,BACH2,DNAH11,PKA,ID2,MCTP2,TLR2,IL6,NPPB,FRMD5,SPINK1,GAS6,GPC3,NTRK3,FBXO32,AMIGO2,F11R,HPGD,MP2,RAB11FIP1,SIGLEC15,FOXD1,ADAM12,RASSF2,ALDH8A1,LGAL52,HAS3,NCA M1,MYOC,ZFH4,HAS1,GCSAM,HOXB7,DUSP2,USP2,RAB31P,KLF2,SIK1,EEF1A2,OR2A7,SIX1,FGF7,CCN2,ICAD,MAP3K9,SUPT20HL1,ABCA8,EXP H5,DAB1,PKDCC,PLPPR3,MYH10,FRMPD4,IGFBP3,FANK1,MP7,ARHGDIG,TRIM63,SLC4A4,SH3RF2,SLC9C1,HUNK,CYR1A,LYVE1,TMEM38A,BEN D5,SLAIN1,PRKD1,CYP3A7,CHL1,KCNG2,EPPK1,SIRPB1,EDA,PARP15,CP EB1,NTRK2,SYT15,HSD17B6,FPR3,SLIT2,IRX3,MPO,ACTC1,HEG1,NRP2,N NMT,CHI3L1,TNIK,LMCD1,SP6,RUNX1,NFIB,APLN,SORBS1,EPB41,PDGFR L,ACKR2,FPR1,STOX2,PLP1,NAV2,PDE7B,INSYNA,MYOZ1,PMEL,CO14A 3,PPP1R13L,SEMA3,CBMAL2,ZNF334,DCLK2,NOTCH2,JOSEC3,JPH3,SIC 38A3,PTGDR,PALM2,AKAP2,PKK4,GNG7,PGF,THBS2,GPR143,LRRC2,ANK 3,DES,PDLM5,PRDM6,IGFBP11,TRABD2A,HTR6,HSPB7,NPAS1,NFIX,PIK3 AP1,NRG1,DMRT2,LIF,ADH1C,SIRPA,NUAK1,CHRM4,CACNA1I,PLK2,STX 11,PALM,CDKL2

Table 2 (continued)

GO ID	CATEGORY	GO Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
GO:0071944	CC	cell periphery	1.05E-11	10.97687663	204	PTGIS,MFAP5,CDH6,TSPAN2,GPC6,PIEZO2,PKP2,NALCN,EMB,MGAM,CO L11A1,RRERG,PLPP4,ITGBL1,NFASC,FGFR2,ANXA8L1,KCNJ12,NPY4R,PAPP A2,CD200,BST2,TENM2,ARHGEF4,FAM110C,HAPLN1,NPY4R2,COMP,SO RBS2,CACNG7,ITGA7,EFEMP1,CXADR,ACAN,LRRN2,CACNG6,PTGS2,LRR C32,APCDD1L,GDF10,CPNE5,TNFAIP8L3,KCNIN3,DHRS3,HMCN1,SPINT2 ,CMKLR2,SLC2A5,TNFSF10,ADAMTSL1,RGS4,KCNE4,SMIM43,SBSPO,NR APGEF5,EPHA3,PLA2G5,PCDHA12,KCNJ2,EVA1A,ALPL,C10ORF90,SGK1 ,CSTA,ATP10A,UlBP1,VEPH1,SLC2A12,RGS7BPCOL4A4,TFPI2,DIO3,DYSF ,EPB41L3,TLR3,ANGPTL1,ANGPTL4,SLC1A7,EPHB2,THBS1,MBPMFAP3L, NOX4,S1PR5,CACNG8,SERPINE2,OR2W3,RAPGEF4,TINAGL1,SYTL2,ADA P1,SHANK2,IGSF9,TLR1,CRLF1,TGFB1,KRT18,CCDC80,DAPK1,VLDLR,GPC 4,LZTS1,KISS1,GEM,FZD7,PTPRN,CELSR2,OR51E2,OMG,EPHA4,LRRRC38,P ODN,SCN4B,SORCS2,CDH8,SGCA,SLC22A23,MGPSULF1,IL20RA,STEAP1 B,TLR2,IL6,GPC3,NTRK3,SLC6A15,AMIGO2,F11R,MPP2,SIGLEC15,ADAM 12,RASSF2,HAS3,NCAM1,MYOCH,HAS1,GCSAM,OR2A7,ESYT3,CCN2,ICA D,ABCA8,MYH10,JFITM10,MPP7,GRAMD1B,ARHGDIG,SLC4A4,HRNR,SLC 9C1,LYVE1,PRKD1,CHL1,KCNG2,BGN,EPPK1,SIRPB1,EDA,STEAP2,NTRK2, SYT15,FPR3,COL8A2,FLG,HEG1,NRP2,CHI3L1,TNIK,RHD,STEAP1,SORBS1 ,EPB41,ART4,ACKR2,MGST1,FPRI,PLP1,NAV2,PMEL,COL4A3,KCNQ4,SSP N,NOTCH2,JOSEC3,JPH3,SLC38A3,PTGDR,PALM2AKAP2,GNG7,THBS2,G PRI43,ANK3,DES,IGFBP11,TRABD2A,HTR6,PIK3AP1,NRGT,ADH1C,SIRPA, CHRM4,CACNA1L,STX11,PALM

Table 2 (continued)

GO ID	CATEGORY	GO Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
GO:0005737	CC	cytoplasm	0.000756935	3.120941328	297	PTGIS,SFRP4,PDE1C,GPC6,PKP2,MDF1,PDLIM3,MGAM,COL11A1,RERGG,DNF,CHRD,L2,FININ,LMO3,CABLES1,INFASC,FGFR2,ANXA8L1,GALNT12,IL33,GAL3ST3,PLCB4,NGFBST2,TENM2,ARHGEF4,FAM110C,ERG,FST,FMO2,AK4,SORBS2,CACNG7,TPD52L1,FGF14,PLN,CXADRACAN,MVH1,CSGA,LNACT1,PTGS2,MTSS1,RASSF9,TNFAIP8L3,KCNJ3,DHRS3,HMNC1,DACT1,SPINT2,NKX3-1,CHN2,DGAT2,SLC2A5,AKR1C3,SCGB3A2,BDNF,KRT7,A1,DAMTSL1,RGS4,NDRG2,IGFN1,THEM5,SCG2,MT1F,EPHA3,PLA2G5,OLA,H,AYA2,EVAT1A,INHBA,ALP,CTOORF90,LXN,MECOM,SGK1,CSTA,ATP10A,RAP1GAP,ULBP1,PLEKH81,SLCZA12,RGS7BP,COL4A4,MYH3,DIO3,DYSF,EPB41L3,TLR3,SPTLC3,PPP1R12B,STM2L,CEL2,MARCHF3,EPH82,ZBED9,FLI1,RASSF5,THBS1,GCH1,MBPPP1R14A,MFAP3L,ID3,TNNT2,LIMCH1,NOX4,IPR5,CACNG8,PLA2G4A,SERPINE2,RAPGEF4,ALDH1B1,PRUNE2,TRH,TINAGL1,SYTL2,ADAP1,SHANK2,TLR1,STXBP6,CRLF1,PCSK1,CALB2,TFGBI,IGF2BP1,SALL4,EYA4,KLHL4,KRT18,ITGB1BP2,DAPK1,WNK4,KRT13,DLX5,FMO3,VLDLR,VDR,GPC4,FAM20A,LZTS1,RARB,FZD7,PTPRN,CELSR2,OR51E2,TLCD4,EPHA4,PODIN,FMO1,DAW1,SORCS2,ENC1,TNNT1,AN,KRD1,SGCA,ZFPM2,SULF1,PIPOX,DEPP1,CYP24A1,BACH2,DNAH11,PKIA,ID2,MCTP2,WSCD1,STEAP1B,TLR2,ARHGEF37,IL6,NPPB,GAS6,GPC3,FBXO32,CCNA1,HPGDS,MPP2,RAB11FIP1,CCNO,BEGAIN,RASSF2,CCDC110,ALDH8A1,SEL1L2,HAS3,NCAM1,MYOC,PPFIA4,HAS1,GCSAM,HOXB7,GA,LNT9,DUSP2,USP2,RAB31P,SIK1,EEF1A2,SIX1,FGF7,ESYT3,CCN2,JCAD,MYOZ3,ABCA8,EXPH5,DAB1,PKDCC,MYH10,IGFBP3,FANK1,MPP7,GRAMD1,B,ARHGDI,TRIM63,HRNR,HUNK,TMEM38A,BENDS,SLAIN1,PRKD1,CYP3A7,BGN,EPPK1,SIRPB1,EDA,STEAP2,PARP15,ACTBL2,CPEB1,NTRK2,SYT15,HSD17B6,SLIT2,IRX3,MPO,COL8A2,ACTC1,FLG,B3GALT5,NNMT,CH13L1,TNIK,LIMCD1,STEAP1,SP6,EXTL1,SORBS1,EPB41,ACKR2,MGST1,FPR1,STOX2,ST3GAL6,MYOM2,PDE7B,MYOZ1,PMEL,COL4A3,SSPN,PPP1R13L,B,MAL2,DCLK2,NOTCH2,DSE,ZNF185,IOSEC3,P4HA3,JP43,SECI4L6,PDK4,PITPNM3,THBS2,DBNDD1,GPR143,GSTO2,DNAH6,ANK3,DES,KRT86,PD,LIM5,NAV1,HSPB7,PNPLA1,PIK3AP1,LIF,MCUB,ADH1C,SIRPA,B3GNT7,NUAK1,PLK2,STX11,PALM,SPSB1,PKP,CDKL2,CTAGE9
GO:0098772	MF	molecular function regulator activity	7.91485E-05	4.101557094	80	BMP6,PKP2,RARRES1,MDF1,RERGG,GDNF,IL33,NGFBST2,ARHGEF4,GDF6,FST,CACNG7,TCF23,FGF14,PLN,EEFEMP1,CACNG6,GDF10,SPINT2,NKX3-1,CHN2,TNFSF10,BDNF,RGS4,KCNE4,RAPGEF5,SCG2,INHBA,LXN,ISGK1,CSTA,RAP1GAP,TFPI2,PPP1R12B,ANGPTL4,THBS1,PPP1R14A,JD3,CACN,G8,SERPINE2,RAPGEF4,TRH,ADAP1,PII16,CRLF1,WNK4,FAM20A,GEM1L,RRC38,SCN4B,IFNE,PKIA,ID2,ARHGEF37,IL6,NPPB,SPINK1,GAS6,GPC3,CCNA1,CCNO,RAB31P,FGF7,IGFBP3,ARHGDI,SH3RF2,PRKD1,EDA,SLIT2,APLN,MYOZ1,COL4A3,SEMA3C,IOSEC3,PGF,IRG1,LIF,MCUB,SIRPA
GO:0005509	MF	calcium ion binding	0.000143688	3.842578472	38	CDH6,ANXA8L1,PLCB4,TENM2,COMP,EEEMP1,ACAN,HMNC1,MEGF6,PLA2G5,PCDHA12,ALPL,DYSF,THBS1,GCH1,CGREF1,PLA2G4A,CALB2,ITGB1BP2,VLDLR,CELSR2,CDH8,SCGAM,GP,SULF1,MCTP2,GAS6,HPGDS,ESYT3,HRNR,SYT15,SLIT2,FLG,HEG1,RUNX1,NOTCH2,PITPNM3,THBS2

Table 3 The enriched pathway terms of the up- and downregulated differentially expressed genes

Pathway ID	Pathway Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
<i>Upregulated genes</i>					
REAC:R-HSA-372790	Signaling by GPCR	0.000169138	3.771759438	44	PLPPR4,LPAR3,MMP3,AVPR1A,RGS7,WNT6,RAMP3,ADCY1,ADRA1D,F2RL2,CCR1,PREX1,ADRA2C,TRPC3,PROK1,SCTR,WNT16,PRKCB,GPR20,PDE3B,WNT5A,TRPC6,HTR1B,GPBAR1,NPW,XK,LHCGR,PIK3CG,GPR183,PTGER4,F2RL3,ADRB2,ACKR3,PIK3R3,CHRM2,WNT9A,GRM4,UCN2,LPAR4,TAS2R1,PTCH1,MGLL,XCR1,CORT
REAC:R-HSA-1474244	Extracellular matrix organization	0.000724319	3.140069834	24	LAMC3,VCAM1,COL6A6,MMP3,COL28A1,MMP12,COL14A1,COL5A3,BMP4,COL7A1,ADAMTS9,COL24A1,PDGFB,MMP7,ADAMTS5,ADAMTS1,MMP8,KDR,COL21A1,P3H2,BMP2,PCOLCE,LAMA5,NID1
REAC:R-HSA-373076	Class A/1 (Rhodopsin-like receptors)	0.011143325	1.952985185	22	PLPPR4,LPAR3,AVPR1A,ADRA1D,F2RL2,CCR1,ADRA2C,PROK1,HTR1B,GPBAR1,NPW,XK,LHCGR,GPR183,PTGER4,F2RL3,ADRB2,ACKR3,CHRM2,LPAR4,XCR1,CORT
REAC:R-HSA-9675108	Nervous system development	0.017667803	1.752817451	31	SEMA3A,L1CAM,ROBO2,CACNA1G,COL6A6,ST8SIA4,EFNA5,EPHA6,TUBA3E,CNTN1,SEMA6A,TRPC3,COL5A3,SH3GL2,SEMA6D,TRPC6,CNTN6,CD24,EPHA5,CACNA1D,PTPRC,ANK1,EPHA7,PIK3R3,ADGRG6,NTN1,CACNA1H,NELL2,PAK3,EFNB2,HMGCR
REAC:R-HSA-162582	Signal Transduction	0.033015787	1.481278352	94	PLPPR4,LAMC3,CILP,LPAR3,COL6A6,MMP3,AVPR1A,TUBA3E,RGS7,WNT6,SOX6,PREX2,NTRK1,CNTN1,FGF10,RAMP3,ADCY1,PIP5K1B,ADRA1D,APOD,F2RL2,CCR1,PREX1,ADRA2C,FLT1,FABP5,PRDM1,TRPC3,DKK2,PROK1,SCTR,WNT16,COL5A3,SH3GL2,PRKCB,GPR20,PDE3B,IL1RL1,PLCG2,NEFL,WNT5A,TRPC6,DOCK4,HTR1B,GPBAR1,NPW,XK,EREG,ELMO1,LHCGR,SCD,DTX1,COL24A1,PDGFB,MM P7,PIK3CG,GPR183,PTGER4,F2RL3,RSP03,SH2D2A,IL2RB,KDR,ADRB2,ACKR3,RGL3,ATP6V0D2,PIK3R3,RTN4R,CRABP2,HECW1,CHRM2,WNT9A,GRM4,UCN2,H2AC20,PDGFD,KREMEN1,BMP2,FAM169A,LPAR4,LAMA5,TAS2R1,PAK3,APOE,PTCH1,MGLL,XCR1,CORT,PGR,WLS,KLK3,STMN2,GREM2
REAC:R-HSA-109582	Hemostasis	0.053792682	1.269276802	32	KIF26A,L1CAM,CLEC3B,TUBA3E,PSG1,F2RL2,ADRA2C,TRPC3,F13A1,PRKCB,PLCG2,TRPC6,DOCK4,TIMP3,SERPINB2,PSG3,CFD,PDGFB,PIK3CG,MMRN1,GUCY1A2,F2RL3,KCNMB4,PSG9,P2RX6,PLAU,PIK3R3,ATP1B2,EPCAM,VPREB3,PDE9A,MGLL
<i>Downregulated genes</i>					
REAC:R-HSA-397014	Muscle contraction	0.005754659	2.23998037	17	KCNJ12,CACNG7,FGF14,PLN,CACNG6,KCNE4,KCNJ2,MYH3,DYSF,TNNT2,CACNG8,SCN4B,TNNT1,ACTC1,SORBS1,DES,CACNA1I
REAC:R-HSA-1630316	Glycosaminoglycan metabolism	0.01248541	1.903597196	12	GPC6,ACAN,CSGALNACT1,GPC4,GPC3,HAS3,HAS1,LYVE1,BGN,ST3GAL6,DSE,B3GNT7

Table 3 (continued)

Pathway ID	Pathway Name	adjusted_p_value	negative_log10_of_adjusted_p_value	Gene Count	Gene
REAC:R-HSA-1474244	Extracellular matrix organization	0.051293235	1.289939908	18	MFAP5,COL11A1,HAPLN1,COMP,ITGA7,EFEMP1,ACAN,COL4A4,THBS1,CTRB2,F11R,ADAM12,CTRB1,NCAM1,BGN,COL8A2,COL4A3,P4HA3
REAC:R-HSA-3000178	ECM proteoglycans	0.051461154	1.288520482	8	HAPLN1,COMP,ITGA7,ACAN,COL4A4,NCAM1,BGN,COL4A3
REAC:R-HSA-5362517	Signaling by Retinoic Acid	0.051461154	1.288520482	6	DHRS3,AKR1C3,RARB,ALDH8A1,PDK4,ADH1C
REAC:R-HSA-3781865	Diseases of glycosylation	0.051461154	1.288520482	11	GPC6,GALNT12,ACAN,ADAMTSL1,BSPPON,THBS1,GPC4,GPC3,BGN,NOTCH2,THBS2

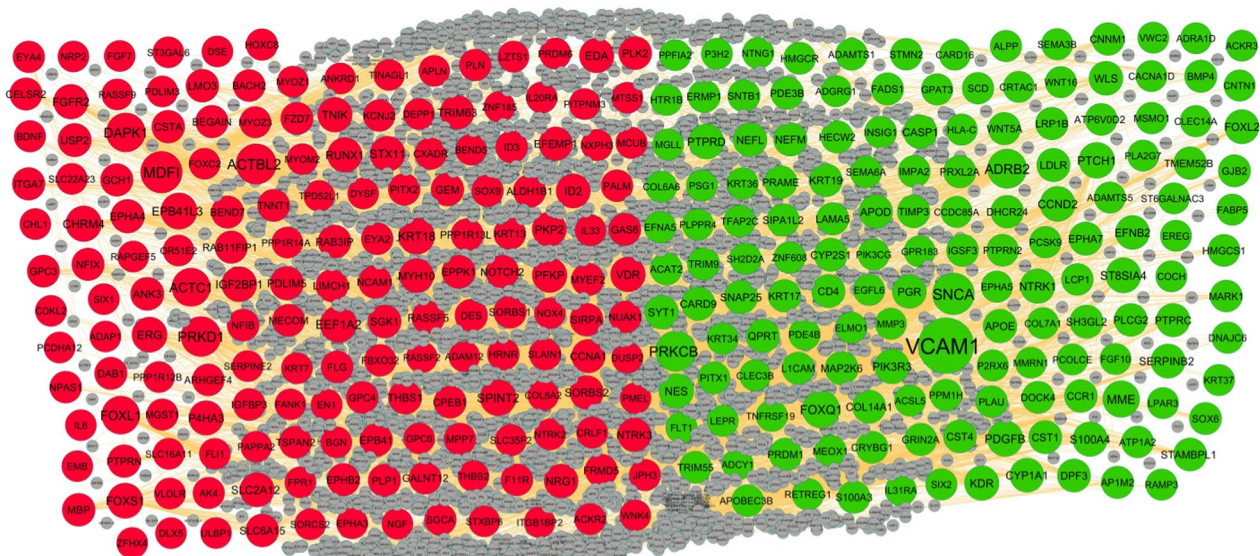


Fig. 3 PPI network of DEGs. Upregulated genes are marked in parrot green; downregulated genes are marked in red

identified the 958 DEGs. The results showed that there were dense regions in PPI, that is, genes closely related to endometriosis. A total of 4871 nodes and 8009 edges were selected to plot the PPI network (Fig. 3). The Network Analyzer plugin of Cytoscape was used to score each node gene by four selected algorithms, including node degree, betweenness, stress and closeness. Finally, we identified ten hub genes (*vcam1*, *snca*, *prkcb*, *adrb2*, *foxq1*, *mdfi*, *actbl2*, *prkd1*, *dap1* and *actc1*) and are listed in Table 4. The top two significant modules from PEWCC were selected for future analysis. Module 1 included 22 nodes and 41 edges (Fig. 4A). The functional enrichment analysis of genes in module 1 was conducted by g:Profiler. These genes were significantly enriched in multicellular organismal process and regulation of biological process. Module 2 included 8 nodes and 14 edges (Fig. 4B). The functional enrichment analysis of genes in module 2 was conducted by g:Profiler. These genes were significantly

enriched in developmental process and biological regulation.

Construction of the miRNA-hub gene regulatory network

We searched for target-regulated hub gene miRNAs using miRNet database and then used the results of this database. By constructing miRNA-hub gene regulatory network networks, we found 2495 nodes (miRNA: 2168; Hub Gene: 327) and 14,692 edges (Fig. 5). We identified 365 miRNAs (ex; *hsa-mir-3143*) targeting regulation of *ccnd2*, 102 miRNAs (ex; *hsa-mir-6888-5p*) targeting regulation of *vcam1*, 89 miRNAs (ex; *hsa-mir-200a-3p*) targeting regulation of *ptprd*, 88 miRNAs (ex; *hsa-mir-3122*) targeting regulation of *pdgfb*, 81 miRNAs (ex; *hsa-mir-17-5p*) targeting regulation of *prkcb*, 241 miRNAs (ex; *hsa-mir-2110*) targeting regulation of *igf2bp1*, 77 miRNAs (ex; *hsa-mir-4432*) targeting regulation of *actc1*, 53 miRNAs

Table 4 Topology table for up- and downregulated genes

Regulation	Node	Degree	Betweenness	Stress	Closeness
Up	VCAM1	416	0.184904	86,834,622	0.341594
Up	SNCA	180	0.086564	32,935,316	0.326731
Up	PRKCB	156	0.05443	19,551,102	0.310589
Up	ADRB2	136	0.056876	10,503,780	0.316941
Up	FOXQ1	102	0.022891	20,702,630	0.264141
Up	MME	93	0.029423	14,457,318	0.274745
Up	PTCH1	78	0.028409	7,457,792	0.26486
Up	PDGFB	76	0.022264	6,008,890	0.261024
Up	ST8SIA4	67	0.023812	5,175,380	0.271404
Up	CCND2	64	0.020133	4,021,362	0.290612
Up	PTPRD	63	0.019057	5,697,616	0.263326
Up	NES	62	0.016543	4,851,666	0.279575
Up	S100A4	58	0.018352	5,689,768	0.278232
Up	PIK3R3	58	0.019257	6,819,154	0.280639
Up	NTRK1	56	0.013319	5,744,746	0.272529
Up	KDR	56	0.011496	7,778,812	0.275476
Up	SERPINB2	52	0.014458	5,702,426	0.255581
Up	FOXL2	51	0.009024	7,152,922	0.25159
Up	CYP1A1	48	0.01608	2,790,170	0.277882
Up	EFNB2	47	0.013517	2,651,168	0.267657
Up	WLS	47	0.013043	3,634,828	0.24982
Up	PGR	46	0.011767	3,597,166	0.275976
Up	SYT1	43	0.008774	2,229,078	0.26394
Up	CASP1	43	0.013201	3,495,008	0.247848
Up	NEFL	43	0.010016	1,997,222	0.276493
Up	SNAP25	42	0.014293	1,986,420	0.284695
Up	APOD	42	0.014537	3,155,838	0.267201
Up	APOE	41	0.012455	1,705,636	0.280963
Up	PTPRC	40	0.010061	2,818,146	0.2708
Up	CARD9	40	0.011367	3,316,374	0.241309
Up	NEFM	40	0.01491	2,636,816	0.293522
Up	QPRT	38	0.009819	8,934,012	0.225947
Up	SH3GL2	37	0.008529	3,627,714	0.25839
Up	EPHA7	37	0.010685	3,258,698	0.263269
Up	CD4	36	0.010006	1,469,630	0.281337
Up	LDLR	36	0.009886	1,569,128	0.256754
Up	MAP2K6	36	0.008317	1,723,230	0.264672
Up	CYP2S1	35	0.010486	1,835,320	0.267966
Up	DPF3	34	0.006119	3,159,488	0.241034
Up	COL14A1	34	0.008718	4,017,460	0.243861
Up	STAMBPL1	33	0.007553	4,960,736	0.233437
Up	CST1	33	0.007158	2,170,272	0.231714
Up	CST4	32	0.006271	2,740,472	0.226906
Up	PLCG2	31	0.008121	1,680,216	0.275476
Up	KRT19	31	0.008239	1,459,230	0.2835
Up	CCR1	29	0.009532	3,308,298	0.246917
Up	PRAME	27	0.006029	2,712,304	0.235482
Up	TMEM52B	27	0.008366	1,251,782	0.265394
Up	DHCR24	26	0.008117	1,620,290	0.284645

Table 4 (continued)

Regulation	Node	Degree	Betweenness	Stress	Closeness
Up	TIMP3	25	0.007343	1,887,972	0.250437
Up	CNNM1	25	0.005697	1,036,690	0.225111
Up	KRT17	25	0.004783	2,178,508	0.269854
Up	PDE3B	25	0.006554	2,226,676	0.246031
Up	PRDM1	24	0.004603	2,157,456	0.257883
Up	S100A3	23	0.005112	2,081,396	0.233628
Up	LAMA5	22	0.004812	1,668,084	0.252819
Up	SIPA1L2	22	0.004587	2,681,374	0.250592
Up	LRP1B	22	0.004855	1,903,866	0.242572
Up	PRXL2A	22	0.005952	857,154	0.244756
Up	AP1M2	21	0.005745	1,327,148	0.25295
Up	BMP4	21	0.005918	1,465,568	0.211152
Up	ERMP1	21	0.005252	1,833,162	0.242307
Up	WNT5A	21	0.005621	839,274	0.243361
Up	GRIN2A	20	0.004087	2,097,912	0.237981
Up	SNTB1	19	0.004951	1,366,392	0.21671
Up	DOCK4	19	0.004269	1,553,902	0.262445
Up	HECW2	19	0.004944	1,653,624	0.23112
Up	FADS1	19	0.003917	1,034,182	0.223786
Up	TRIM9	18	0.003815	745,666	0.244707
Up	SOX6	18	0.003932	1,523,690	0.236042
Up	GJB2	18	0.00357	1,889,844	0.220141
Up	L1CAM	17	0.002057	874,050	0.249079
Up	HMGCR	17	0.003664	1,039,586	0.22993
Up	ACAT2	17	0.003431	920,074	0.241214
Up	MARK1	16	0.002652	1,684,916	0.259313
Up	TRIM55	16	0.001206	524,732	0.240036
Up	HMGCS1	16	0.003905	954,734	0.219377
Up	INSIG1	15	0.003029	1,038,904	0.223047
Up	PITX1	15	0.003043	1,063,398	0.219288
Up	CRYBG1	15	0.002626	695,160	0.262106
Up	IGSF3	15	0.00201	920,740	0.242717
Up	IMPA2	15	0.00307	1,138,144	0.227266
Up	MEOX1	14	0.003339	829,668	0.222802
Up	ATP6V0D2	14	0.002463	486,050	0.221524
Up	TFAP2C	13	0.002502	1,073,038	0.244769
Up	PTPRN2	13	0.002218	428,790	0.226631
Up	ATP1A2	12	0.002637	1,131,342	0.233035
Up	ADAMTS1	12	0.0024	617,034	0.225842
Up	SEMA6A	12	0.003041	437,668	0.233437
Up	APOBEC3B	12	0.001441	744,566	0.221746
Up	PLAU	11	0.003237	803,628	0.243617
Up	LCP1	11	0.002165	543,558	0.22579
Up	EFNA5	10	8.84E-04	151,924	0.243922
Up	RETREG1	10	0.001241	470,002	0.215797
Up	STMN2	10	0.002634	545,182	0.208403
Up	PPM1H	9	0.001276	937,346	0.238098
Up	HTR1B	9	0.00207	271,180	0.241681
Up	MSMO1	8	0.001453	551,916	0.229042

Table 4 (continued)

Regulation	Node	Degree	Betweenness	Stress	Closeness
Up	KRT36	8	7.68E-04	350,894	0.240059
Up	PCSK9	7	3.29E-04	54,918	0.224446
Up	DNAJC6	7	0.001693	628,446	0.220171
Up	SCD	7	0.001379	435,390	0.231021
Up	KRT34	7	0.003303	1,108,236	0.179504
Up	ALPP	5	0.001644	337,932	0.19351
Up	PDE4B	5	7.08E-04	200,632	0.228311
Up	CLEC14A	5	0.001237	443,264	0.182555
Up	GPAT3	5	8.73E-04	87,548	0.215234
Up	NTNG1	4	0.00111	125,664	0.21288
Up	P2RX6	4	5.27E-04	122,142	0.203863
Up	TNFRSF19	4	9.59E-04	559,034	0.234359
Up	ZNF608	4	8.37E-04	272,030	0.208296
Up	HLA-C	4	4.32E-04	96,322	0.228365
Up	SIX2	3	0.002226	1,176,074	0.232756
Up	RAMP3	3	8.22E-04	758,918	0.155682
Up	FLT1	3	2.74E-04	63,344	0.237412
Up	FABP5	3	4.57E-04	103,188	0.213702
Up	SH2D2A	3	4.10E-04	212,434	0.243678
Up	EGFL6	3	2.04E-04	63,224	0.222151
Up	COL7A1	3	9.87E-04	306,868	0.253622
Up	KRT37	3	8.22E-04	275,598	0.132128
Up	CNTN1	3	4.36E-04	235,384	0.249705
Up	SEMA3B	3	1	6	1
Up	MMP3	2	4.76E-05	19,768	0.225049
Up	GPR183	2	5.02E-05	27,508	0.203684
Up	EPHA5	2	8.99E-06	2564	0.215691
Up	IL31RA	2	3.46E-05	26,030	0.212973
Up	CCDC85A	2	1.60E-05	5982	0.186687
Up	WNT16	2	0	0	0.207099
Up	PSG1	2	2.29E-04	125,530	0.232345
Up	ACSL5	2	1.08E-04	106,606	0.247181
Up	PIK3CG	2	2.70E-04	137,430	0.238506
Up	LEPR	2	2.99E-04	76,112	0.231847
Up	FGF10	1	0	0	0.21491
Up	ELMO1	1	0	0	0.207895
Up	ADRA1D	1	0	0	0.178118
Up	P3H2	1	0	0	0.200313
Up	MMRN1	1	0	0	0.219347
Up	COCH	1	0	0	0.209407
Up	PLA2G7	1	0	0	0.200033
Up	PLPPR4	1	0	0	0.196606
Up	VWC2	1	0	0	0.193656
Up	ST6GALNAC3	1	0	0	0.209407
Up	MGLL	1	0	0	0.240677
Up	CLEC3B	1	0	0	0.242005
Up	CARD16	1	0	0	0.198628
Up	COL6A6	1	0	0	0.195886
Up	EREG	1	0	0	0.209389

Table 4 (continued)

Regulation	Node	Degree	Betweenness	Stress	Closeness
Up	ADGRG1	1	0	0	0.240677
Up	PCOLCE	1	0	0	0.207187
Up	LPAR3	1	0	0	0.165313
Up	ADCY1	1	0	0	0.188751
Up	ACKR3	1	0	0	0.180429
Up	CRTAC1	1	0	0	0.203038
Up	ADAMTS5	1	0	0	0.200288
Up	CACNA1D	1	0	0	0.176234
Up	PPFIA2	1	0	0	0.208448
Down	MDFI	180	0.067201	24,938,630	0.277565
Down	ACTBL2	159	0.059788	16,984,080	0.312604
Down	PRKD1	151	0.049413	25,348,404	0.299078
Down	DAPK1	135	0.043258	14,691,546	0.296454
Down	ACTC1	125	0.043583	13,786,990	0.30907
Down	FOXL1	119	0.031148	26,714,620	0.26612
Down	EEF1A2	96	0.030658	12,573,334	0.292657
Down	EPB41L3	88	0.027315	13,917,190	0.278966
Down	KRT18	86	0.033018	9,650,494	0.319249
Down	SPINT2	85	0.027392	6,835,460	0.266689
Down	CHRM4	78	0.024102	10,167,440	0.263869
Down	FOXS1	72	0.016114	9,987,450	0.261276
Down	ERG	70	0.015816	8,115,936	0.283484
Down	IGF2BP1	68	0.018742	4,590,692	0.29807
Down	STX11	67	0.018202	6,035,790	0.271495
Down	VDR	63	0.017576	4,272,470	0.275569
Down	PFKP	63	0.019322	10,071,716	0.289505
Down	FGFR2	62	0.017882	4,630,474	0.273724
Down	TNIK	61	0.017992	5,615,768	0.280526
Down	EDA	61	0.017336	5,807,424	0.250476
Down	RUNX1	60	0.018065	4,954,702	0.282973
Down	P4HA3	59	0.019144	7,280,554	0.270709
Down	ID2	55	0.016051	4,769,732	0.275678
Down	PKP2	55	0.020937	6,848,742	0.291937
Down	SLC2A12	55	0.015964	7,708,246	0.235084
Down	MYH10	52	0.016668	3,589,202	0.30441
Down	NRG1	50	0.016348	3,547,318	0.264831
Down	TRIM63	48	0.012682	4,517,170	0.262615
Down	SORBS2	45	0.00868	6,589,870	0.267186
Down	EPHA4	43	0.013734	2,446,220	0.275429
Down	NTRK3	42	0.010949	4,470,618	0.254578
Down	CCNA1	42	0.009251	3,433,956	0.256971
Down	PDLIM5	42	0.013999	2,000,086	0.281548
Down	EFEMP1	42	0.014364	2,399,978	0.254751
Down	THBS1	41	0.012427	2,896,698	0.261318
Down	ANK3	40	0.010421	4,772,040	0.248506
Down	USP2	40	0.012325	2,824,276	0.271343
Down	SGK1	38	0.007985	3,779,252	0.269659
Down	EPPK1	38	0.007359	4,225,478	0.276414
Down	SIRPA	35	0.008354	4,887,612	0.273248

Table 4 (continued)

Regulation	Node	Degree	Betweenness	Stress	Closeness
Down	RAB3IP	34	0.008005	2,997,686	0.246805
Down	PLK2	34	0.011158	1,806,358	0.269003
Down	SLC6A15	33	0.00856	4,749,196	0.239752
Down	RASSF5	32	0.007015	3,193,748	0.249705
Down	MBP	32	0.010185	1,574,138	0.275008
Down	NOTCH2	32	0.007274	2,605,336	0.262106
Down	EPB41	32	0.008073	4,217,818	0.277123
Down	GEM	32	0.006616	4,083,892	0.257488
Down	CSTA	31	0.007847	3,030,162	0.254192
Down	LMO3	31	0.009479	1,668,282	0.254485
Down	NCAM1	30	0.009068	1,465,722	0.279977
Down	SORBS1	30	0.004624	2,039,478	0.267878
Down	DES	29	0.005178	1,191,798	0.262814
Down	PTPRN	29	0.00818	1,615,368	0.250039
Down	MYEF2	29	0.006925	2,882,312	0.252124
Down	EPHB2	28	0.005108	1,607,522	0.262885
Down	ALDH1B1	28	0.007171	3,795,748	0.243592
Down	ITGA7	27	0.007364	1,448,790	0.24742
Down	BEGAIN	27	0.005823	1,454,784	0.243849
Down	GCH1	26	0.006139	2,464,430	0.240558
Down	MECOM	25	0.005644	2,581,520	0.241789
Down	NFIX	25	0.004898	1,145,270	0.242102
Down	RAB11FIP1	25	0.005257	2,872,472	0.269749
Down	PLP1	25	0.006069	1,380,662	0.25009
Down	DAB1	24	0.005216	1,464,652	0.258472
Down	LIMCH1	24	0.00482	2,909,102	0.245745
Down	PPP1R13L	23	0.003793	1,790,426	0.243714
Down	GAS6	23	0.004951	1,039,182	0.213496
Down	FRMD5	23	0.008404	1,763,548	0.260297
Down	BEND7	23	0.005734	1,408,600	0.237632
Down	CELSR2	23	0.006178	1,023,294	0.238074
Down	FZD7	23	0.005034	1,869,974	0.232656
Down	GALNT12	22	0.005278	1,572,318	0.221878
Down	CPEB1	22	0.003768	839,108	0.260451
Down	TNNT1	21	0.002906	698,182	0.250257
Down	GPC3	21	0.004711	1,741,574	0.228012
Down	TSPAN2	21	0.0057	972,012	0.248379
Down	IGFBP3	19	0.004017	1,687,726	0.236363
Down	ID3	18	0.001278	690,358	0.212211
Down	NFIB	18	0.006922	1,031,030	0.260982
Down	GPC4	18	0.003629	1,410,500	0.24296
Down	KRT13	18	0.001814	1,014,686	0.248392
Down	FLG	18	0.002769	1,392,370	0.234846
Down	BACH2	17	0.004043	1,105,092	0.231593
Down	NUAK1	17	0.002249	854,382	0.242053
Down	ARHGEF4	17	0.002473	1,190,348	0.242681
Down	HRNR	17	0.002458	1,173,000	0.254698
Down	F11R	16	0.002671	1,064,048	0.24369
Down	MPP7	16	0.003136	1,190,282	0.243495

Table 4 (continued)

Regulation	Node	Degree	Betweenness	Stress	Closeness
Down	PALM	16	0.003087	1,250,900	0.236489
Down	PITX2	15	0.002541	901,710	0.225267
Down	NPAS1	15	0.00459	1,076,534	0.226199
Down	EYA2	14	0.00257	759,552	0.228129
Down	KCNJ2	14	0.002222	782,768	0.215119
Down	FBXO32	13	0.002724	515,166	0.242476
Down	SGCA	13	0.003283	940,578	0.222619
Down	SOX9	13	0.003999	579,538	0.246068
Down	EYA4	12	0.00376	1,740,072	0.201699
Down	PLN	12	0.002418	280,268	0.216142
Down	SLAIN1	12	0.001293	817,642	0.236328
Down	HOXC8	11	0.00227	382,190	0.215539
Down	CXADR	11	0.001515	411,744	0.239941
Down	AK4	10	0.002393	770,268	0.214257
Down	BDNF	9	0.001499	722,196	0.218746
Down	MGST1	9	0.001054	383,496	0.220891
Down	JPH3	9	0.001571	342,404	0.213928
Down	SLC35F2	9	0.002137	334,890	0.208127
Down	MCUB	8	0.001343	376,614	0.228773
Down	EN1	7	0.001288	650,718	0.214239
Down	NXPH3	7	0.001042	394,474	0.225685
Down	PRDM6	6	8.27E-04	180,818	0.199541
Down	ACKR2	6	0.001234	909,848	0.168077
Down	SIX1	5	0.003887	1,852,370	0.236581
Down	CRLF1	5	4.88E-04	106,980	0.20325
Down	LZTS1	5	8.75E-04	197,898	0.219357
Down	CDKL2	5	0.001644	383,036	0.220071
Down	EPHA3	4	8.78E-05	38,542	0.232634
Down	SLC16A11	4	0.001233	310,374	0.20308
Down	MYOZ3	4	8.48E-04	232,490	0.20818
Down	DUSP2	3	8.09E-06	4042	0.210113
Down	VLDLR	3	1.30E-04	45,712	0.232179
Down	FPR1	3	1.55E-05	5552	0.223858
Down	PCDHA12	3	8.37E-04	124,810	0.228472
Down	SERPINE2	2	3.16E-04	17,692	0.221949
Down	NTRK2	2	2.65E-04	169,496	0.24658
Down	BGN	2	5.42E-05	11,718	0.190197
Down	IL20RA	2	3.31E-04	115,232	0.234925
Down	PPP1R12B	2	6.15E-05	31,120	0.224054
Down	KRT7	2	4.36E-05	26,098	0.228784
Down	NOX4	2	1.13E-04	68,694	0.246442
Down	ANKRD1	2	0	0	0.208582
Down	PPP1R14A	2	1.75E-04	131,520	0.245497
Down	NGF	1	0	0	0.214173
Down	FGF7	1	0	0	0.21491
Down	RASSF2	1	0	0	0.199819
Down	ADAM12	1	0	0	0.191183
Down	ZFHX4	1	0	0	0.210194
Down	EMB	1	0	0	0.168806

Table 4 (continued)

Regulation	Node	Degree	Betweenness	Stress	Closeness
Down	ST3GAL6	1	0	0	0.209407
Down	DSE	1	0	0	0.209389
Down	OR51E2	1	0	0	0.18939
Down	GPC6	1	0	0	0.195477
Down	NRP2	1	0	0	0.213477
Down	THBS2	1	0	0	0.207187
Down	CHL1	1	0	0	0.199051
Down	ULBP1	1	0	0	0.185683
Down	SLC22A23	1	0	0	0.174346
Down	RAPGEF5	1	0	0	0.199051
Down	IL33	1	0	0	0.198628
Down	ADAP1	1	0	0	0.230234
Down	MYOZ1	1	0	0	0.208002
Down	DYSF	1	0	0	0.208127
Down	DLX5	1	0	0	0.177326
Down	FLI1	1	0	0	0.220881
Down	COL8A2	1	0	0	0.196059
Down	TINAGL1	1	0	0	0.217271
Down	PMEL	1	0	0	0.208448
Down	APLN	1	0	0	0.217271
Down	MTSS1	1	0	0	0.208448
Down	DEPP1	1	0	0	0.220891
Down	IL6	1	0	0	0.205342
Down	BEND5	1	0	0	0.21344
Down	RASSF9	1	0	0	0.202682
Down	MYOM2	1	0	0	0.200173
Down	FOXC2	1	0	0	0.183859
Down	FANK1	1	0	0	0.254631
Down	ZNF185	1	0	0	0.238167
Down	ITGB1BP2	1	0	0	0.18209
Down	PDLIM3	1	0	0	0.172315
Down	SORCS2	1	0	0	0.179491
Down	PITPNM3	1	0	0	0.195886
Down	TPD52L1	1	0	0	0.228676
Down	PAPPA2	1	0	0	0.191183
Down	WNK4	1	0	0	0.212396
Down	STXBP6	1	0	0	0.221615

(ex; *hsa-mir-556-3p*) targeting regulation of *epb41l3*, 48 mirnas (ex; *hsa-mir-10b-5p*) targeting regulation of *dapk1* and 41 mirnas (ex; *hsa-mir-1229-5p*) targeting regulation of *mdfi*, and are listed in Table 5.

Construction of the TF-hub gene regulatory network

We searched for target-regulated hub gene TFs using NetworkAnalyst database and then used the results of this database. By constructing TF-hub gene regulatory network networks, we found 520 nodes (TF: 198; Hub

Gene: 322) and 8331 edges (Fig. 6). We identified 59 TFs (ex; *tcf3*) targeting regulation of *ptch1*, 42 tfs (ex; *phc1*) targeting regulation of *ccnd2*, 37 tfs (ex; *nr1i2*) targeting regulation of *prkcb*, 36 tfs (ex; *hoxc9*) targeting regulation of *st8sia4*, 34 tfs (ex; *rnf2*) targeting regulation of *foxq1*, 44 tfs (ex; *clock*) targeting regulation of *dapk1*, 38 tfs (ex; *prdm14*) targeting regulation of *igf2bp1*, 36 tfs (ex; *smarca4*) targeting regulation of *krt18*, 34 tfs (ex; *trim28*) targeting regulation of *stx11* and 31 tfs (ex; *htt*) targeting regulation of *epb41l3*, and are listed in Table 5.

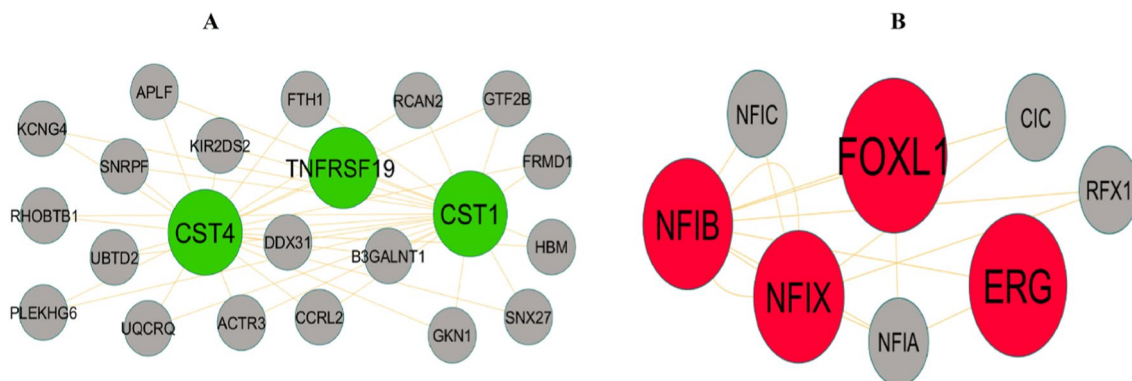


Fig. 4 Modules selected from the PPI network. **A** The most significant module was obtained from PPI network with 22 nodes and 41 edges for upregulated genes **B** The most significant module was obtained from PPI network with 8 nodes and 14 edges for downregulated genes. Upregulated genes are marked in parrot green; downregulated genes are marked in red



Fig. 5 Hub gene—miRNA regulatory network. The olive green color diamond nodes represent the key miRNAs; upregulated genes are marked in green; downregulated genes are marked in red

Receiver operating characteristic curve (ROC) analysis

The ROC curve was used to evaluate the diagnostic value of hub genes. As shown in Fig. 7, the AUC values of *vcam1*, *snc*, *prkcb*, *adrb2*, *foxq1*, *mdfi*, *actbl2*, *prkd1*, *dapk1* and *actc1* in endometriosis were 0.904, 0.907, 0.903, 0.926, 0.901, 0.910, 0.923, 0.892, 0.895 and 0.898, respectively. Thus, the hub genes have good diagnostic efficiency in endometriosis and normal control samples.

Discussion

Endometriosis is a key cause of serious reproductive disorder in the female population and leads to a public health burden. Lack of early screening and diagnosis of

endometriosis result in progressive worsening including dysmenorrhea, dyspareunia, chronic pelvic pain, irregular uterine bleeding and infertility. The advance stage of endometriosis seriously affects the recovery from female reproductive diseases. Therefore, it is necessary to identify potential novel biomarkers for early diagnosis and targeted therapy of endometriosis. With the advancement of bioinformatics methods and NGS technology, it has started to be widely applied to identify potential novel biomarkers. This investigation used NGS data to conduct bioinformatics analysis for identifying novel target genes and pathways involved in the occurrence and development of endometriosis. Bioinformatics and NGS

Table 5 MiRNA–hub gene and TF–hub gene topology table

Regulation	Hub Genes	Degree	MicroRNA	Regulation	Hub Genes	Degree	TF
Up	CCND2	365	hsa-mir-3143	Up	PTCH1	59	TCF3
Up	VCAM1	102	hsa-mir-6888-5p	Up	CCND2	42	PHC1
Up	PTPRD	89	hsa-mir-200a-3p	Up	PRKCB	37	NR1I2
Up	PDGFB	88	hsa-mir-3122	Up	ST8SIA4	36	HOXC9
Up	PRKCB	81	hsa-mir-17-5p	Up	FOXQ1	34	RNF2
Up	PIK3R3	78	hsa-mir-93-5p	Up	PDGFB	34	RELA
Up	FOXQ1	62	hsa-mir-5706	Up	NES	32	KLF4
Up	MME	54	hsa-mir-186-5p	Up	MME	31	BMI1
Up	PTCH1	52	hsa-mir-372-3p	Up	PIK3R3	30	RUNX1
Up	SNCA	37	hsa-mir-369-3p	Up	PTPRD	27	PAX3
Up	S100A4	36	hsa-mir-194-3p	Up	ADRB2	27	GATA2
Up	ST8SIA4	31	hsa-mir-181c-5p	Up	SNCA	25	SCLY
Up	ADRB2	22	hsa-mir-10b-5p	Up	S100A4	20	ARNT
Up	NES	21	hsa-mir-487b-5p	Up	VCAM1	16	SMAD3
Up	NTRK1	4	hsa-mir-147a	Up	NTRK1	13	JARID2
Down	IGF2BP1	241	hsa-mir-2110	Down	DAPK1	44	CLOCK
Down	ACTC1	77	hsa-mir-4432	Down	IGF2BP1	38	PRDM14
Down	EPB41L3	53	hsa-mir-556-3p	Down	KRT18	36	SMARCA4
Down	DAPK1	48	hsa-mir-10b-5p	Down	STX11	34	TRIM28
Down	MDFI	41	hsa-mir-1229-5p	Down	EPB41L3	31	HTT
Down	FOXL1	36	hsa-mir-7-5p	Down	FOXL1	29	STAT3
Down	SPINT2	29	hsa-mir-4677-3p	Down	ERG	28	TFAP2C
Down	KRT18	27	hsa-mir-126-3p	Down	SPINT2	26	NACC1
Down	EEF1A2	27	hsa-mir-16-5p	Down	EEF1A2	24	DNAJC2
Down	STX11	21	hsa-mir-373-3p	Down	CHRM4	22	SOX9
Down	PRKD1	19	hsa-mir-34a-5p	Down	PRKD1	21	MTF2
Down	ACTBL2	14	hsa-mir-599	Down	ACTBL2	18	YAP1
Down	ERG	9	hsa-mir-145-5p	Down	MDFI	17	TET1
Down	FOXS1	8	hsa-mir-101-3p	Down	ACTC1	15	TBX5
Down	CHRM4	4	hsa-mir-146a-5p	Down	FOXS1	11	EP300

data analysis might become to identify effective drugs for treating endometriosis in future.

In this investigation, we analyzed the endometriosis GSE243039 screened from the GEO database. It includes 20 normal control samples and 20 endometriosis samples. Compared to normal controls, we found 958 DEGs (including 479 upregulated genes and 479 downregulated genes). Research has shown that *pcsk9* [48], *cntn4* [49], *sema3a* [50], *sfrp4* [51], *mfap5* [52], *bmp6* [53], *cdh6* [54], *piezo2* [55] and *pkp2* [56] play an important role in the pathogenesis of inflammation. Some studies have shown that altered expression of genes includes *pcsk9* [57], *sema3a* [58] and *sfrp4* [59] promotes the pain. Studies have revealed that genes including *pcsk9* [60], *cntn4* [61], *sema3a* [62], *ptgis* [63], *sfrp4* [64], *mfap5* [65], *cdh6* [66], *gpc6* [67] and *pkp2* [68] play a key role in ovarian cancer. A study indicates that genes including *pcsk9* [69], *sfrp4* [70] and *bmp6* [71] have been identified in polycystic

ovarian syndrome. Genes including *pcsk9* [72], *cntn4* [49], *sema3a* [73], *sfrp4* [74], *mfap5* [75], *bmp6* [76], *pde1c* [77] and *pkp2* [78] are altered expressed in cardiovascular diseases. The genes including *pcsk9* [79], *apcdd1* [80], *sfrp4* [81], *mfap5* [82] and *pkp2* [83] have been identified to be involved in the development of obesity. Studies show that genes including *pcsk9* [84] and *sfrp4* [85] have been known to be involved in gestational diabetes mellitus. Recent reports have revealed that genes including *pcsk9* [86], *sema3a* [87], *sfrp4* [81] and *bmp6* [88] play an important role in the pathogenesis of diabetes mellitus. Recent reports have revealed that genes including *pcsk9* [89], *sema3a* [90], *ptgis* [91] and *piezo2* [92] have a significant prognostic potential in hypertension. Studies have found that genes including *adamts19* [93] and *bmp6* [94] play an indispensable role in infertility. Previous studies have reported that the genes including *sema3a* [95], *sfrp4* [96] and *bmp6* [94] are a key regulator of endometriosis.

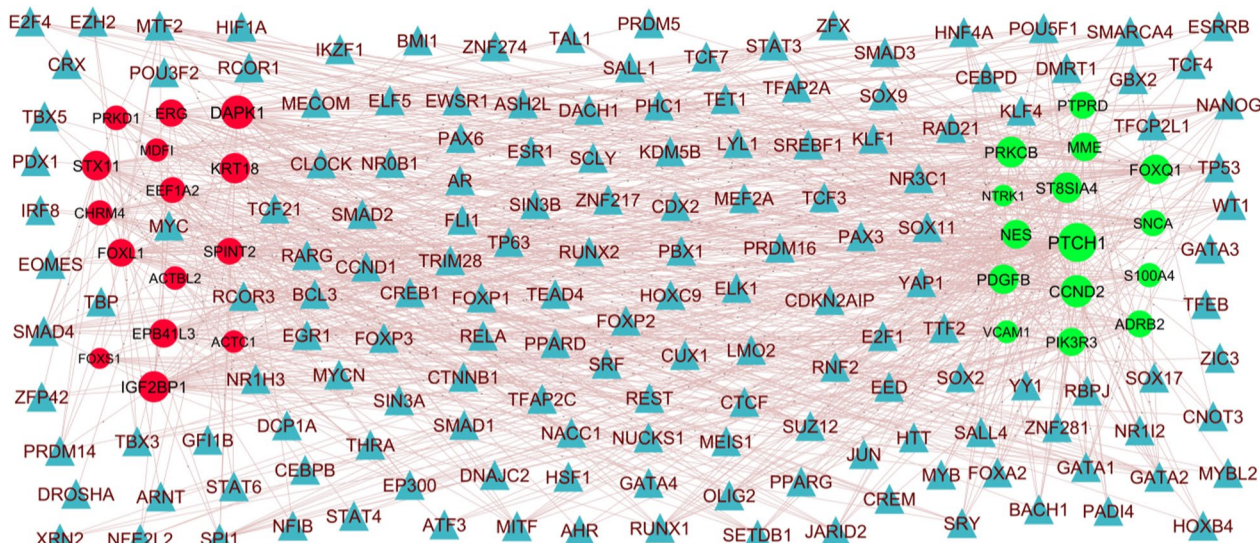


Fig. 6 Hub gene—TF regulatory network. The blue color triangle nodes represent the key TFs; upregulated genes are marked in dark green; downregulated genes are marked in dark red

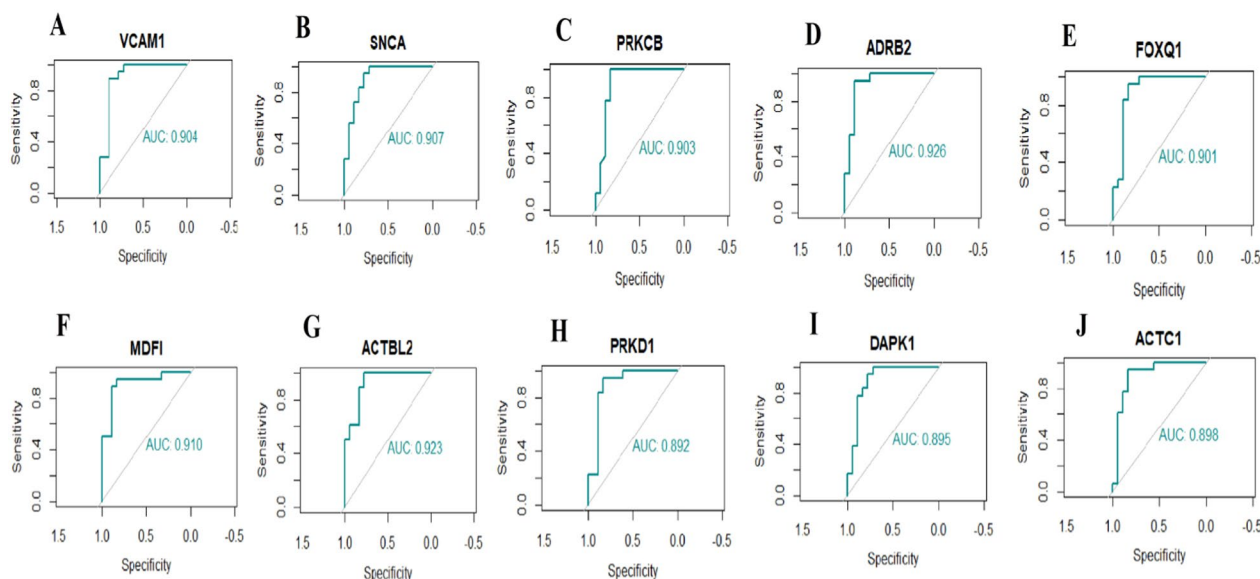


Fig. 7 ROC curve analyses of hub genes. **A** VCAM1 **B** SNCA **C** PRKCB **D** ADRB2 **E** FOXQ1 **F** MDFI **G** ACTBL2 **H** PRKD1 **I** DAPK1 **J** ACTC1

Recently, increasing evidence demonstrated that genes including *ptgis* [97] and *sfrp4* [98] might be potential therapeutic targets for endometrial cancer treatment. Studies have found that genes including *sfrp4* [99] and *mfap5* [100] are used as prognostic markers for cervical cancer. This result suggests that these genes might play a key role in the progression of endometriosis.

In this investigation, we identified enriched genes in GO terms and signaling pathways that might be utilized as diagnostic, prognostic and therapeutic targets in endometriosis. Signaling pathways including extracellular matrix organization [101], nervous system development [102], signal transduction [103], hemostasis [104], muscle contraction [105], signaling by retinoic acid [106] and diseases of glycosylation [107] were responsible for advancement of endometriosis. Recently, mounting

researches have revealed that genes including *l1cam* [108], *hsd17b2* [109], *vcam1* [110], *sox6* [111], *fgf10* [112], *mmp12* [113], *ccr1* [114], *prok1* [115], *prl* [116], *timp3* [117], *adamts9* [118], *ndnf* [119], *lhcr* [120], *pdgfb* [121], *ldlr* [122], *cd4* [123], *foxl2* [124], *trpa1* [125], *adrb2* [126], *plau* [127], *epcam* [128], *ucn2* [129], *cyp1a1* [130], *ntn1* [131], *il15* [132], *bmp2* [133], *apoe* [134], *casp1* [135], *abcg2* [136], *ace* [137], *pgr* [138], *alpp* [139], *lpar4* [140], *atrnl1* [141], *hla-c* [142], *mmp3* [143], *pdlim3* [144], *nfasc* [145], *il33* [146], *ngf* [147], *comp* [148], *fst* [149], *efemp1* [150], *gata6* [151], *tcf21* [152], *ptgs2* [153], *hoxc8* [154], *akr1c3* [155], *bdnf* [119], *epha3* [156], *inhba* [157], *rap1gap* [158], *tlr3* [159], *nox4* [160], *tgfb1* [161], *igf2bp1* [162], *dlx5* [163], *vdr* [164], *fzd7* [165], *id2* [166], *tlr2* [167], *il6* [168], *gas6* [169], *dusp2* [170], *fgf7* [171], *ccn2* [172], *igfbp3* [173], *chl1* [174], *bgn* [175], *ntrk2* [176], *slit2* [177], *notch2* [178], *lif* [179], *cd200* [180], *bst2* [181], *dysf* [182], *dapk1* [183], *kiss1* [184], *fpr1* [185] and *trh* [186] were vital for the onset and developmental process of endometriosis. A great number of studies have indicated that genes including *l1cam* [187], *ajap1* [188], *hsd17b2* [189], *vcam1* [190], *grp* [191], *aqp8* [192], *wnt6* [193], *fabp4* [194], *sox6* [195], *ntrk1* [196], *cntn1* [197], *mmp12* [198], *lag3* [199], *sox18* [200], *ccr1* [201], *flt1* [202], *prdm1* [203], *trpc3* [204], *dkk2* [205], *rnf157* [206], *dhcr24* [207], *bmp4* [208], *prl* [209], *foxq1* [210], *wnt5a* [211], *meox1* [212], *dock4* [213], *timp3* [214], *adamts9* [215], *ndnf* [216], *neto1* [217], *cd24* [218], *lhcr* [219], *scd* [220], *pdgfb* [221], *mmrn1* [222], *ldlr* [223], *cd4* [224], *foxl2* [225], *trpa1* [226], *epha5* [227], *tox* [228], *cst4* [229], *rspo3* [230], *map2k6* [231], *nes* [232], *tmem119* [233], *padi2* [234], *mmp8* [235], *kdr* [236], *adrb2* [237], *mgat3* [238], *ptprc* [239], *pitx1* [240], *kl* [241], *plau* [242], *znf365* [243], *pik3r3* [244], *sox8* [245], *ccnd2* [246], *crabp2* [247], *pcdh9* [248], *epcam* [249], *clec14a* [250], *cyp1a1* [251], *ntn1* [252], *pdgfd* [253], *cldn3* [254], *lepr* [255], *il15* [256], *bmp2* [257], *lama5* [258], *ntng1* [259], *krt19* [260], *ros1* [261], *apoe* [262], *ptch1* [263], *itpka* [264], *casp1* [265], *nid1* [266], *abcg2* [267], *ace* [268], *pgr* [269], *wls* [270], *klk3* [271], *lrp1b* [272], *ly6k* [273], *alpp* [274], *prame* [275], *slco4a1* [276], *egfl6* [277], *gpbar1* [278], *elmo1* [279], *wnk2* [280], *il2rb* [281], *diras2* [282], *galnt14* [283], *rtkn2* [284], *atrnl1* [285], *s100a4* [286], *macc1* [287], *mmp3* [288], *col11a1* [289], *cables1* [290], *fgfr2* [291], *il33* [292], *ngf* [293], *foxc2* [294], *comp* [295], *fst* [296], *sorbs2* [297], *efemp1* [298], *gata6* [299], *tcf21* [300], *ptgs2* [301], *mtss1* [302], *dact1* [303], *hoxc8* [304], *pitx2* [305], *tnfsf10* [306], *bdnf* [307], *krt7* [308], *ndrg2* [309], *eya2* [310], *inhba* [311], *sgk1* [312], *slc2a12* [313], *dio3* [314], *epb41l3* [315], *tlr3* [316], *angptl4* [317], *ephb2* [318], *fli1* [319], *thbs1* [320], *id3* [321], *nox4* [322], *tgfb1* [323], *igf2bp1* [324], *sall4* [325], *dlx5* [326], *vdr* [327], *lzts1* [328], *fzd7* [329], *en2* [330], *enc1* [331], *ifne* [332], *tnnt1* [333],

ankrd1 [334], *sox9* [335], *mgp* [336], *sulf1* [337], *cyp24a1* [338], *dnah11* [339], *tlr2* [340], *il6* [341], *nppb* [342], *spink1* [343], *gpc3* [344], *ntrk3* [345], *amigo2* [346], *foxd1* [347], *adam12* [348], *dusp2* [349], *usp2* [350], *klf2* [351], *sik1* [352], *six1* [310], *fgf7* [353], *myh10* [354], *igfbp3* [355], *lyve1* [356], *actbl2* [357], *slit2* [358], *actc1* [359], *nnmt* [360], *chi3l1* [361], *runx1* [362], *nfib* [363], *notch2* [364], *pgf* [365], *thbs2* [366], *nav1* [367], *nrg1* [368], *plk2* [369], *itgbl1* [370], *cd200* [371], *bst2* [372], *kcnn3* [373], *hmcn1* [374], *veph1* [375], *tfpi2* [376], *sylt2* [377], *ccdc80* [378], *dapk1* [379], *kiss1* [380], *il20ra* [381], *has3* [382], *has1* [382], *mgst1* [383], *fpr1* [384] and *sh3rf2* [385] are closely associated with the onset and progression of ovarian cancer. A previous study reported that the genes including *l1cam* [386], *hsd17b2* [387], *grp* [388], *fabp4* [389], *sox6* [390], *mmp12* [391], *apod* [392], *lag3* [393], *cst1* [394], *flt1* [395], *dhcr24* [396], *prl* [397], *wnt5a* [398], *timp3* [399], *cd24* [400], *lhcr* [401], *mmrn1* [402], *cd4* [403], *adamts5* [404], *adamts1* [405], *padi2* [406], *mark1* [407], *kl* [408], *plau* [409], *sox8* [410], *crabp2* [411], *ptprd* [412], *epcam* [413], *irx2* [414], *sema3b* [415], *cyp1a1* [416], *pdgfd* [417], *lepr* [418], *apoe* [419], *casp1* [420], *mgll* [421], *nid1* [422], *abcg2* [423], *ace* [424], *pgr* [425], *hpse2* [426], *lmtk3* [427], *alpp* [428], *egfl6* [429], *cacna2d3* [430], *mctp1* [431], *hkdc1* [432], *s100a4* [433], *macc1* [434], *mmp3* [435], *fgfr2* [436], *il33* [437], *foxc2* [438], *itga7* [439], *efemp1* [440], *gata6* [441], *bhlhe41* [442], *tcf21* [443], *gdf10* [444], *nkx3-1* [445], *akr1c3* [446], *sgk1* [447], *rap1gap* [448], *fli1* [449], *nox4* [450], *serpine2* [451], *igsf9* [452], *igf2bp1* [453], *sall4* [454], *vdr* [455], *celsr2* [456], *enc1* [457], *sox9* [458], *cyp24a1* [338], *il6* [459], *gas6* [460], *klf2* [461], *six1* [462], *igfbp3* [463], *lyve1* [464], *chl1* [465], *bgn* [466], *slit2* [467], *nrp2* [468], *nnmt* [469], *runx1* [470], *thbs2* [471], *hspb7* [472], *nrg1* [473], *tfpi2* [474], *dapk1* [183], *has3* [475], *has1* [475], *steap1* [476] and *mgst1* [477] play an important role in the pathophysiology of endometrial cancer. Study demonstrated that genes including *vcam1* [478], *aqp8* [479], *l1cam* [480], *fabp4* [481], *psg1* [482], *sox6* [483], *mmp12* [484], *apod* [485], *lag3* [486], *sox18* [487], *flt1* [488], *fabp5* [489], *bmp4* [490], *prl* [491], *foxq1* [492], *wnt5a* [493], *frzb* [494], *cpe* [495], *ereg* [496], *ndnf* [497], *cd24* [498], *scd* [499], *ldlr* [500], *cd4* [501], *foxl2* [502], *krt17* [503], *nes* [504], *mgat3* [505], *mark1* [506], *kl* [507], *plau* [508], *epha7* [509], *pik3r3* [510], *ccnd2* [511], *hecw1* [512], *epcam* [513], *batf2* [514], *cyp1a1* [515], *mstn* [516], *il15* [517], *syt7* [518], *pak3* [519], *krt19* [520], *ros1* [521], *cubn* [522], *ptch1* [523], *casp1* [524], *abcg2* [525], *pgr* [526], *hpse2* [527], *lrp1b* [528], *alpp* [529], *cyp2s1* [530], *doc2b* [531], *msmo1* [532], *sorcs1* [533], *hla-c* [534], *s100a4* [535], *macc1* [536], *mmp3* [537], *fgfr2* [538], *il33* [539], *ngf* [540], *foxc2* [541], *sorbs2* [542], *itga7* [543], *efemp1* [544], *gata6* [545], *tcf21* [546], *ptgs2* [547], *mtss1* [548],

dact1 [549], *spint2* [550], *nkx3-1* [551], *hoxc8* [552], *bdnf* [497], *ndrg2* [553], *epha3* [554], *eya2* [555], *inhba* [556], *alpl* [557], *sgk1* [558], *rap1gap* [559], *epb41l3* [560], *tlr3* [561], *angptl4* [562], *ephb2* [563], *fli1* [564], *thbs1* [565], *nox4* [566], *tgfb1* [567], *igf2bp1* [568], *sall4* [569], *vdr* [570], *rarb* [571], *epha4* [572], *enc1* [573], *sox9* [574], *sulf1* [575], *tlr2* [576], *il6* [577], *gpc3* [578], *ntrk3* [579], *ccna1* [580], *amigo2* [581], *foxd1* [582], *ccno* [583], *adam12* [584], *rassf2* [585], *hoxb7* [586], *klf2* [587], *sik1* [588], *six1* [589], *fgf7* [590], *igfbp3* [591], *chl1* [592], *eppk1* [593], *slit2* [594], *flg* [595], *nrp2* [596], *nnmt* [597], *chi3l1* [598], *runx1* [599], *apln* [600], *sema3c* [601], *notch2* [602], *thbs2* [603], *pnpla1* [604], *bst2* [605], *hmcn1* [606], *ulbp1* [607], *tfpi2* [608], *dapk1* [609], *kiss1* [610], *fpr1* [611] and *pik3ap1* [612] can participate in the occurrence and development of cervical cancer. The abnormal expression of genes including *cbln2* [613], *sdk1* [614], *vcam1* [615], *six2* [616], *avpr1a* [617], *epha6* [618], *fabp4* [619], *psg1* [620], *ano1* [621], *sox6* [622], *fgf10* [623], *pla2g7* [624], *mmp12* [625], *adra1d* [626], *lag3* [627], *flt1* [628], *fabp5* [629], *prdm1* [630], *trpc3* [631], *igsf3* [632], *bmp4* [633], *il1rl1* [634], *prl* [635], *nefl* [636], *wnt5a* [637], *timp3* [638], *ndnf* [639], *snap25* [640], *cd24* [641], *pdgfb* [642], *ldlr* [643], *cd4* [644], *trpa1* [645], *adamts1* [646], *pde4b* [647], *nes* [648], *th* [649], *psg9* [650], *cacna1d* [651], *mmp8* [652], *adrb2* [653], *kl* [654], *plau* [655], *ptprd* [656], *sema3b* [657], *ucn2* [658], *cyp2j2* [659], *cyp1a1* [660], *atp1a2* [661], *cldn3* [662], *mstn* [663], *lepr* [664], *il15* [665], *cacna1h* [666], *bmp2* [667], *lama5* [668], *ros1* [669], *apoe* [670], *casp1* [671], *pde9a* [672], *efnb2* [673], *abcg2* [674], *ace* [675], *pgr* [676], *slc35f3* [677], *ica1* [678], *alpp* [679], *trpc6* [680], *gpbar1* [681], *pnpla3* [682], *hla-c* [683], *s100a4* [684], *macc1* [685], *mmp3* [686], *gdnf* [687], *fgfr2* [688], *il33* [689], *ngf* [690], *pappa2* [691], *comp* [692], *gata6* [693], *acana* [694], *tcf21* [695], *ptgs2* [696], *pitx2* [697], *akr1c3* [698], *bdnf* [699], *sgk1* [700], *tlr3* [701], *angptl4* [702], *fli1* [703], *thbs1* [704], *id3* [705], *nox4* [706], *pcsk1* [707], *itgb1bp2* [708], *wnk4* [709], *dlx5* [710], *vdr* [711], *epha4* [712], *mgp* [713], *cyp24a1* [714], *id2* [715], *tlr2* [716], *il6* [717], *nppb* [718], *gas6* [719], *f11r* [720], *foxd1* [721], *adam12* [722], *ncam1* [723], *usp2* [724], *klf2* [725], *sik1* [726], *fgf7* [727], *igfbp3* [728], *bgn* [729], *ntrk2* [730], *nnmt* [731], *chi3l1* [732], *runx1* [733], *apln* [734], *stox2* [735], *kcnq4* [736], *notch2* [737], *pgf* [738], *thbs2* [739], *pdlim5* [740], *prdm6* [741], *htr6* [742], *nrg1* [743], *cd200* [744], *bst2* [745], *kcnn3* [746], *slc2a5* [747], *tfpi2* [748], *dysf* [749], *ccdc80* [750], *dapk1* [751], *kiss1* [752], *slc4a4* [753], *steap2* [754], *sorbs1* [755], *ackr2* [756], *fpr1* [757], *gpr143* [758] and *trh* [759] contributes to the progression of hypertension. Study showed that the genes including *robo2* [760], *vcam1* [761], *grp* [762], *fabp4* [763], *ano1* [764], *sox6* [765], *tfap2c* [766], *ramp3* [767], *pla2g7* [768], *mmp12* [769], *faim2* [770], *apod* [771], *lag3* [772], *sox18* [773], *f2rl2* [774], *ccr1* [775], *flt1* [776], *fabp5* [629], *trpc3* [777], *thsd7a* [778], *dkk2* [779], *prkcb* [780], *dhcr24* [781], *pde3b* [782], *bmp4* [783], *il1rl1* [784], *mypn* [785], *plcg2* [786], *prl* [787], *wnt5a* [788], *meox1* [789], *timp3* [790], *frzb* [791], *cpe* [792], *adamts9* [793], *ndnf* [794], *pdgfb* [795], *pik3cg* [796], *ldlr* [797], *cd4* [798], *trpa1* [799], *f2rl3* [800], *c1ql1* [801], *adamts5* [802], *pde4b* [803], *nes* [804], *th* [805], *mmp8* [806], *kdr* [807], *adrb2* [808], *ackr3* [809], *ptprc* [810], *kl* [811, 812], *plau* [813], *ccnd2* [814], *ptgs1* [815], *insig1* [816], *irx2* [817], *siglec1* [818], *ucn2* [819], *cyp2j2* [820], *cyp1a1* [821], *astn2* [822], *ntn1* [823], *pdgfd* [824], *mstn* [663], *lepr* [664], *il15* [825], *cacna1h* [826], *bmp2* [827], *syt7* [828], *zbtb46* [829], *ros1* [830], *apoe* [831], *cubn* [832], *rbm20* [833], *casp1* [834], *pde9a* [835], *abcg2* [836], *hmgcr* [837], *ace* [838], *grem2* [839], *palmd* [840], *lrp1b* [841], *alpp* [842], *trpc6* [843], *gpbar1* [844], *myzap* [845], *prodh* [846], *il2rb* [847], *cdhr3* [848], *pnpla3* [849], *fads1* [850], *hla-c* [851], *s100a4* [852], *mmp3* [853], *pdlim3* [854], *gdnf* [855], *fgfr2* [856], *il33* [857], *ngf* [858], *hapln1* [859], *foxc2* [860], *comp* [861], *fst* [862], *sorbs2* [863], *itga7* [864], *pln* [865], *gata6* [866], *bhlhe41* [867], *acana* [868], *tcf21* [869], *ptgs2* [870], *dact1* [871], *pitx2* [872], *akr1c3* [873], *bdnf* [874], *ndrg2* [875], *eya2* [876], *sgk1* [877], *rap1gap* [878], *dio3* [879], *tlr3* [880], *angptl4* [881], *ephb2* [882], *thbs1* [883], *tnnt2* [884], *nox4* [885], *s1pr5* [886], *serpine2* [887], *pcsk1* [888], *tgfb1* [889], *sall4* [890], *eya4* [891], *itgb1bp2* [708], *vdr* [892], *gpc4* [893], *celsr2* [894], *epha4* [895], *tnnt1* [896], *ankrd1* [897], *zfp2m2* [898], *sox9* [899], *mgp* [900], *cyp24a1* [901], *dnah11* [902], *tlr2* [903], *il6* [904], *gas6* [905], *gpc3* [906], *ntrk3* [907], *amigo2* [908], *f11r* [909], *adam12* [722], *ncam1* [910], *usp2* [911], *klf2* [912], *sik1* [913], *six1* [914], *fgf7* [915], *ccn2* [916], *icad* [917], *igfbp3* [918], *lyve1* [919], *prkd1* [920], *bgn* [921], *eda* [922], *slit2* [923], *actc1* [924], *nrp2* [925], *chi3l1* [926], *runx1* [927], *apln* [928], *myom2* [929], *myoz1* [930], *ppp1r13l* [931], *thbs2* [932], *des* [933], *pdlim5* [934], *hspb7* [935], *nrg1* [936], *plk2* [937], *itgbl1* [938], *cd200* [939], *kcnn3* [940], *kcnj2* [941], *eva1a* [942], *tfpi2* [943], *dysf* [944], *adap1* [945], *ccdc80* [946], *dapk1* [947], *scn4b* [948], *esyt3* [949], *abca8* [950], *heg1* [951], *fpr1* [952], *sspn* [953], *adh1c* [954], *sirpa* [955] and *trh* [956] might be related to the pathophysiology of cardiovascular diseases. A study showed genes including *hsd17b2* [109], *efna5* [957], *mmp12* [958], *prok1* [959], *prl* [960], *nlrp2* [961], *ndnf* [962], *mei4* [963], *cd24* [964], *lhcr* [965], *cd4* [966], *foxl2* [967], *kdr* [968], *adrb2* [969], *cyp1a1* [970], *ntn1* [971], *mstn* [972], *bmp2* [973], *apoe* [974], *ace* [975], *pgr* [976], *grem2* [977], *alpp* [978], *mmp3* [979], *gdnf* [980], *fgfr2* [981], *il33* [982], *ngf* [983], *comp* [984], *cccr2* [985], *fst* [986], *gata6* [987], *ptgs2* [153], *bdnf* [988], *sgk1* [989], *angptl4* [990], *thbs1* [991], *id3* [992], *nox4* [993], *igf2bp1* [994], *sall4* [995], *vdr* [996], *sulf1* [997], *tlr2* [998], *il6*

[999], *gpc3* [1000], *ccno* [1001], *igfbp3* [1002], *chl1* [1003], *ntrk2* [1004], *slit2* [1005], *apln* [1006], *notch2* [1007], *pgf* [1008], *lif* [1009], *cd200* [1010], *tfpi2* [1011], *kiss1* [752] and *trh* [1012] are highly prone to infertility. A study indicated that activation of genes including *vcam1* [761], *stra6* [1013], *coch* [1014], *grp* [1015], *aqp8* [1016], *fabp4* [1017], *ano1* [1018], *sox6* [1019], *tfap2c* [1020], *ntrk1* [1021], *cntn1* [1022], *fgf10* [1023], *pla2g7* [768], *mmp12* [1024], *lcp1* [1025], *snca* [1026], *apod* [1027], *lag3* [1028], *ccr1* [1029], *cst1* [1030], *retreg1* [1031], *flt1* [1032], *fabp5* [1033], *prdm1* [1034], *trpc3* [1035], *prok1* [1036], *wnt16* [1037], *f13a1* [1038], *dhcr24* [1039], *pde3b* [1040], *bmp4* [783], *il1rl1* [1041], *plcg2* [1042], *prl* [1043], *foxq1* [1044], *nefl* [1045], *wnt5a* [1046], *timp3* [1047], *serpinb2* [1048], *frzb* [1049], *nlrp2* [1050], *cpe* [1051], *adamts9* [1052], *npw* [1053], *ereg* [1054], *ndnf* [1055], *snap25* [1056], *syt1* [1057], *scd* [1058], *pdgfb* [1059], *ldlr* [1060], *cd4* [1061], *gpr183* [1062], *trpa1* [125], *ptger4* [1063], *adamts5* [1064], *rspo3* [1065], *krt17* [1066], *adamts1* [1067], *pde4b* [1068], *nes* [804], *sh2d2a* [1069], *th* [1070], *tmem119* [1071], *mmp8* [1072], *adrb2* [1073], *ackr3* [1074], *mgat3* [1075], *tnfrsf9* [1076], *txk* [1077], *kl* [811], *plau* [1078], *epha7* [1079], *znf365* [1080], *pik3r3* [1081], *sox8* [1082], *ccnd2* [1083], *ptgs1* [1084], *batf2* [1085], *ucn2* [1086], *cyp2j2* [1087], *clec14a* [1088], *cyp1a1* [1089], *ntn1* [1090], *mstn* [1091], *lepr* [1092], *cd248* [1093], *il15* [825], *bmp2* [1094], *ros1* [1095], *mme* [1096], *apoe* [1097], *ptch1* [1098], *casp1* [1099], *mgll* [1100], *efnb2* [1101], *abcg2* [1102], *hmgcr* [1103], *ace* [1104], *pgr* [1105], *grem2* [839], *rgs7* [1106], *chst1* [1107], *alpp* [1108], *trpc6* [1109], *slco4a1* [276], *cyp4b1* [1110], *gpbar1* [844], *elmo1* [1111], *doc2b* [1112], *cd163l1* [1113], *slco2a1* [1114], *il2rb* [1115], *b4galnt2* [1116], *slc37a2* [1117], *pnpla3* [1118], *fads1* [1119], *hla-c* [1120], *st3gal5* [1121], *s100a4* [1122], *macc1* [1123], *cort* [1124], *mmp3* [1125], *gdnf* [1126], *lmo3* [1127], *nfasc* [1128], *fgfr2* [1129], *il33* [1130], *ngf* [1131], *hapln1* [1132], *gdf6* [1133], *foxc2* [1134], *comp* [1135], *fst* [1136], *itga7* [1137], *gata6* [1138], *acan* [1139], *pcf21* [1140], *ptgs2* [1141], *mtss1* [1142], *dhhrs3* [1143], *nkx3-1* [1144], *tnfsf10* [1145], *bdnf* [1146], *ndrg2* [1147], *epha3* [1148], *pla2g5* [1149], *mecom* [1150], *sgk1* [1151], *tlr3* [1152], *angptl4* [1153], *ephb2* [1154], *fli1* [1155], *thbs1* [1156], *mbp* [1157], *id3* [1158], *nox4* [1159], *s1pr5* [1160], *pi16* [1161], *igf2bp1* [1162], *sall4* [1163], *vldlr* [1164], *vdr* [1165], *fam20a* [1166], *epha4* [1167], *ankrd1* [1168], *sgca* [1169], *sox9* [1170], *mgp* [1171], *cyp24a1* [1172], *tlr2* [1173], *il6* [1174], *gas6* [1175], *ntrk3* [1176], *adam12* [1177], *ncam1* [1178], *myoc* [1179], *usp2* [1180], *klf2* [1181], *sik1* [1182], *six1* [1183], *fgf7* [1184], *ccn2* [1185], *igfbp3* [1186], *lyve1* [1187], *prkd1* [1188], *bgn* [1189], *slit2* [1190], *irx3* [1191], *actc1* [1192], *flg* [1193], *nrp2* [1194], *nnmt* [1195], *chi3l1* [1196], *runx1* [1197], *nfib* [1198], *apln* [1199], *plp1* [1200], *nav2* [1201], *notch2* [1202], *pgf* [1203], *thbs2* [1204], *nrg1* [1205], *lif* [1206], *plk2* [1207], *nalc1* [1208], *cd200* [1209], *kcnn3* [1210], *eva1a* [1211], *tfpi2* [1212], *dysf* [1213], *syt2* [1214], *tlr1* [1215], *ccdc80* [946], *dapk1* [947], *kiss1* [1216], *gem* [1217], *il20ra* [1218], *has3* [1219], *has1* [1220], *slc4a4* [1221], *sirpb1* [1222], *steap1* [1223], *ackr2* [1224], *fpr1* [1225], *gng7* [1226], *igfbpl1* [1227], *pik3ap1* [1228], *adh1c* [1229], *lxn* [1230] and *trh* [1231] has been observed in inflammation. Studies have suggested that genes including *vcam1* [1232], *aqp8* [1233], *fabp4* [1234], *fabp5* [1235], *bmp4* [1236], *prl* [960], *wnt5a* [1237], *adamts9* [1238], *ndnf* [1239], *lhcr* [1240], *ldlr* [1241], *cd4* [1242], *adamts5* [1243], *map2k6* [1244], *adamts1* [1245], *pde4b* [1246], *th* [1247], *mmp8* [1248], *adrb2* [969], *kl* [1249], *epha7* [1250], *ucn2* [1251], *cyp1a1* [970], *lepr* [1252], *il15* [1253], *bmp2* [1254], *apoe* [1255], *casp1* [1256], *ace* [1257], *pgr* [1258], *grem2* [1259], *sorcs1* [1260], *hkdc1* [1261], *fads1* [1262], *s100a4* [1263], *il33* [1243], *ngf* [1264], *comp* [1265], *fst* [1266], *gata6* [1267], *acan* [1268], *akr1c3* [1269], *bdnf* [1270], *angptl4* [1271], *nox4* [1272], *vdr* [1273], *gpc4* [1274], *tlr2* [1275], *il6* [1276], *igfbp3* [1277], *tnik* [1278], *apln* [1279], *pgf* [1280], *nrg1* [1281], *lif* [1282], *angptl1* [1283], *kiss1* [1284], *sorbs1* [1285] and *trh* [1286] can be used as important therapeutic targets for polycystic ovarian syndrome. A recent study found that genes including *vcam1* [1287], *stra6* [1288], *aqp8* [1289], *fabp4* [1290], *sox6* [1291], *ramp3* [1292], *mmp12* [1293], *faim2* [770], *ccr1* [1294], *ism1* [1295], *flt1* [1296], *fabp5* [1297], *thsd7a* [1298], *sctr* [1299], *wnt16* [1300], *prkcb* [1301], *pde3b* [1302], *il1rl1* [1041], *prl* [1303], *wnt5a* [1304], *htr1b* [1305], *timp3* [1306], *cpe* [1051], *ereg* [1307], *ndnf* [1308], *snap25* [1309], *cd24* [1310], *scd* [1058], *pdgfb* [1311], *ldlr* [1312], *cd4* [1313], *trpa1* [1314], *map2k6* [1315], *pde4b* [1316], *th* [1317], *mmp8* [1318], *adrb2* [1319], *kl* [1320], *plau* [1321], *ptgs1* [1084], *insig1* [1322], *bmp8a* [1323], *ucn2* [1324], *ntn1* [1090], *pdgfd* [1325], *mstn* [1326], *lepr* [1092], *il15* [1327], *bmp2* [1328], *apoe* [1329], *casp1* [1330], *mgll* [1331], *nid1* [1332], *abcg2* [1333], *ace* [1334], *pgr* [1335], *grem2* [1336], *lrp1b* [1337], *alpp* [1338], *trpc6* [1339], *egfl6* [1340], *gpbar1* [1341], *aif1l* [1342], *gpat3* [1343], *sorcs1* [1260], *slc37a2* [1344], *fads1* [1345], *acsl5* [1346], *ptprn2* [1347], *s100a4* [1348], *macc1* [1349], *cort* [1350], *mmp3* [1351], *gdnf* [1352], *lmo3* [1353], *cables1* [1354], *il33* [1355], *ngf* [1356], *foxc2* [1357], *fst* [1136], *pln* [1358], *acan* [1359], *ptgs2* [1360], *gdf10* [1361], *cpne5* [1362], *dgat2* [1363], *bdnf* [1364], *rgs4* [1365], *epha3* [1366], *pla2g5* [1367], *sgk1* [700], *tlr3* [1368], *angptl4* [1369], *ephb2* [1370], *thbs1* [1371], *id3* [1372], *nox4* [1373], *pcsk1* [1374], *wnk4* [1375], *vldlr* [1376], *vdr* [1377], *gpc4* [1378], *ifne* [1379], *zfp2* [1380], *tlr2* [1381], *il6* [1382], *spink1* [1383], *gas6* [1384], *f11r* [1385], *siglec15* [1386], *adam12* [1387], *myoc* [1388], *usp2* [1389], *sik1* [1390], *ccn2* [1391], *igfbp3* [1392], *lyve1*

[1393], *bgn* [1394], *eda* [1395], *ntrk2* [1396], *slit2* [1397], *irx3* [1191], *nnmt* [1398], *chi3l1* [1399], *runx1* [1400], *apln* [1401], *pgf* [1402], *htr6* [742], *nrg1* [1403], *npy4r* [1404], *ccdc80* [1405], *kiss1* [1406], *slc6a15* [1407], *esyt3* [1408], *sorbs1* [1409], *slc38a3* [1410], *lxn* [1411] and *trh* [1412] are potential targets for obesity. Studies have shown that genes including *vcam1* [1413], *stra6* [1414], *aqp8* [1415], *fabp4* [1416], *flt1* [1417], *bmp4* [633], *prl* [1418], *adamts9* [1419], *ndnf* [1420], *dtx1* [1421], *cd4* [1422], *adamts5* [1423], *mmp8* [1424], *adrb2* [1425], *ptprd* [1426], *insig1* [1427], *lepr* [1428], *il15* [1429], *apoe* [1430], *ace* [1431], *lrp1b* [1432], *alpp* [1433], *hkdc1* [1434], *pnpla3* [1435], *fads1* [1436], *mmp3* [1437], *fgfr2* [1438], *il33* [1439], *foxc2* [1440], *fst* [1441], *hoxc8* [1442], *bdnf* [1443], *ndrg2* [1444], *angptl4* [1445], *tgfb1* [1446], *vdr* [1447], *gpc4* [1448], *cyp24a1* [1449], *tlr2* [1450], *il6* [1451], *klf2* [1452], *igfbp3* [1453], *slit2* [1454], *apln* [1455], *notch2* [1456], *pgf* [1457], *nrg1* [1458], *tlr1* [1459], *ccdc80* [1460] and *kiss1* [1461] are the contributing factors to gestational diabetes mellitus pathogenesis. A previous study identified genes including *vcam1* [1462], *stra6* [1013], *wnt6* [1463], *fabp4* [1464], *sox6* [1465], *pla2g7* [1466], *mmp12* [1467], *faim2* [770], *snca* [1468], *apod* [1469], *lag3* [1470], *prex1* [1471], *flt1* [1472], *fabp5* [1473], *trpc3* [1474], *thsd7a* [1475], *prkcb* [1476], *pde3b* [1477], *bmp4* [1478], *prl* [1479], *nefl* [1480], *wnt5a* [1481], *timp3* [1306], *cpe* [1482], *adamts9* [1483], *ndnf* [1484], *snap25* [1485], *scd* [1058], *ldlr* [1486], *cd4* [1487], *trpa1* [1488], *rspo3* [1489], *pde4b* [1490], *th* [1491], *cacna1d* [1492], *mmp8* [1318], *kdr* [1493], *adrb2* [653], *kl* [1494], *plau* [1321], *ccnd2* [1495], *ptprd* [1496], *siglec1* [1497], *ucn2* [1324], *cyp2j2* [1498], *cyp1a1* [1499], *ntn1* [1090], *mstn* [1326], *lepr* [1500], *il15* [1501], *bmp2* [1502], *apoe* [1503], *cubn* [1504], *casp1* [1505], *mgll* [1506], *efnb2* [1507], *nid1* [1508], *abcg2* [1102], *ace* [1509], *stmn2* [1510], *ica1* [1511], *trpc6* [1512], *gpbar1* [1513], *elmo1* [1514], *doc2b* [1515], *ank1* [1516], *sorcs1* [1517], *hkdc1* [1518], *pnpla3* [1519], *fads1* [1520], *acs15* [1521], *hla-c* [1522], *s100a4* [1523], *cort* [1524], *mmp3* [1525], *gdnf* [1526], *cables1* [1354], *il33* [1527], *ngf* [1528], *foxc2* [1529], *comp* [1530], *fst* [1531], *sorbs2* [1532], *gata6* [1533], *ptgs2* [1141], *dact1* [1534], *dgat2* [1535], *bdnf* [1484], *ndrg2* [1536], *sgk1* [1537], *tlr3* [1152], *angptl4* [1538], *ephb2* [1539], *thbs1* [1540], *mbp* [1541], *nox4* [1542], *pi16* [1543], *pcsk1* [888], *tgfb1* [1544], *igf2bp1* [1545], *wnk4* [1546], *vldlr* [1547], *vdr* [1548], *gpc4* [1274], *ptprn* [1549], *epha4* [712], *sox9* [1550], *mgp* [1551], *cyp24a1* [1552], *tlr2* [1553], *il6* [1554], *nppb* [1555], *spink1* [1556], *gas6* [1557], *f11r* [1558], *foxd1* [1559], *adam12* [1560], *klf2* [1561], *sik1* [1562], *fgf7* [1563], *igfbp3* [918], *lyve1* [1393], *eda* [1395], *slit2* [1564], *irx3* [1565], *nnmt* [1398], *chi3l1* [1566], *runx1* [1567], *apln* [1568], *col4a3* [1569], *notch2* [1570], *pdlim5* [740], *nrg1*

[1571], *dmrt2* [1572], *npy4r* [1573], *cd200* [1574], *bst2* [1575], *tspi2* [1576], *kiss1* [1406], *mpp7* [1577], *sorbs1* [1409], *slc38a3* [1578], *chn2* [1579] and *trh* [1580] have been implicated in diabetes mellitus pathology. A previous bioinformatics study suggested that genes including *grp* [1581], *avpr1a* [1582], *ano1* [1583], *ntrk1* [1584], *fgf10* [1585], *mmp12* [1586], *snca* [1587], *ccr1* [1588], *flt1* [1589], *fabp5* [1590], *trpc3* [1591], *bmp4* [1592], *prl* [1593], *wnt5a* [1594], *timp3* [1595], *serpinb2* [1596], *nlrp2* [1050], *npw* [1053], *ereg* [1597], *ndnf* [1598], *snap25* [1056], *syt1* [1599], *cd4* [1600], *gpr183* [1601], *trpa1* [1602], *pde4b* [1603], *th* [1604], *mmp8* [1605], *adrb2* [1606], *mgat3* [1607], *plau* [1608], *astrn2* [1609], *il15* [1610], *bmp2* [1611], *apoe* [1612], *pde9a* [1613], *mgll* [1100], *efnb2* [1101], *hmgcr* [1614], *ace* [1615], *syt9* [1616], *trpc6* [1617], *xcr1* [1618], *s100a4* [1619], *mmp3* [1620], *gdnf* [1621], *il33* [1622], *ngf* [1623], *gdf6* [1133], *comp* [1624], *acan* [1625], *ptgs2* [1626], *gdf10* [1627], *bdnf* [1628], *ndrg2* [1629], *sgk1* [1630], *tlr3* [1631], *ephb2* [1632], *mbp* [1633], *nox4* [1619], *shank2* [1634], *pi16* [1635], *dlx5* [1636], *vdr* [1637], *zfhx2* [1638], *epha4* [1639], *cyp24a1* [1640], *id2* [1641], *tlr2* [1642], *il6* [1643], *spink1* [1644], *gas6* [1645], *klf2* [1646], *six1* [1647], *chl1* [1648], *slit2* [1649], *runx1* [1650], *notch2* [1651], *pgf* [1652], *nrg1* [1653], *nalc1* [1654], *lxn* [1655] and *trh* [1656] might play a role in the development of pain. Therefore, studying the enriched genes involved in the regulation of endometriosis might be helpful to clarify the incidence or molecular pathogenic mechanisms of various complications including ovarian cancer, endometrial cancer, cervical cancer, hypertension, cardiovascular diseases, infertility, inflammation, polycystic ovarian syndrome, obesity, gestational diabetes mellitus, diabetes mellitus and pain.

Establishing PPI network and module analysis is friendly for researchers to investigate the underlying molecular mechanism of endometriosis for the reason that the DEGs would be grouped and ordered in the network judging by their interactions. PPI network and module analyses could help to find hub genes involved in the regulation of endometriosis. A recent study suggested that the hub genes including *vcam1* [110], *adrb2* [126] and *dapk1* [183] might take part in the progression of endometriosis. Recent evidence indicates that the hub genes including *vcam1* [190], *adrb2* [237], *foxq1* [210], *actb12* [357], *dapk1* [379], *actc1* [359], *cst4* [229], *nfib* [363], *nfix* [1657] and *erg* [1658] are potential therapeutic targets in ovarian cancer. Previous studies have reported that hub genes including *vcam1* [478], *foxq1* [492], *dapk1* [609] and *erg* [1659] participate in the progression of cervical cancer. Studies have shown that hub genes including *vcam1* [615], *adrb2* [653] and *dapk1* [751] are involved in the regulation of hypertension.

Several studies have found hub genes including *vcam1* [761], *prkcb* [780], *adrb2* [808], *prkd1* [920], *dapk1* [947], *actc1* [924] and *nfix* [1660] expression levels were significantly altered in cardiovascular diseases. Studies show that hub genes including *vcam1* [761], *snca* [1026], *adrb2* [1073], *foxq1* [1044], *prkd1* [1188], *dapk1* [947], *actc1* [1192], *cst1* [1030], *nfib* [1198] and *erg* [1661] are mainly involved in progression of inflammation. Previous studies have shown that hub genes including *vcam1* [1232] and *adrb2* [969] were identified to be closely associated with polycystic ovarian syndrome. Many studies have confirmed that hub genes including *vcam1* [1287], *prkcb* [1301] and *adrb2* [1319] were an important participant in obesity. Previous studies have found that hub genes including *vcam1* [1413] and *adrb2* [1425] were shown to be primarily involved in gestational diabetes mellitus. A growing number of studies have demonstrated that hub genes including *vcam1* [1462], *snca* [1468], *prkcb* [1476] and *adrb2* [653] play an important role in progression of diabetes mellitus. Accumulating evidence shows that hub gene *adrb2* [969] is an important risk factor for infertility. Recent study reported that hub gene *adrb2* [1606] plays a crucial role in pain progression. A recent study showed that hub genes including *dapk1* [183], *cst1* [394] and *nfix* [1662] plays an important role in the pathogenesis of endometrial cancer. Our findings suggested *mdfi*, *tnfrsf19* and *foxl1* as potential novel diagnostic biomarkers for endometriosis. This investigation identified the possible hub genes that were highly correlated with the PPI network to find the novel biomarkers associated in the pathogenesis of endometriosis. Our ROC curve analysis showed that hub genes have diagnostic value for endometriosis.

In this investigation, the miRNA-hub gene regulatory network and TF-hub gene regulatory network of the hub genes in endometriosis were analyzed by using miRNet and NetworkAnalyst database. These analyses could help to find some miRNAs, TFs and hub genes involved in the regulation of endometriosis. Studies have shown that biomarkers including *ccnd2* [246], *vcam1* [190], *pdgfb* [221], *ptch1* [263], *foxq1* [210], *igf2bp1* [324], *actc1* [359], *epb41l3* [315], *dapk1* [379], *hsa-mir-17-5p* [1663], *tcf3* [1664], *rnf2* [1665], *clock* [1666], *smarca4* [1667] and *trim28* [1668] can lead to ovarian cancer. Studies reported that biomarkers including *ccnd2* [511], *vcam1* [478], *ptch1* [523], *foxq1* [492], *igf2bp1* [568], *epb41l3* [560], *dapk1* [609], *hsa-mir-17-5p* [1669], *tcf3* [1670] and *trim28* [1671] were proposed to contribute to the development of cervical cancer. A previous study reported that biomarkers including *ccnd2* [814], *vcam1* [615], *pdgfb* [795], *prkcb* [780], *actc1* [924], *dapk1* [947], *hsa-mir-17-5p* [1672], *hsa-mir-2110* [1673], *tcf3* [1674] and

smarca4 [1675] play a key role in cardiovascular diseases. Accumulated evidence has demonstrated that biomarkers including *ccnd2* [1083], *vcam1* [761], *pdgfb* [1059], *ptch1* [1098], *foxq1* [1044], *igf2bp1* [1162], *actc1* [1192], *dapk1* [947], *hsa-mir-2110* [1676], *hsa-mir-10b-5p* [1677], *tcf3* [1678], *nr1i2* [1679] and *trim28* [1680] are associated with inflammation. Studies have shown that biomarkers including *ccnd2* [1495], *vcam1* [1462], *ptprd* [1496], *prkcb* [1476], *igf2bp1* [1545], *hsa-mir-200a-3p* [1681] and *hsa-mir-10b-5p* [1682] were identified to be associated with diabetes mellitus. A previous study found that biomarkers including *vcam1* [110], *pdgfb* [121], *igf2bp1* [162], *dapk1* [183] and *hsa-mir-17-5p* [1683] have been found in endometriosis. Recent studies have identified biomarkers including *vcam1* [1232], *hsa-mir-17-5p* [1684] and *hsa-mir-2110* [1685] are involved in the pathogenesis and progression of polycystic ovarian syndrome. A previous study reported that biomarkers including *vcam1* [1287], *pdgfb* [1311], *prkcb* [1301], *hsa-mir-17-5p* [1686], *hsa-mir-10b-5p* [1687] and *trim28* [1688] are associated with the pathogenesis and development of obesity. Many studies have shown that biomarkers including *vcam1* [1413], *ptprd* [1426] and *hsa-mir-17-5p* [1689] are likely to be important in the development of gestational diabetes mellitus. Recent studies have demonstrated that biomarkers including *igf2bp1* [453], *dapk1* [183], *ptprd* [412], *tcf3* [1690], *smarca4* [1691] and *trim28* [1692] are important in the development of endometrial cancer. Research has shown that biomarkers including *ptprd* [656], *pdgfb* [642], *dapk1* [751], *hsa-mir-4432* [1693], *smarca4* [1694] and *trim28* [1695] might be potential therapeutic targets for hypertension. Recent studies have proposed that the biomarkers including *igf2bp1* [994], *hsa-mir-17-5p* [1696] and *smarca4* [1697] serve a vital role in infertility. Study has suggested that *TRIM28* [1698] might be involved in the development of pain. New biomarkers associated with diagnosis were identified in this study: *st8sia4*, *mdfi*, *krt18*, *stx11*, *hsa-mir-3143*, *hsa-mir-6888-5p*, *hsa-mir-3122*, *hsa-mir-556-3p*, *hsa-mir-1229-5p*, *phc1*, *hoxc9*, *prdm14* and *htt* (huntingtin). We suggest that exercise can regulate the expression of these miRNAs, TFs and hub genes, thereby inhibiting the occurrence and development of endometriosis.

There are few limitations in our investigation. While our investigation presents promising results, several limitations should be accepted. In vivo and in vitro validation experiments for hub genes and clinical trials are required to assess the correlation linking clinical parameters and the hub genes in endometriosis pathogenesis. In this investigation, we did not account for the potential confounding effects of demographic variables. We will conduct more in-depth research in the future.

Conclusions

The current investigation identified biomarkers and pathways which might be involved in endometriosis progression through the integrated analysis of NGS dataset. These results might contribute to a better understanding of the molecular mechanisms which underlie endometriosis and provide a series of potential biomarkers. However, further experiments are required to verify the findings of the current investigations. Therefore, further experiments with additional patient cohorts are also required to confirm the results of these investigations. In vivo and in vitro investigation of gene and pathway interaction is essential to delineate the specific roles of the identified biomarkers, which might help to confirm biomarker functions and reveal the molecular mechanisms underlying endometriosis.

Abbreviations

DEGs	Differentially expressed genes
NGS	Next-generation sequencing
GEO	Gene expression omnibus
GO	Gene ontology
PPI	Protein–protein interaction
miRNA	Micro-ribonucleic acid
TF	Transcription factor
ROC	Receiver operating characteristic curve
VCAM1	Vascular cell adhesion molecule 1
SNCA	Synuclein alpha
PRKCB	Protein kinase C beta
ADRB2	Adrenoceptor beta 2
FOXQ1	Forkhead box Q1
MDF1	MyoD family inhibitor
ACTBL2	Actin beta-like 2
PRKD1	Protein kinase D1
DAPK1	Death-associated protein kinase 1
ACTC1	Actin alpha cardiac muscle 1

Acknowledgements

I thank very much to Peixin Jiang, Baylor College of Medicine, Houston, TX, USA, the author who deposited their NGS dataset GSE243039, into the public GEO database.

Author contributions

BV performed writing original draft and review and editing. CV contributed to software and investigation.

Funding

The authors received no financial support for the research.

Availability of data and materials

The datasets supporting the conclusions of this article are available in the GEO (Gene Expression Omnibus) (<https://www.ncbi.nlm.nih.gov/geo/>) repository. [(GSE243039) <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE243039>].

Declarations

Ethics approval and consent to participate

This article does not contain any studies with human participants or animals performed by any of the authors.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Department of Pharmaceutical Chemistry, K.L.E. College of Pharmacy, Gadag, Karnataka 582101, India. ²Biostatistics and Bioinformatics, Chanabasava Nilaya, Bharthinagar, Dharwad, Karnataka 580001, India.

Received: 18 July 2024 Accepted: 23 August 2024

Published online: 12 October 2024

References

- Czyzyk A, Podfigurna A, Szeliga A, Meczekalski B (2017) Update on endometriosis pathogenesis. *Minerva Ginecol* 69(5):447–461. <https://doi.org/10.23736/S0026-4784.17.04048-5>
- Vercellini P, Viganò P, Somigliana E, Fedele L (2014) Endometriosis: pathogenesis and treatment. *Nat Rev Endocrinol* 10(5):261–275. <https://doi.org/10.1038/nrendo.2013.255>
- Burney RO, Giudice LC (2012) Pathogenesis and pathophysiology of endometriosis. *Fertil Steril* 98(3):511–519. <https://doi.org/10.1016/j.fertnstert.2012.06.029>
- Taylor HS, Kotlyar AM, Flores VA (2021) Endometriosis is a chronic systemic disease: clinical challenges and novel innovations. *Lancet* 397(10276):839–852. [https://doi.org/10.1016/S0140-6736\(21\)00389-5](https://doi.org/10.1016/S0140-6736(21)00389-5)
- Li J, Liu R, Tang S, Feng F, Liu C, Wang L, Zhao W, Zhang T, Yao Y, Wang X et al (2019) Impact of endometriosis on risk of ovarian, endometrial and cervical cancers: a meta-analysis. *Arch Gynecol Obstet* 299(1):35–46. <https://doi.org/10.1007/s00404-018-4968-1>
- Schüler-Toprak S, Ortmann O, Buechler C, Treeck O (2022) The complex roles of adipokines in polycystic ovary syndrome and endometriosis. *Biomedicines* 10(10):2503. <https://doi.org/10.3390/biomedicines10102503>
- Marchandot B, Curtiaud A, Matsushita K, Trimailla A, Host A, Faller E, Garbin O, Akladios C, Jesel L, Morel O (2022) Endometriosis and cardiovascular disease. *Eur Heart J Open* 2(1):oeac001. <https://doi.org/10.1093/ehjopen/oeac001>
- Pantelis A, Machairiotis N, Lapatsanis DP (2021) The formidable yet unresolved interplay between endometriosis and obesity. *Sci World J* 2021:6653677. <https://doi.org/10.1155/2021/6653677>
- Salmeri N, Li Piani L, Cavoretto PI, Somigliana E, Viganò P, Candiani M (2023) Endometriosis increases the risk of gestational diabetes: a meta-analysis stratified by mode of conception, disease localization and severity. *Sci Rep* 13(1):8099. <https://doi.org/10.1038/s41598-023-35236-y>
- Alhallak I, Quick CM, Graham GL, Simmen RCM (2023) A pilot study on the co-existence of diabetes and endometriosis in reproductive-age women: potential for endometriosis progression. *Reprod Sci* 30(8):2429–2438. <https://doi.org/10.1007/s43032-023-01190-3>
- Mu F, Rich-Edwards J, Rimm EB, Spiegelman D, Forman JP, Missmer SA (2017) Association between endometriosis and hypercholesterolemia or hypertension. *Hypertension* 70(1):59–65. <https://doi.org/10.1161/HYPERTENSIONAHA.117.09056>
- Fung JN, Montgomery GW (2018) Genetics of endometriosis: State of the art on genetic risk factors for endometriosis. *Best Pract Res Clin Obstet Gynaecol* 50:61–71. <https://doi.org/10.1016/j.bpobgyn.2018.01.012>
- Coilet E, Courbiere B, Agostini A, Boublil L, Bretelle F, Netter A (2022) Endometriosis and environmental factors: a critical review. *J Gynecol Obstet Hum Reprod* 51(7):102418. <https://doi.org/10.1016/j.jogoh.2022.102418>
- Zanelotti A, Decherney AH (2017) Surgery and endometriosis. *Clin Obstet Gynecol* 60(3):477–484. <https://doi.org/10.1097/GRF.0000000000000291>
- Brown J, Crawford TJ, Datta S, Prentice A (2018) Oral contraceptives for pain associated with endometriosis. *Cochrane Database Syst Rev* 5(5):CD001019. <https://doi.org/10.1002/14651858.CD001019.pub3>
- Vercellini P, Buggio L, Berlanda N, Barbara G, Somigliana E, Bosari S (2016) Estrogen-progestins and progestins for the management of

- endometriosis. *Fertil Steril* 106(7):1552–1571.e2. <https://doi.org/10.1016/j.fertnstert.2016.10.022>
17. Brown J, Crawford TJ, Allen C, Hopewell S, Prentice A (2017) Nonsteroidal anti-inflammatory drugs for pain in women with endometriosis. *Cochrane Database Syst Rev* 1(1):CD004753. <https://doi.org/10.1002/14651858.CD004753.pub4>
 18. Jeng CJ, Chuang L, Shen J (2014) A comparison of progestogens or oral contraceptives and gonadotropin-releasing hormone agonists for the treatment of endometriosis: a systematic review. *Expert Opin Pharmacother* 15(6):767–773. <https://doi.org/10.1517/14656566.2014.888414>
 19. Ahn SH, Singh V, Tayade C (2017) Biomarkers in endometriosis: challenges and opportunities. *Fertil Steril* 107(3):523–532. <https://doi.org/10.1016/j.fertnstert.2017.01.009>
 20. Absenger Y, Hess-Stumpp H, Kreft B, Krätzschar J, Haendler B, Schütze N, Regidor PA, Winterhager E (2004) CYP11A1, a deregulated gene in endometriosis. *Mol Hum Reprod* 10(6):399–407. <https://doi.org/10.1093/molehr/gah053>
 21. Smolarz B, Szyłto K, Romanowicz H (2020) The genetic background of endometriosis: can ESR2 and CYP19A1 genes be a potential risk factor for its development? *Int J Mol Sci* 21(21):8235. <https://doi.org/10.3390/ijms21218235>
 22. Zanatta A, Rocha AM, Carvalho FM, Pereira RM, Taylor HS, Motta EL, Baracat EC, Serafini PC (2010) The role of the Hoxa10/HOXA10 gene in the etiology of endometriosis and its related infertility: a review. *J Assist Reprod Genet* 27(12):701–710. <https://doi.org/10.1007/s10815-010-9471-y>
 23. Mathew D, Drury JA, Valentijn AJ, Vasieva O, Hapangama DK (2016) In silico, in vitro and in vivo analysis identifies a potential role for steroid hormone regulation of FOXD3 in endometriosis-associated genes. *Hum Reprod* 31(2):345–354. <https://doi.org/10.1093/humrep/dev307>
 24. Dentillo DB, Meola J (2010) Deregulation of LOXL1 and HTRA1 gene expression in endometriosis. *Reprod Sci* 17(11):1016–1023. <https://doi.org/10.1177/1933719110377662>
 25. Matsuzaki S, Darcha C (2015) Co-operation between the AKT and ERK signaling pathways may support growth of deep endometriosis in a fibrotic microenvironment in vitro. *Hum Reprod* 30(7):1606–1616. <https://doi.org/10.1093/humrep/dev108>
 26. Matsuzaki S, Darcha C (2013) Involvement of the Wnt/ β -catenin signaling pathway in the cellular and molecular mechanisms of fibrosis in endometriosis. *PLoS ONE* 8(10):e76808. <https://doi.org/10.1371/journal.pone.0076808>
 27. Makker A, Goel MM, Das V, Agarwal A (2012) PI3K-Akt-mTOR and MAPK signaling pathways in polycystic ovarian syndrome, uterine leiomyomas and endometriosis: an update. *Gynecol Endocrinol* 28(3):175–181. <https://doi.org/10.3109/09513590.2011.583955>
 28. Su RW, Strug MR, Joshi NR, Jeong JW, Miele L, Lessey BA, Young SL, Fazleabas AT (2015) Decreased Notch pathway signaling in the endometrium of women with endometriosis impairs decidualization. *J Clin Endocrinol Metab* 100(3):E433–E442. <https://doi.org/10.1210/jc.2014-3720>
 29. Huang F, Cao J, Liu Q, Zou Y, Li H, Yin T (2013) MAPK/ERK signal pathway involved expression of COX-2 and VEGF by IL-1 β induced in human endometriosis stromal cells in vitro. *Int J Clin Exp Pathol* 6(10):2129–2136
 30. Clough E, Barrett T (2016) The gene expression omnibus database. *Methods Mol Biol* 1418:93–110. https://doi.org/10.1007/978-1-4939-3578-9_5
 31. Ganekal P, Vastrad B, Kavatagimath S, Vastrad C, Kotrashetti S (2023) Bioinformatics and next-generation data analysis for identification of genes and molecular pathways involved in subjects with diabetes and obesity. *Medicina (Kaunas)* 59(2):309. <https://doi.org/10.3390/medicina59020309>
 32. Alur V, Raju V, Vastrad B, Vastrad C, Kavatagimath S, Kotturshetti S (2023) Bioinformatics analysis of next generation sequencing data identifies molecular biomarkers associated with type 2 diabetes mellitus. *Clin Med Insights Endocrinol Diabetes* 16:11795514231155636. <https://doi.org/10.1177/11795514231155636>
 33. Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, Smyth GK (2015) limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res* 43(7):e47. <https://doi.org/10.1093/nar/gkv007>
 34. Solari A, Goeman JJ (2017) Minimally adaptive BH: a tiny but uniform improvement of the procedure of Benjamini and Hochberg. *Biom J* 59(4):776–780. <https://doi.org/10.1002/bimj.201500253>
 35. Thomas PD (2017) The gene ontology and the meaning of biological function. *Methods Mol Biol* 1446:15–24. https://doi.org/10.1007/978-1-4939-3743-1_2
 36. Fabregat A, Jupe S, Matthews L, Sidiropoulos K, Gillespie M, Garapati P, Haw R, Jassal B, Korninger F (2018) The reactome pathway knowledgebase. *Nucleic Acids Res* 46(D1):D649–D655. <https://doi.org/10.1093/nar/gkx1132>
 37. Reimand J, Kull M, Peterson H, Hansen J, Vilo J (2007) g:Profiler—a web-based toolset for functional profiling of gene lists from large-scale experiments. *Nucl Acids Res* 35:W193–W200. <https://doi.org/10.1093/nar/gkm226>
 38. Alanis-Lobato G, Andrade-Navarro MA, Schaefer MH (2017) HIPPIE v2.0: enhancing meaningfulness and reliability of protein-protein interaction networks. *Nucl Acids Res* 45:408–414. <https://doi.org/10.1093/nar/gkw985>
 39. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B (2003) Ideker T Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome Res* 13(11):2498–2504. <https://doi.org/10.1101/gr.1239303>
 40. Luo X, Guo L, Dai XJ, Wang Q, Zhu W, Miao X, Gong H (2017) Abnormal intrinsic functional hubs in alcohol dependence: evidence from a voxel-wise degree centrality analysis. *Neuropsychiatr Dis Treat* 13:2011–2020. <https://doi.org/10.2147/NDT.S142742>
 41. Li Y, Li W, Tan Y, Liu F, Cao Y, Lee KY (2017) Hierarchical decomposition for betweenness centrality measure of complex networks. *Sci Rep* 7:46491. <https://doi.org/10.1038/srep46491>
 42. Gilbert M, Li Z, Wu XN, Rohr L, Gombos S, Harter K, Schulze WX (2021) Comparison of path-based centrality measures in protein-protein interaction networks revealed proteins with phenotypic relevance during adaptation to changing nitrogen environments. *J Proteomics* 235:104114. <https://doi.org/10.1016/j.jprot.2021.104114>
 43. Li G, Li M, Wang J, Li Y, Pan Y (2020) United neighborhood closeness centrality and orthology for predicting essential proteins. *IEEE/ACM Trans Comput Biol Bioinform* 17(4):1451–1458. <https://doi.org/10.1109/TCBB.2018.2889978>
 44. Zaki N, Efimov D, Berengueres J (2013) Protein complex detection using interaction reliability assessment and weighted clustering coefficient. *BMC Bioinform* 14:163. <https://doi.org/10.1186/1471-2105-14>
 45. Fan Y, Xia J (2018) miRNet-Functional analysis and visual exploration of miRNA-target interactions in a network context. *Methods Mol Biol* 1819:215–233. https://doi.org/10.1007/978-1-4939-8618-7_10
 46. Zhou G, Soufan O, Ewald J, Hancock REW, Basu N, Xia J (2019) NetworkAnalyst 3.0: a visual analytics platform for comprehensive gene expression profiling and meta-analysis. *Nucl Acids Res* 47:W234–W241. <https://doi.org/10.1093/nar/gkz240>
 47. Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez JC, Müller M (2011) pROC: an open-source package for R and S+ to analyze and compare ROC curves. *BMC Bioinform* 12:77. <https://doi.org/10.1186/1471-2105-12-77>
 48. Frostegård J (2022) The role of PCSK9 in inflammation, immunity, and autoimmune diseases. *Expert Rev Clin Immunol* 18(1):67–74. <https://doi.org/10.1080/1744666X.2022.2017281>
 49. Wang K, Huang XT, Miao YP, Bai XL, Jin F (2022) MiR-148a-3p attenuates apoptosis and inflammation by targeting CNTN4 in atherosclerosis. *Ann Transl Med* 10(22):1201. <https://doi.org/10.21037/atm-22-3768>
 50. Zhang H, Lu Y, Wu B, Xia F (2021) Semaphorin 3A mitigates lipopolysaccharide-induced chondrocyte inflammation, apoptosis and extracellular matrix degradation by binding to Neuropilin-1. *Bioengineered* 12(2):9641–9654. <https://doi.org/10.1080/21655979.2021.1974806>
 51. Zhang J, Yang Z, Liang Z, Wang M, Hu C, Chang C, Shi L, Ji Q, Liu L (2022) Secreted frizzled-related protein 4 exerts anti-atherosclerotic effects by reducing inflammation and oxidative stress. *Eur J Pharmacol* 923:174901. <https://doi.org/10.1016/j.ejphar.2022.174901>
 52. Dong XY, Yin JX, Zhang H, Liao Y (2022) High glucose stimulating ECM remodeling and an inflammatory phenotype in the IPFF via upregulation of MFAP5 expression. *Biochem Biophys Res Commun* 601:93–100. <https://doi.org/10.1016/j.bbrc.2022.02.077>

53. Varga E, Pap R, Jánosa G, Sipos K, Pandur E (2021) IL-6 regulates hepcidin expression via the BMP/SMAD pathway by altering BMP6, TMPRSS5 and TFR2 expressions at normal and inflammatory conditions in BV2 microglia. *Neurochem Res* 46(5):1224–1238. <https://doi.org/10.1007/s11064-021-03322-0>
54. Schöffski P, Concin N, Suarez C, Subbiah V, Ando Y, Ruan S, Wagner JP, Mansfield K, Zhu X, Origuchi S et al (2021) A phase 1 study of a CDH6-targeting antibody-drug conjugate in patients with advanced solid tumors with evaluation of inflammatory and neurological adverse events. *Oncol Res Treat* 44(10):547–556. <https://doi.org/10.1159/000518549>
55. Dubin AE, Schmidt M, Mathur J, Petrus MJ, Xiao B, Coste B, Patapoutian A (2012) Inflammatory signals enhance piezo2-mediated mechanosensitive currents. *Cell Rep* 2(3):511–517. <https://doi.org/10.1016/j.celrep.2012.07.014>
56. Pérez-Hernández M, Marrón-Liñares GM, Schlamp F, Heguy A, van Opbergen CJM, Mezzano V, Zhang M, Liang FX, Cerrone M, Delmar M (2021) Transcriptomic coupling of PKP2 with inflammatory and immune pathways endogenous to adult cardiac myocytes. *Front Physiol* 11:623190. <https://doi.org/10.3389/fphys.2020.623190>
57. Peng J, Liu MM, Jin JL, Cao YX, Guo YL, Wu NQ, Zhu CG, Dong Q, Sun J, Xu RX et al (2022) NAFLD fibrosis score is correlated with PCSK9 and improves outcome prediction of PCSK9 in patients with chest pain: a cohort study. *Lipids Health Dis* 21(1):3. <https://doi.org/10.1186/s12944-021-01610-w>
58. Maeda T, Yamada D, Kawahara K (2016) Cancer pain relief achieved by disrupting tumor-driven semaphorin 3A signaling in mice. *Neurosci Lett* 632:147–151. <https://doi.org/10.1016/j.neulet.2016.08.060>
59. Chen Y, Zhu J, Chen L, Shen Y, Zhang J, Wang Q (2022) SFRP4+IGFBP5hi NKT cells induced neural-like cell differentiation to contribute to adenomyosis pain. *Front Immunol* 13:945504. <https://doi.org/10.3389/fimmu.2022.945504>
60. Jacome Sanz D, Raivola J, Karvonen H, Arjama M, Barker H, Murumägi A, Ungureanu D (2021) Evaluating targeted therapies in ovarian cancer metabolism: novel role for PCSK9 and second generation mTOR inhibitors. *Cancers (Basel)* 13(15):3727. <https://doi.org/10.3390/cancers13153727>
61. Manderson EN, Birch AH, Shen Z, Mes-Masson AM, Provencher D, Tonin PN (2009) Molecular genetic analysis of a cell adhesion molecule with homology to L1CAM, contactin 6, and contactin 4 candidate chromosome 3p26pter tumor suppressor genes in ovarian cancer. *Int J Gynecol Cancer* 19(4):513–525. <https://doi.org/10.1111/IGC.0b013e3181a3cd38>
62. Jiang H, Qi L, Wang F, Sun Z, Huang Z, Xi Q (2015) Decreased semaphorin 3A expression is associated with a poor prognosis in patients with epithelial ovarian carcinoma. *Int J Mol Med* 35(5):1374–1380. <https://doi.org/10.3892/ijmm.2015.2142>
63. Wu X, Lu W, Xu C, Jiang C, Zhang W, Zhang D, Cui S, Zhuo Z, Cui Y, Mei H et al (2023) PTGS1 may be a predictive marker for ovarian cancer by regulating fatty acid metabolism. *Comput Math Methods Med* 2023:2397728. <https://doi.org/10.1155/2023/2397728>
64. Varier L, Sundaram SM, Gamit N, Warriar S (2023) An overview of ovarian cancer: the role of cancer stem cells in chemoresistance and a precision medicine approach targeting the wnt pathway with the antagonist sFRP4. *Cancers (Basel)* 15(4):1275. <https://doi.org/10.3390/cancers15041275>
65. Kujawa KA, Zembala-Nożyńska E, Syrkis JP, Cortez AJ, Kupryjańczyk J, Lisowska KM (2022) Microfibril associated protein 5 (MFAP5) is related to survival of ovarian cancer patients but not useful as a prognostic biomarker. *Int J Mol Sci* 23(24):15994. <https://doi.org/10.3390/ijms232415994>
66. Bartolomé RA, Robles J, Martín-Regalado Á, Pintado-Berninches L, Burdiel M, Jaén M, Aizpurúa C, Imbaud JI, Casal JI (2021) CDH6-activated α 5 β 3 crosstalks with α 2 β 1 to trigger cellular adhesion and invasion in metastatic ovarian and renal cancers. *Mol Oncol* 15(7):1849–1865. <https://doi.org/10.1002/1878-0261.12947>
67. Karapetsas A, Giannakakis A, Dangaj D, Lanitis E, Kynigopoulos S, Lambropoulou M, Tanyi JL, Galanis A, Kakolyris S, Trypsianis G et al (2015) Overexpression of GPC6 and TMEM132D in early stage ovarian cancer correlates with CD8+ T-lymphocyte infiltration and increased patient survival. *Biomed Res Int* 2015:712438. <https://doi.org/10.1155/2015/712438>
68. Gao L, Li X, Guo Q, Nie X, Hao Y, Liu Q, Liu J, Zhu L, Yan L, Lin B (2020) Identification of PKP 2/3 as potential biomarkers of ovarian cancer based on bioinformatics and experiments. *Cancer Cell Int* 20:509. <https://doi.org/10.1186/s12935-020-01602-3>
69. Olaniyi KS, Areloegbe SE, Areola ED, Sabinari IW, Fafure AA, Agbana RD, Atuma CL, Shah MZUH, Ajadi IO, Olatunji LA (2023) Low-dose spironolactone combats dyslipidemia and hepatic inflammation by modulating PCSK9 in rat model of polycystic ovarian syndrome. *Toxicol Appl Pharmacol* 473:116604. <https://doi.org/10.1016/j.taap.2023.116604>
70. Bicer M, Alarслан P, Guler A, Demir I, Aslanipour B, Calan M (2020) Elevated circulating levels of secreted frizzled-related protein 4 in relation to insulin resistance and androgens in women with polycystic ovary syndrome. *J Endocrinol Invest* 43(3):305–313. <https://doi.org/10.1007/s40618-019-01108-4>
71. Xin X, Chang HM, Leung PCK, Dong L, Li J, Lian F, Wu H (2024) Bone morphogenetic protein 6 induces downregulation of pentraxin 3 expression in human granulosa lutein cells in women with polycystic ovary syndrome. *J Assist Reprod Genet* 41(1):31–48. <https://doi.org/10.1007/s10815-023-02972-z>
72. Guo Y, Yan B, Tai S, Zhou S, Zheng XL (2021) PCSK9: Associated with cardiac diseases and their risk factors? *Arch Biochem Biophys* 704:108717. <https://doi.org/10.1016/j.jabb.2020.108717>
73. Li C, Zhao Y, Li F, Wang Z, Qiu Z, Yang Y, Xiong W, Wang R, Chen H, Xu F et al (2023) Semaphorin3A exacerbates cardiac microvascular rarefaction in pressure overload-induced heart disease. *Adv Sci (Weinh)* 10(21):e2206801. <https://doi.org/10.1002/adv.202206801>
74. Zeng W, Cao Y, Jiang W, Kang G, Huang J, Xie S (2019) Knockdown of Sfrp4 attenuates apoptosis to protect against myocardial ischemia/reperfusion injury. *J Pharmacol Sci* 140(1):14–19. <https://doi.org/10.1016/j.jpsh.2019.04.003>
75. Cheng B, Zhong JP, Fu WJ, Chen HJ, Fang L, Li GL, Li JW, Wen MH, Lv YB, Wang HB (2022) I. Microfiber-associated protein 5 (MFAP5): A promising approach to discover new biomarkers for heart failure and cardiac remodeling. *Int J Cardiol* 366:68–69. <https://doi.org/10.1016/j.ijcard.2022.07.008>
76. Lu G, Ge Z, Chen X, Ma Y, Yuan A, Xie Y, Pu J (2023) BMP6 knockdown enhances cardiac fibrosis in a mouse myocardial infarction model by upregulating AP-1/CEMIP expression. *Clin Transl Med* 13(6):e1296. <https://doi.org/10.1002/ctm2.1296>
77. Knight WE, Chen S, Zhang Y, Oikawa M, Wu M, Zhou Q, Miller CL, Cai Y, Mickelsen DM, Moravec C et al (2016) PDE1C deficiency antagonizes pathological cardiac remodeling and dysfunction. *Proc Natl Acad Sci USA* 113(45):E7116–E7125. <https://doi.org/10.1073/pnas.1607728113>
78. Cerrone M, Marrón-Liñares GM, van Opbergen CJM, Costa S, Bourfiss M, Pérez-Hernández M, Schlamp F, Sanchis-Gomar F, Malkani K, Drenkova K et al (2022) Role of plakophilin-2 expression on exercise-related progression of arrhythmogenic right ventricular cardiomyopathy: a translational study. *Eur Heart J* 43(12):1251–1264. <https://doi.org/10.1093/eurheartj/ehab772>
79. Macchi C, Greco MF, Favero C, Dioni L, Cantone L, Hoxha M, Vigna L, Solazzo G, Corsini A, Banach M et al (2022) Associations among PCSK9 levels, atherosclerosis-derived extracellular vesicles, and their mirna content in adults with obesity. *Front Cardiovasc Med* 8:785250. <https://doi.org/10.3389/fcvm.2021.785250>
80. Yiew NKH, Chatterjee TK, Tang YL, Pellenberg R, Stansfield BK, Bagi Z, Fulton DJ, Stepp DW, Chen W, Patel V et al (2017) A novel role for the Wnt inhibitor APCDD1 in adipocyte differentiation: Implications for diet-induced obesity. *J Biol Chem* 292(15):6312–6324. <https://doi.org/10.1074/jbc.M116.758078>
81. Bukhari SA, Yasmin A, Zahoor MA, Mustafa G, Sarfraz I, Rasul A (2019) Secreted frizzled-related protein 4 and its implication in obesity and type-2 diabetes. *IUBMB Life* 71(11):1701–1710. <https://doi.org/10.1002/iub.2123>
82. Vaittinen M, Kolehmainen M, Rydén M, Eskelinen M, Wabitsch M, Pihlajamäki J, Uusitupa M, Pulkkinen L (2015) MFAP5 is related to obesity-associated adipose tissue and extracellular matrix remodeling and inflammation. *Obesity (Silver Spring)* 23(7):1371–1378. <https://doi.org/10.1002/oby.21103>
83. Lluh A, Latorre J, Serena-Maione A, Espadas I, Caballano-Infantes E, Moreno-Navarrete JM, Oliveras-Cañellas N, Ricart W, Malagón MM, Martín-Montalvo A et al (2023) Impaired Plakophilin-2 in obesity breaks

- cell cycle dynamics to breed adipocyte senescence. *Nat Commun* 14(1):5106. <https://doi.org/10.1038/s41467-023-40596-0>
84. Wu Y, Shi J, Su Q, Yang Z, Qin L (2022) Correlation between circulating PCSK9 levels and gestational diabetes mellitus in a chinese population. *Front Endocrinol (Lausanne)* 13:826757. <https://doi.org/10.3389/fendo.2022.826757>
 85. Schuitemaker JHN, Beernink RHJ, Franx A, Cremers TIFH, Koster MPH (2020) First trimester secreted Frizzled-Related Protein 4 and other adipokine serum concentrations in women developing gestational diabetes mellitus. *PLoS ONE* 15(11):e0242423. <https://doi.org/10.1371/journal.pone.0242423>
 86. Hsu LA, Teng MS, Wu S, Chou HH, Ko YL (2022) Common and rare PCSK9 variants associated with low-density lipoprotein cholesterol levels and the risk of diabetes mellitus: a mendelian randomization study. *Int J Mol Sci* 23(18):10418. <https://doi.org/10.3390/ijms231810418>
 87. Qiao Q, Xu X, Song Y, Song S, Zhu W, Li F (2018) Semaphorin 3A promotes osteogenic differentiation of BMSC from type 2 diabetes mellitus rats. *J Mol Histol* 49(4):369–376. <https://doi.org/10.1007/s10735-018-9776-1>
 88. Guo Q, Wang W, Abboud R, Guo Z (2020) Impairment of maturation of BMP-6 (35 kDa) correlates with delayed fracture healing in experimental diabetes. *J Orthop Surg Res* 15(1):186. <https://doi.org/10.1186/s13018-020-01705-7>
 89. Ye P, Jiang XM, Qian WC, Zhang J (2023) Inhibition of PCSK9 improves the development of pulmonary arterial hypertension via down-regulating Notch3 expression. *Cardiovasc Drugs Ther.* <https://doi.org/10.1007/s10557-023-07458-9>
 90. Viazzi F, Ramesh G, Jayakumar C, Leoncini G, Garneri D, Pontremoli R (2015) Increased urine semaphorin-3A is associated with renal damage in hypertensive patients with chronic kidney disease: a nested case-control study. *J Nephrol* 28(3):315–320. <https://doi.org/10.1007/s40620-014-0097-5>
 91. Wang XJ, Xu XQ, Sun K, Liu KQ, Li SQ, Jiang X, Zhao QH, Wang L, Peng FH, Ye J et al (2020) Association of rare PTGIS variants with susceptibility and pulmonary vascular response in patients with idiopathic pulmonary arterial hypertension. *JAMA Cardiol* 5(6):677–684. <https://doi.org/10.1001/jamacardio.2020.0479>
 92. Huo L, Gao Y, Zhang D, Wang S, Han Y, Men H, Yang Z, Qin X, Wang R, Kong D et al (2021) Piezo2 channel in nodose ganglia neurons is essential in controlling hypertension in a pathway regulated directly by Nedd4-2. *Pharmacol Res* 164:105391. <https://doi.org/10.1016/j.phrs.2020.105391>
 93. Pyun JA, Kim S, Cha DH, Kwack K (2013) Epistasis between IGF2R and ADAMTS19 polymorphisms associates with premature ovarian failure. *Hum Reprod* 28(11):3146–3154. <https://doi.org/10.1093/humrep/det365>
 94. De Conto E, Matte U, Cunha-Filho JS (2021) BMP-6 and SMAD4 gene expression is altered in cumulus cells from women with endometriosis-associated infertility. *Acta Obstet Gynecol Scand* 100(5):868–875. <https://doi.org/10.1111/aogs.13931>
 95. Liang Y, Wang W, Huang J, Tan H, Liu T, Shang C, Liu D, Guo L, Yao S (2015) Potential role of semaphorin 3A and its receptors in regulating aberrant sympathetic innervation in peritoneal and deep infiltrating endometriosis. *PLoS ONE* 10(12):e0146027. <https://doi.org/10.1371/journal.pone.0146027>
 96. Lin J, Zong L, Kennedy SH, Zondervan KT (2011) Coding regions of INHBA, SFRP4 and HOXA10 are not implicated in familial endometriosis linked to chromosome 7p13-15. *Mol Hum Reprod* 17(10):605–611. <https://doi.org/10.1093/molehr/gar035>
 97. Wang B, Ge S, Wang Z, Wang W, Wang Y, Leng H, Ma X (2023) Analysis and experimental validation of fatty acid metabolism-related genes prostacyclin synthase (PTGIS) in endometrial cancer. *Aging (Albany NY)* 15(19):10322–10346. <https://doi.org/10.18632/aging.205080>
 98. Guo L, Chen H, Chen J, Gao C, Fu X, Zhou S, Wu W, Li T, Lin J, Yang T et al (2023) PBX1-promoted SFRP4 transcription inhibits cell proliferation and epithelial-mesenchymal transition in endometrial carcinoma. *Tissue Cell* 82:102083. <https://doi.org/10.1016/j.tice.2023.102083>
 99. Zhang H, Chen R, Shao J (2020) MicroRNA-96-5p facilitates the viability, migration, and invasion and suppresses the apoptosis of cervical cancer cells by negatively modulating SFRP4. *Technol Cancer Res Treat* 19:1533033820934132. <https://doi.org/10.1177/1533033820934132>
 100. Li Q, Zhang Y, Jiang Q (2018) MFAP5 suppression inhibits migration/invasion, regulates cell cycle and induces apoptosis via promoting ROS production in cervical cancer. *Biochem Biophys Res Commun* 507(1–4):51–58. <https://doi.org/10.1016/j.bbrc.2018.10.146>
 101. Cook CJ, Wiggin N, Fogg KC (2024) Characterizing the extracellular matrix transcriptome of endometriosis. *Reprod Sci* 31(2):413–429. <https://doi.org/10.1007/s43032-023-01359-w>
 102. Fan P, Li T (2022) Unveil the pain of endometriosis: from the perspective of the nervous system. *Expert Rev Mol Med* 24:e36. <https://doi.org/10.1017/erm.2022.26>
 103. Park S, Lim W, Bazer FW, Song G (2017) Naringenin induces mitochondria-mediated apoptosis and endoplasmic reticulum stress by regulating MAPK and AKT signal transduction pathways in endometriosis cells. *Mol Hum Reprod* 23(12):842–854. <https://doi.org/10.1093/molehr/gax057>
 104. Seki S, Ito K, Takemura N, Oikawa R, Koutake H, Mihara F, Yagi J, Nakanishi M, Tomio K, Oishi H et al (2022) Laparoscopic hemostasis for abdominal blunt massive hemorrhage due to endometriosis. *Asian J Endosc Surg* 15(2):376–379. <https://doi.org/10.1111/ases.13008>
 105. Arena A, Degli Esposti E, Cocchi L, Orsini B, Lenzi J, Del Forno S, Raimondo D, Youssef A, Seracchioli R (2022) Three-dimensional ultrasound evaluation of pelvic floor muscle contraction in women affected by deep infiltrating endometriosis: application of a quick contraction scale. *J Ultrasound Med* 41(12):2973–2979. <https://doi.org/10.1002/jum.15996>
 106. Yamagata Y, Takaki E, Shinagawa M, Okada M, Jozaki K, Lee L, Sato S, Maekawa R, Taketani T, Asada H et al (2015) Retinoic acid has the potential to suppress endometriosis development. *J Ovarian Res* 8:49. <https://doi.org/10.1186/s13048-015-0179-6>
 107. Solkiewicz K, Kacperczyk M, Krotkiewski H, Jędryka M, Kratz EM (2022) O-glycosylation changes in Serum immunoglobulin G are associated with inflammation development in advanced endometriosis. *Int J Mol Sci* 23(15):8087. <https://doi.org/10.3390/ijms23158087>
 108. Silveira CG, Finas D, Hunold P, Köster F, Stroschein K, Canny GO, Moldenhauer G, Altevogt P, Rody A, Hornung D (2013) L1 cell adhesion molecule as a potential therapeutic target in murine models of endometriosis using a monoclonal antibody approach. *PLoS ONE* 8(12):e82512. <https://doi.org/10.1371/journal.pone.0082512>
 109. Osiński M, Wirstlein P, Wender-Ożegowska E, Mikołajczyk M, Jagodziński PP, Szczepańska M (2018) HSD3B2, HSD17B1, HSD17B2, ESR1, ESR2 and AR expression in infertile women with endometriosis. *Ginekol Pol* 89(3):125–134. <https://doi.org/10.5603/GPa2018.0022>
 110. Kuessel L, Wenzl R, Proestling K, Balendran S, Pateisky P (2017) Soluble VCAM-1/soluble ICAM-1 ratio is a promising biomarker for diagnosing endometriosis. *Hum Reprod* 32(4):770–779. <https://doi.org/10.1093/humrep/dex02>
 111. Zhang D, Li Y, Tian J, Zhang H, Wang S (2015) MiR-202 promotes endometriosis by regulating SOX6 expression. *Int J Clin Exp Med* 8(10):17757–17764
 112. Signorile PG, Baldi A, Viceconte R, Vincenzi B, Montella M (2023) Adenogenesis factors FGF7, FGF10, FGF23, IFN- τ and HGF in endometriosis tissue respect to eutopic endometrium: an immunohistochemical study. *Crit Rev Eukaryot Gene Expr* 33(4):85–94. <https://doi.org/10.1615/CritRevEukaryotGeneExpr.2023047178>
 113. Borghese B, Chiche JD, Vernerey D, Chenot C, Mir O, Bijaoui G, Bonaiti-Pellié C, Chapron C (2008) Genetic polymorphisms of matrix metalloproteinase 12 and 13 genes are implicated in endometriosis progression. *Hum Reprod* 23(5):1207–1213. <https://doi.org/10.1093/humrep/den007>
 114. Li T, Wang J, Guo X, Yu Q, Ding S, Xu X, Peng Y, Zhu L, Zou G, Zhang X (2020) Possible involvement of crosstalk between endometrial cells and mast cells in the development of endometriosis via CCL8/CCR1. *Biomed Pharmacother* 129:110476. <https://doi.org/10.1016/j.biopha.2020.110476>
 115. Tiberi F, Tropea A, Apa R, Romani F, Lanzone A, Marana R (2010) Prokineticin 1 mRNA expression in the endometrium of healthy women and in the eutopic endometrium of women with endometriosis. *Fertil Steril* 93(7):2145–2149. <https://doi.org/10.1016/j.fertnstert.2009.01.105>
 116. Lee GJ, Porreca F, Navratilova E (2023) Prolactin and pain of endometriosis. *Pharmacol Ther* 247:108435. <https://doi.org/10.1016/j.pharmthera.2023.108435>

117. Kotronis K, Zafrakas M, Papisozomenou P, Timologou A, Miliaras D, Tarlatzis BC, Grimbizis G (2019) Protein expression pattern of tissue inhibitor of metalloproteinase-3 (TIMP3) in endometriosis and normal endometrium. *Gynecol Endocrinol* 35(12):1103–1106. <https://doi.org/10.1080/09513590.2019.1625880>
118. Wan Y, Gu C, Kong J, Sui J, Zuo L, Song Y, Chen J (2022) Long noncoding RNA ADAMTS9-AS1 represses ferroptosis of endometrial stromal cells by regulating the miR-6516-5p/GPX4 axis in endometriosis. *Sci Rep* 12(1):2618. <https://doi.org/10.1038/s41598-022-04963-z>
119. Jafarabady K, Shafiee A, Bahri RA, Mohammadi I, Amini MJ, Rajai S, Akbarzadeh D, Abhari FS, Movahed E, Parvari S et al (2024) Brain-derived neurotrophic factor (BDNF) as a potential marker of endometriosis: a systematic review and meta-analysis. *BMC Womens Health* 24(1):39. <https://doi.org/10.1186/s12905-023-02877-0>
120. Geng T, Sun Y, Cheng L, Cao Y, Zhang M, Hong Z, Ma L, Zhang Y (2022) Downregulation of LHCGR attenuates COX-2 expression and induces luteinized unruptured follicle syndrome in endometriosis. *Front Endocrinol (Lausanne)* 13:853563. <https://doi.org/10.3389/fendo.2022.853563>
121. Mohagheghian Yaghoubi H, Samadi M, Tajik N, Babaheidarian P, Movahedinia S, Rashidi N, Delbandi AA (2020) Immunomodulatory effects of vitamin D3 on gene expression of MDGF, EGF and PDGFB in endometriosis. *Reprod Biomed Online* 41(5):782–789. <https://doi.org/10.1016/j.rbmo.2020.05.013>
122. Gibran L, Maranhão RC, Tavares ER, Carvalho PO, Abrão MS, Podgac S (2017) mRNA levels of low-density lipoprotein receptors are overexpressed in the foci of deep bowel endometriosis. *Hum Reprod* 32(2):332–339. <https://doi.org/10.1093/humrep/dew303>
123. Pashizeh F, Mansouri R, Davari-Tanha F et al (2020) Alterations of CD4+T cell subsets in blood and peritoneal fluid in different stages of endometriosis. *Int J Fertil Steril* 14(3):201–208. <https://doi.org/10.22074/ijfs.2020.6127>
124. Governini L, Carrarelli P, Rocha AL, Leo VD, Luddi A, Arcuri F, Piomboni P, Chapron C, Bilezikjian LM, Petraglia F (2014) FOXL2 in human endometrium: hyperexpressed in endometriosis. *Reprod Sci* 21(10):1249–1255. <https://doi.org/10.1177/1933719114522549>
125. Zhu H, Wang Y, He Y, Yu W (2022) Inflammation-mediated macrophage polarization induces TRPV1/TRPA1 heteromers in endometriosis. *Am J Transl Res* 14(5):3066–3078
126. Long Q, Liu X, Qi Q, Guo SW (2016) Chronic stress accelerates the development of endometriosis in mouse through adrenergic receptor β_2 . *Hum Reprod* 31(11):2506–2519. <https://doi.org/10.1093/humrep/dew237>
127. Sillem M, Pifti S, Monga B, Buvari P, Shamia U, Runnebaum B (1997) Soluble urokinase-type plasminogen activator receptor is over-expressed in uterine endometrium from women with endometriosis. *Mol Hum Reprod* 3(12):1101–1105. <https://doi.org/10.1093/molehr/3.12.1101>
128. Liu D, Yang N, Liang Y, Chen M, Yang F, Liu L, Yao S (2020) Increased expression of epithelial cell adhesion molecule and its possible role in epithelial-mesenchymal transition in endometriosis. *J Obstet Gynaecol Res* 46(10):2066–2075. <https://doi.org/10.1111/jog.14401>
129. Novembri R, Carrarelli P, Toti P, Rocha AL, Borges LE, Reis FM, Piomboni P, Florio P, Petraglia F (2011) Urocortin 2 and urocortin 3 in endometriosis: evidence for a possible role in inflammatory response. *Mol Hum Reprod* 17(9):587–593. <https://doi.org/10.1093/molehr/gar020>
130. Fan W, Huang Z, Xiao Z, Li S, Ma Q (2016) The cytochrome P4501A1 gene polymorphisms and endometriosis: a meta-analysis. *J Assist Reprod Genet* 33(10):1373–1383. <https://doi.org/10.1007/s10815-016-0783-4>
131. Guo X, Ding S, Li T, Wang J, Yu Q, Zhu L, Xu X, Zou G, Peng Y, Zhang X (2020) Macrophage-derived netrin-1 is critical for neuroangiogenesis in endometriosis. *Int J Biol Macromol* 148:226–237. <https://doi.org/10.1016/j.ijbiomac.2020.01.130>
132. Belleis P, Frediani Barbeiro D, Gueuvoghlian-Silva BY, Kalil J, Abrão MS, Podgac S (2019) Interleukin-15 and interleukin-7 are the major cytokines to maintain endometriosis. *Gynecol Obstet Invest* 84(5):435–444. <https://doi.org/10.1159/000496607>
133. Janusz J, Janusz A, Kondera-Anasz Z, Sikora J, Smycz-Kubańska M, Englisz A, Wendlocha D, Mielczarek-Palacz A (2021) Participation of selected soluble BMP-2 and BMP-7 bone morphogenetic proteins and their soluble type I ALK-1 and type II BMPRII receptors in formation and development of endometriosis. *Biomedicines* 9(10):1292. <https://doi.org/10.3390/biomedicines9101292>
134. Liu YJ, Xing F, Zong K, Wang MY, Ji DM, Zhao YH, Xia YH, Wang A, Shi LG, Ding SM et al (2021) Increased ApoE expression in follicular fluid and the ApoE genotype are associated with endometriosis in chinese women. *Front Endocrinol (Lausanne)* 12:779183. <https://doi.org/10.3389/fendo.2021.779183>
135. Hang Y, Tan L, Chen Q, Liu Q, Jin Y (2021) E3 ubiquitin ligase TRIM24 deficiency promotes NLRP3/caspase-1/IL-1 β -mediated pyroptosis in endometriosis. *Cell Biol Int* 45(7):1561–1570. <https://doi.org/10.1002/cbin.11592>
136. Liu S, Zhou J, Wen J (2015) Expression and significance of CD133 and ABCG2 in endometriosis. *Clin Exp Obstet Gynecol* 42(6):771–775
137. Kowalczyńska LJ, Ferenc T, Wojciechowski M, Mordalska A, Pogoda K, Malinowski A (2014) Endometriosis and RAS system gene polymorphisms: the association of ACE A2350G polymorphism with endometriosis in Polish individuals. *DNA Cell Biol* 33(5):328–335. <https://doi.org/10.1089/dna.2013.2255>
138. Reis FM, Coutinho LM, Vannuccini S, Batteux F, Chapron C, Petraglia F (2020) Progesterone receptor ligands for the treatment of endometriosis: the mechanisms behind therapeutic success and failure. *Hum Reprod Update* 26(4):565–585. <https://doi.org/10.1093/humupd/dmaa009>
139. Kang JO, Hudak WA, Crowley WJ, Criswell BS (1990) Placental-type alkaline phosphatase in peritoneal fluid of women with endometriosis. *Clin Chim Acta* 186(2):285–294. [https://doi.org/10.1016/0009-8981\(90\)90046-u](https://doi.org/10.1016/0009-8981(90)90046-u)
140. Kowalczyk-Zieba I, Woclawek-Potocka I, Wasniewski T, Boruszewska D, Grycmacher K, Sinderewicz E, Staszkievicz J, Wolczynski S (2019) LPAR2 and LPAR4 are the main receptors responsible for LPA actions in ovarian endometriotic cysts. *Reprod Sci* 26(1):139–150. <https://doi.org/10.1177/1933719118766263>
141. Chen X, Liu M (2022) CircATRNL1 increases acid-sensing ion channel 1 to advance epithelial-mesenchymal transition in endometriosis by binding to microRNA-103a-3p. *Reprod Biol* 22(2):100643. <https://doi.org/10.1016/j.repbio.2022.100643>
142. Chou YC, Chen CH, Chen MJ, Chang CW, Chen PH, Yu MH, Chen YJ, Tsai EM, Yang PS, Lin SY et al (2020) Killer cell immunoglobulin-like receptors (KIR) and human leukocyte antigen-C (HLA-C) allorecognition patterns in women with endometriosis. *Sci Rep* 10(1):4897. <https://doi.org/10.1038/s41598-020-61702-y>
143. Muharam R, Rahmala Febri R, Mutia K, Iffanolida PA, Maidarti M, Wiweko B, Hestiantoro A (2023) Down-regulation of miR-93 negatively correlates with overexpression of VEGFA and MMP3 in endometriosis: a cross-sectional study. *Int J Fertil Steril* 17(1):28–33. <https://doi.org/10.22074/ijfs.2022.543884.1233>
144. Gan L, Sun J, Sun J (2022) Bioinformatical analysis identifies PDLIM3 as a potential biomarker associated with immune infiltration in patients with endometriosis. *PeerJ* 10:e13218. <https://doi.org/10.7717/peerj.13218>
145. Chen P, Yao M, Fang T, Ye C, Du Y, Jin Y, Wu R (2022) Identification of NFASC and CHL1 as two novel hub genes in endometriosis using integrated bioinformatic analysis and experimental verification. *Pharmgenom Pers Med* 15:377–392. <https://doi.org/10.2147/PGPM.S354957>
146. He B, Teng XM, Hao F, Zhao M, Chen ZQ, Li KM, Yan Q (2022) Decreased intracellular IL-33 impairs endometrial receptivity in women with adenomyosis. *Front Endocrinol (Lausanne)* 13:928024. <https://doi.org/10.3389/fendo.2022.928024>
147. Liu D, Liu M, Yu P, Li H (2023) Brain-derived neurotrophic factor and nerve growth factor expression in endometriosis: a systematic review and meta-analysis. *Taiwan J Obstet Gynecol* 62(5):634–639. <https://doi.org/10.1016/j.tjog.2023.07.003>
148. Janša V, Klančič T, Pušič M, Klein M, Vrtačnik Bokal E, Ban Frangež H, Rižner TL (2021) Proteomic analysis of peritoneal fluid identified COMP and TGFBI as new candidate biomarkers for endometriosis. *Sci Rep* 11(1):20870. <https://doi.org/10.1038/s41598-021-00299-2>
149. Florio P, Reis FM, Torres PB, Calonaci F, Abrão MS, Nascimento LL, Franchini M, Cianferoni L, Petraglia F (2009) High serum follistatin levels in women with ovarian endometriosis. *Hum Reprod* 24(10):2600–2606. <https://doi.org/10.1093/humrep/dep195>

150. Wan Y, Song Y, Chen J, Kong J, Gu C, Huang J, Zuo L (2022) Upregulated fibulin-1 increased endometrial stromal cell viability and migration by repressing EFEMP1-dependent ferroptosis in endometriosis. *Biomed Res Int* 2022:4809415. <https://doi.org/10.1155/2022/4809415>
151. Cheng W, Shan J, Ding J, Liu Y, Sun S, Xu L, Yu C (2023) herapeutic effects of Huayu Jiedu formula on endometriosis via downregulating GATA 6 expression. *Heliyon* 10(1):e23149. <https://doi.org/10.1016/j.heliyon.2023.e23149>
152. Zhu J, Xu Z, Wu P, Zeng C, Peng C, Zhou Y, Xue Q (2023) MicroRNA-92a-3p inhibits cell proliferation and invasion by regulating the transcription factor 21/steroidogenic factor 1 axis in endometriosis. *Reprod Sci* 30(7):2188–2197. <https://doi.org/10.1007/s43032-021-00734-9>
153. da Luz CM, da Broi MG, Donabela FC, Paro de Paz CC, Meola J, Navarro PA (2017) PTGS2 down-regulation in cumulus cells of infertile women with endometriosis. *Reprod Biomed Online* 35(4):379–386. <https://doi.org/10.1016/j.rbmo.2017.06.021>
154. Mihara Y, Maekawa R, Sato S, Shimizu N, Doi-Tanaka Y, Takagi H, Shirafuta Y, Shinagawa M, Tamura I, Taketani T et al (2020) An integrated genomic approach identifies HOXC8 as an upstream regulator in ovarian endometrioma. *J Clin Endocrinol Metab* 105(12):dgaa618. <https://doi.org/10.1210/clinem/dgaa618>
155. Rižner TL, Penning TM (2020) Aldo-keto reductase 1C3-assessment as a new target for the treatment of endometriosis. *Pharmacol Res* 152:104446. <https://doi.org/10.1016/j.phrs.2019.104446>
156. Xu H, Gao Y, Shu Y, Wang Y, Shi Q (2019) EPHA3 enhances macrophage autophagy and apoptosis by disrupting the mTOR signaling pathway in mice with endometriosis. *Biosci Rep* 39(7):BSR20182274. <https://doi.org/10.1042/BSR20182274>
157. Lin J, Zong L, Kennedy SH, Zondervan KT (2011) Coding regions of INHBA, SFRP4 and HOXA10 are not implicated in familial endometriosis linked to chromosome 7p13-15. *Mol Hum Reprod* 17(10):605–611. <https://doi.org/10.1093/molehr/gar035>
158. Dehghanian M, Yarahmadi G, Sandoghsaz RS, Khodadadian A, Shamsi F, Vahidi Mehrjardi MY (2023) Evaluation of Rap1GAP and EPAC1 gene expression in endometriosis disease. *Adv Biomed Res* 12:101. https://doi.org/10.4103/abr.abr_86_22
159. Almasi MZ, Hosseini E, Jafari R, Aflatoonian K, Aghajanzpour S, Ramazanali F, Moini A, Shahhoseini M, Afsharian P, Aflatoonian R (2021) Evaluation of Toll-like receptor 3 (TLR3) signaling pathway genes and its genetic polymorphisms in ectopic and eutopic endometrium of women with endometriosis. *J Gynecol Obstet Hum Reprod* 50(9):102153. <https://doi.org/10.1016/j.jogoh.2021.102153>
160. Wang X, Jiang X, Lv X, Wang X, Lin A, Li Y (2024) NADPH oxidase 4-mediated oxidative stress contributes to endometriosis. *J Appl Genet* 65(1):113–120. <https://doi.org/10.1007/s13353-023-00810-7>
161. Janša V, Pušić Novak M, Ban Frangež H, Rižner TL (2023) TGFBI as a candidate biomarker for non-invasive diagnosis of early-stage endometriosis. *Hum Reprod* 38(7):1284–1296. <https://doi.org/10.1093/humrep/dead091>
162. Huang Z, Shen F, Chen J, Xie B, Chen X, Zhao Y, Chen S (2024) LncRNA linc01194 promotes the progress of endometrial carcinoma by up-regulating SOX2 through binding to IGF2BP1. *J Gynecol Oncol* 35(2):e21. <https://doi.org/10.3802/jgo.2024.35.e21>
163. Bellessort B, Le Cardinal M, Bachelot A, Narboux-Nême N, Garagnani P, Pirazzini C, Barbieri O, Mastracci L, Jonchere V, Duvernois-Berthet E et al (2016) Dlx5 and Dlx6 control uterine adenogenesis during post-natal maturation: possible consequences for endometriosis. *Hum Mol Genet* 25(1):97–108. <https://doi.org/10.1093/hmg/ddv452>
164. Jafari M, Khodaverdi S, Sadri M, Moradi Z, Mohammadi T, Heidari S, Akhavan Sales Z, Delbandi AA (2021) Association between vitamin D receptor (VDR) and vitamin D binding protein (VDBP) genes polymorphisms to endometriosis susceptibility in Iranian women. *Reprod Sci* 28(12):3491–3497. <https://doi.org/10.1007/s43032-021-00598-z>
165. Lan S, Zhang Z, Li Q (2023) FZD7: a potential biomarker for endometriosis. *Medicine (Baltimore)* 102(40):e35406. <https://doi.org/10.1097/MD.00000000000035406>
166. Araujo FM, Meola J, Rosa-E-Silva JC, Paz CCP, Ferriani RA, Nogueira AA (2017) Increased expression of ID2, PRELP and SMOC2 genes in patients with endometriosis. *Braz J Med Biol Res* 50(7):e5782. <https://doi.org/10.1590/1414-431X20175782>
167. Sobstyl M, Niedźwiedzka-Rystwej P, Grywalska E, Korona-Głowniak I, Sobstyl A, Bednarek W, Roliński J (2020) Toll-like receptor 2 expression as a new hallmark of advanced endometriosis. *Cells* 9(8):1813. <https://doi.org/10.3390/cells9081813>
168. Burns KA, Thomas SY, Hamilton KJ, Young SL, Cook DN, Korach KS (2018) Early endometriosis in females is directed by immune-mediated estrogen receptor α and IL-6 cross-talk. *Endocrinology* 159(1):103–118. <https://doi.org/10.1210/en.2017-00562>
169. Sun WS, Misao R, Iwagaki S, Fujimoto J, Tamaya T (2002) Coexpression of growth arrest-specific gene 6 and receptor tyrosine kinases, Axl and Sky, in human uterine endometrium and ovarian endometriosis. *Mol Hum Reprod* 8(6):552–558. <https://doi.org/10.1093/molehr/8.6.552>
170. Hsiao KY, Chang N, Tsai JL, Lin SC, Tsai SJ, Wu MH (2017) Hypoxia-inhibited DUSP2 expression promotes IL-6/STAT3 signaling in endometriosis. *Am J Reprod Immunol* 78(4):e12690. <https://doi.org/10.1111/aji.12690.10.1111/aji.12690>
171. Signorile PG, Baldi A, Viceconte R, Vincenzi B, Montella M (2023) Adenogenesis factors FGF7, FGF10, FGF23, IFN- τ and HGF in endometriosis tissue respect to eutopic endometrium: an immunohistochemical study. *Crit Rev Eukaryot Gene Expr* 33(4):85–94. <https://doi.org/10.1615/CritRevEukaryotGeneExpr.2023047178>
172. Zhang Y, Chang X, Wu D, Deng M, Miao J, Jin Z (2021) Down-regulation of exosomal miR-214-3p targeting CCN2 contributes to endometriosis fibrosis and the role of exosomes in the horizontal transfer of miR-214-3p. *Reprod Sci* 28(3):715–727. <https://doi.org/10.1007/s43032-020-00350-z>
173. Kai K, Joshi NR, Burns GW, Hrbek SM, Vegter EL, Ochoa-Bernal MA, Song Y, Moldovan GE, Sempere LF, Miyadahira EH et al (2023) MicroRNA-210-3p regulates endometriotic lesion development by targeting IGFBP3 in baboons and women with endometriosis. *Reprod Sci* 30(10):2932–2944. <https://doi.org/10.1007/s43032-023-01253-5>
174. Chen P, Yao M, Fang T, Ye C, Du Y, Jin Y, Wu R (2022) Identification of NFASC and CHL1 as two novel hub genes in endometriosis using integrated bioinformatic analysis and experimental verification. *Pharmacogenomics Pers Med* 15:377–392. <https://doi.org/10.2147/PGPM.S354957>
175. Kocbek V, Hevir-Kene N, Bersinger NA, Mueller MD, Rižner TL (2014) Increased levels of biglycan in endometriomas and peritoneal fluid samples from ovarian endometriosis patients. *Gynecol Endocrinol* 30(7):520–524. <https://doi.org/10.3109/09513590.2014.898055>
176. Lee HC, Lin SC, Wu MH, Tsai SJ (2021) Inhibiting NTRK2 signaling causes endometriotic lesion regression. *Reproduction* 161(1):11–19. <https://doi.org/10.1530/REP-20-0163>
177. Guo SW, Zheng Y, Lu Y, Liu X, Geng JG (2013) Slit2 overexpression results in increased microvessel density and lesion size in mice with induced endometriosis. *Reprod Sci* 20(3):285–298. <https://doi.org/10.1177/1933719112452940>
178. Xu A, Jiang M, Li S, Fei Q (2020) Down-regulation of circ_0061140 attenuates ectopic endometrial cell proliferation, migration and invasion in endometriosis via inactivating Notch2. *Gene* 757:144926. <https://doi.org/10.1016/j.gene.2020.144926>
179. Moberg C, Bourlev V, Ilyasova N, Olovsson M (2015) Endometrial expression of LIF and its receptor and peritoneal fluid levels of IL-1 α and IL-6 in women with endometriosis are associated with the probability of pregnancy. *Arch Gynecol Obstet* 292(2):429–437. <https://doi.org/10.1007/s00404-015-3626-0>
180. Hamilton M, Turpin V, Ayoub A, Reihani A, Arredondo J, Ask K, Clark DA, Foster WG (2023) Circulating CD200 is increased in the secretory phase of women with endometriosis as is endometrial mRNA, and endometrial stromal cell CD200R1 is increased in spite of reduced mRNA. *Am J Reprod Immunol* 89(1):e13655. <https://doi.org/10.1111/aji.13655>
181. Jiang L, Wang S, Xia X, Zhang T, Wang X, Zeng F, Ma J, Fang X (2024) Novel diagnostic biomarker BST2 identified by integrated transcriptomics promotes the development of endometriosis via the TNF- α /NF- κ B signaling pathway. *Biochem Genet*. <https://doi.org/10.1007/s10528-024-10666-z>
182. Peng LS, Li ZM, Chen G, Liu FY, Luo Y, Guo JB, Gao GD, Deng YH, Xu LX, Zhou JY et al (2021) Frequent DYSF rare variants/mutations in 152 Han Chinese samples with ovarian endometriosis. *Arch Gynecol Obstet* 304(3):671–677. <https://doi.org/10.1007/s00404-021-06094-8>

183. Tian X, Xu L, Wang P (2015) MiR-191 inhibits TNF- α induced apoptosis of ovarian endometriosis and endometrioid carcinoma cells by targeting DAPK1. *Int J Clin Exp Pathol* 8(5):4933–4942
184. Timologou A, Zafarakas M, Grimbizis G, Miliaras D, Kotronis K, Stamatopoulos P, Tarlatzis BC (2016) Immunohistochemical expression pattern of metastasis suppressors KAI1 and KISS1 in endometriosis and normal endometrium. *Eur J Obstet Gynecol Reprod Biol* 199:110–115. <https://doi.org/10.1016/j.ejogrb.2016.02.004>
185. Fusco R, D'amico R, Cordaro M, Gugliandolo E, Siracusa R, Peritore AF, Crupi R, Impellizzeri D, Cuzzocrea S, Di Paola R et al (2018) Absence of formyl peptide receptor 1 causes endometriotic lesion regression in a mouse model of surgically-induced endometriosis. *Oncotarget* 9(59):31355–31366. <https://doi.org/10.18632/oncotarget.258>
186. Cunha-Filho JS, Gross JL, Lemos NA, Dias EC, Vettori D, Souza CA, Passos EP (2002) Prolactin and growth hormone secretion after thyrotrophin-releasing hormone infusion and dopaminergic (DA2) blockade in infertile patients with minimal/mild endometriosis. *Hum Reprod* 17(4):960–965. <https://doi.org/10.1093/humrep/17.4.960>
187. Doberstein K, Spivak R, Reavis HD, Hooda J, Feng Y, Kroeger PT Jr, Stuckelberger S, Mills GB, Devins KM, Schwartz LE et al (2022) L1CAM is required for early dissemination of fallopian tube carcinoma precursors to the ovary. *Commun Biol* 5(1):1362. <https://doi.org/10.1038/s42003-022-04314-8>
188. Xie W, Wang W, Meng S, Wu X, Liu X, Liu Y, Kang X, Su Y, Lv X, Guo L et al (2023) A novel hypoxia-stimulated lncRNA HIF1A-AS3 binds with YBX1 to promote ovarian cancer tumorigenesis by suppressing p21 and AJAP1 transcription. *Mol Carcinog* 62(12):1860–1876. <https://doi.org/10.1002/mc.23620>
189. Plourde M, Manhes C, Leblanc G, Durocher F, Dumont M, Sinilnikova O (2008) Mutation analysis and characterization of HSD17B2 sequence variants in breast cancer cases from French Canadian families with high risk of breast and ovarian cancer. *J Mol Endocrinol* 40(4):161–172. <https://doi.org/10.1677/JME-07-0101>
190. Song J, Sokoll LJ, Zhang Z, Chan DW (2023) VCAM-1 complements CA-125 in detecting recurrent ovarian cancer. *Clin Proteom* 20(1):25. <https://doi.org/10.1186/s12014-023-09414-z>
191. Jia Y, Shi H, Fan D (2015) Significance of gastrin-releasing peptide in ovarian cancer ES2 cells. *Oncol Lett* 10(1):359–363. <https://doi.org/10.3892/ol.2015.3240>
192. Ma J, Zhou C, Yang J, Ding X, Zhu Y, Chen X (2016) Expression of AQP6 and AQP8 in epithelial ovarian tumor. *J Mol Histol* 47(2):129–134. <https://doi.org/10.1007/s10735-016-9657-4>
193. Bao H, Wu W, Li Y, Zong Z, Chen S (2022) WNT6 participates in the occurrence and development of ovarian cancer by upregulating/activating the typical Wnt pathway and Notch1 signaling pathway. *Gene* 846:146871. <https://doi.org/10.1016/j.gene.2022.146871>
194. Mukherjee A, Chiang CY, Dafotis HA, Nieman KM, Fahrman JF, Lastra RR, Romero IL, Fiehn O, Lengyel E (2020) Adipocyte-induced FABP4 expression in ovarian cancer cells promotes metastasis and mediates carboplatin resistance. *Cancer Res* 80(8):1748–1761. <https://doi.org/10.1158/0008-5472.CAN-19-1999>
195. Chapel DB, Hirsch MS (2022) SOX6 expression is sensitive for peritoneal epithelioid malignant mesothelioma, but not specific in the differential diagnosis with tubo-ovarian serous neoplasia. *Am J Surg Pathol* 46(2):213–219. <https://doi.org/10.1097/PAS.0000000000001792>
196. Endo Y, Watanabe T, Saito M, Saito K, Suzuki R, Sano H, Natori Y, Sasaki E, Ueda M, Kamo N et al (2022) A rare case of recurrent ovarian cancer with TPM3-NTRK1 gene rearrangement: a case report. *Mol Clin Oncol* 16(4):90. <https://doi.org/10.3892/mco.2022.2523>
197. Han Y, You J, Han Y, Liu Y, Huang M, Lu X, Chen J, Zheng Y (2021) LINC00184 promotes ovarian cancer cells proliferation and cisplatin resistance by elevating CNTN1 expression via sponging miR-1305. *Onco Targets Ther* 14:2711–2726. <https://doi.org/10.2147/OTT.S280490>
198. Li Y, Jia JH, Kang S, Zhang XJ, Zhao J, Wang N, Zhou RM, Sun DL, Duan YN, Wang DJ (2009) The functional polymorphisms on promoter region of matrix metalloproteinase-12, -13 genes may alter the risk of epithelial ovarian carcinoma in Chinese. *Int J Gynecol Cancer* 19(1):129–133. <https://doi.org/10.1111/IGC.0b013e31819a1d8e>
199. Eurich K, De La Cruz P, Laguna A, Woodman M, McAdams J, Lips E, Ebbott J, DiSilvestro J, Ribeiro J, James N (2023) Multiplex serum immune profiling reveals circulating LAG-3 is associated with improved patient survival in high grade serous ovarian cancer. *Gynecol Oncol* 174:200–207. <https://doi.org/10.1016/j.ygyno.2023.05.015>
200. Lawrenson K, Fonseca MAS, Liu AY, Segato Dezem F, Lee JM, Lin X, Corona RI, Abbasi F, Vavra KC, Dinh HQ et al (2019) A study of high-grade serous ovarian cancer origins implicates the SOX18 transcription factor in tumor development. *Cell Rep* 29(11):3726–3735.e4. <https://doi.org/10.1016/j.celrep.2019.10.122>
201. Krishnan V, Tallapragada S, Schaar B, Kamat K, Chanana AM, Zhang Y, Patel S, Parkash V, Rinker-Schaeffer C, Folkins AK et al (2020) Omental macrophages secrete chemokine ligands that promote ovarian cancer colonization of the omentum via CCR1. *Commun Biol* 3(1):524. <https://doi.org/10.1038/s42003-020-01246-z>
202. Takei Y, Mizukami H, Saga Y, Yoshimura I, Hasumi Y, Takayama T, Kohno T, Matsushita T, Okada T, Kume A et al (2007) Suppression of ovarian cancer by muscle-mediated expression of soluble VEGFR-1/Flt-1 using adeno-associated virus serotype 1-derived vector. *Int J Cancer* 120(2):278–284. <https://doi.org/10.1002/ijc.22307>
203. Mitamura T, Zhai T, Hatanaka KC, Hatanaka Y, Amano T, Wang L, Tanaka S, Watari H (2022) Germline PRDM1 Variant rs2185379 in long-term recurrence-free survivors of advanced ovarian cancer. *Pharmgenom Pers Med* 15:977–984. <https://doi.org/10.2147/PGPM.S387120>
204. Shen Z, Gu L, Liu Y, Wang L, Zhu J, Tang S, Wei X, Wang J, Zhang S, Wang X et al (2022) PLAA suppresses ovarian cancer metastasis via METTL3-mediated m6A modification of TRPC3 mRNA. *Oncogene* 41(35):4145–4158. <https://doi.org/10.1038/s41388-022-02411-w>
205. Fraungruber P, Kaltofen T, Heublein S, Kuhn C, Mayr D, Burges A, Mahner S, Rathert P, Jeschke U, Trillsch F (2021) G protein-coupled estrogen receptor correlates with Dkk2 expression and has prognostic impact in ovarian cancer patients. *Front Endocrinol (Lausanne)* 12:564002. <https://doi.org/10.3389/fendo.2021.564002>
206. Xu P, Xu S, Pan H, Dai C, Xu Y, Wang L, Cong Y, Zhang H, Cao J, Ge L et al (2023) Differential effects of the lncRNA RNF157-AS1 on epithelial ovarian cancer cells through suppression of DIRAS3- and ULK1-mediated autophagy. *Cell Death Dis* 14(2):140. <https://doi.org/10.1038/s41419-023-05668-5>
207. Fuller PJ, Alexiadis M, Jobling T, McNeilage J (2005) Seladin-1/DHCR24 expression in normal ovary, ovarian epithelial and granulosa tumours. *Clin Endocrinol (Oxf)* 63(1):111–115. <https://doi.org/10.1111/j.1365-2265.2005.02308.x>
208. Coffman LG, Choi YJ, McLean K, Allen BL, di Magliano MP, Buckanovich RJ (2016) Human carcinoma-associated mesenchymal stem cells promote ovarian cancer chemotherapy resistance via a BMP4/HH signaling loop. *Oncotarget* 7(6):6916–6932. <https://doi.org/10.18632/oncotarget.6870>
209. Alkharusi A, AlMuslahi A, AlBalushi N, AlAjmi R, AlRawahi S, AlFarqani A, Norstedt G, Zadjali F (2021) Connections between prolactin and ovarian cancer. *PLoS ONE* 16(8):e0255701. <https://doi.org/10.1371/journal.pone.0255701>
210. Wu J, Wu Y, Chen S, Guo Q, Shao Y, Liu C, Lin K, Wang S, Zhu J, Chen X et al (2024) PARP1-stabilised FOXQ1 promotes ovarian cancer progression by activating the LAMB3/WNT/ β -catenin signalling pathway. *Oncogene* 43(12):866–883. <https://doi.org/10.1038/s41388-024-02943-3>
211. Zhou W, Mei J, Gu D, Xu J, Wang R, Wang H, Liu C (2021) Wnt5a: A promising therapeutic target in ovarian cancer. *Pathol Res Pract* 219:153348. <https://doi.org/10.1016/j.prp.2021.153348>
212. Li J, Sun Y, Zhi X, Sun Y, Abudousalamu Z, Lin Q, Li B, Yao L, Chen M (2024) Unraveling the molecular mechanisms of lymph node metastasis in ovarian cancer: focus on MEOX1. *J Ovarian Res* 17(1):61. <https://doi.org/10.1186/s13048-024-01384-6>
213. Zhao Q, Zhong J, Lu P, Feng X, Han Y, Ling C, Guo W, Zhou W, Yu F (2021) DOCK4 is a platinum-chemosensitive and prognostic-related biomarker in ovarian cancer. *PPAR Res* 2021:6629842. <https://doi.org/10.1155/2021/6629842>
214. Hakamy S, Assidi M, Jafri MA, Nedjati T, Alkhatabi H, Al-Qahtani A, Al-Maghrabi J, Sait K, Al-Qahtani M, Buhmeida A et al (2021) Assessment of prognostic value of tissue inhibitors of metalloproteinase 3 (TIMP3) protein in ovarian cancer. *Libyan J Med* 16(1):1937866. <https://doi.org/10.1080/19932820.2021.1937866>
215. Cai L, Hu X, Ye L, Bai P, Jie Y, Shu K (2022) Long non-coding RNA ADAMTS9-AS1 attenuates ferroptosis by Targeting microRNA-587/

- solute carrier family 7 member 11 axis in epithelial ovarian cancer. *Bioengineered* 13(4):8226–8239. <https://doi.org/10.1080/21655979.2022.2049470>
216. Xu Y, Jiang WG, Wang HC, Martin T, Zeng YX, Zhang J, Qi Y (2019) BDNF activates TrkB/PLC γ 1 signaling pathway to promote proliferation and invasion of ovarian cancer cells through inhibition of apoptosis. *Eur Rev Med Pharmacol Sci* 23(12):5093–5100. https://doi.org/10.26355/eurerv_201906_1817
 217. Xu Y, Wang W, Chen J, Mao H, Liu Y, Gu S, Liu Q, Xi Q, Shi W (2020) High neuropilin and toll-like 1 expression associated with metastasis and poor survival in epithelial ovarian cancer via regulation of actin cytoskeleton. *J Cell Mol Med* 24(16):9114–9124. <https://doi.org/10.1111/jcmm.15547>
 218. Gu Y, Zhou G, Tang X, Shen F, Ding J, Hua K (2023) The biological roles of CD24 in ovarian cancer: old story, but new tales. *Front Immunol* 14:1183285. <https://doi.org/10.3389/fimmu.2023.1183285>
 219. Zhong Y, Wang Y, Huang J, Xu X, Pan W, Gao S, Zhang Y, Su M (2019) Association of hCG and LHCG expression patterns with clinicopathological parameters in ovarian cancer. *Pathol Res Pract* 215(4):748–754. <https://doi.org/10.1016/j.prp.2019.01.001>
 220. Carbone M, Melino G (2019) Stearoyl CoA desaturase regulates ferroptosis in ovarian cancer offering new therapeutic perspectives. *Cancer Res* 79(20):5149–5150. <https://doi.org/10.1158/0008-5472.CAN-19-2453>
 221. Szubert S, Moszynski R, Szpuredk D, Romaniuk B, Sajdak S, Nowicki M, Michalak S (2019) The expression of platelet-derived growth factor receptors (PDGFRs) and their correlation with overall survival of patients with ovarian cancer. *Ginekol Pol* 90(5):242–249. <https://doi.org/10.5603/GPa2019.0045>
 222. Saini A, Chandra KB, Kumar V, Mathur SR, Sharma JB, Kumar S, Yadav S (2020) Analysis of multimerin 1 (MMRN1) expression in ovarian cancer. *Mol Biol Rep* 47(12):9459–9468. <https://doi.org/10.1007/s11033-020-06027-9>
 223. Liu L, Sun YH, An R, Cheng RJ, Li N, Zheng JH (2023) LDLR promotes autophagy-mediated cisplatin resistance in ovarian cancer associated with the PI3K/AKT/mTOR signaling pathway. *Kaohsiung J Med Sci* 39(8):779–788. <https://doi.org/10.1002/kjm.212696>
 224. Arman Karakaya Y, Atıgan A, Güler ÖT, Demiray AG, Bir F (2021) The relation of CD3, CD4, CD8 and PD-1 expression with tumor type and prognosis in epithelial ovarian cancers. *Ginekol Pol* 92(5):344–351. <https://doi.org/10.5603/GPa2021.0080>
 225. Kommoss S, Anglesio MS, Mackenzie R, Yang W, Senz J, Ho J, Bell L, Lee S, Lorette J, Huntsman DG, Blake GC (2013) FOXL2 molecular testing in ovarian neoplasms: diagnostic approach and procedural guidelines. *Mod Pathol* 26(6):860–867. <https://doi.org/10.1038/modpathol.2012.226>
 226. Cao KY, Yan TM, Zhang JZ, Chan TF, Li J, Li C, Lai-Han Leung E, Gao J, Zhang BX, Jiang ZH (2022) A tRNA-derived fragment from Chinese yew suppresses ovarian cancer growth via targeting TRPA1. *Mol Ther Nucleic Acids* 27:718–732. <https://doi.org/10.1016/j.omtn.2021.12.037>
 227. Chen X, Wang X, Wei X, Wang J (2016) EphA5 protein, a potential marker for distinguishing histological grade and prognosis in ovarian serous carcinoma. *J Ovarian Res* 9(1):83. <https://doi.org/10.1186/s13048-016-0292-1>
 228. Li S, Yang S, Hong Y (2022) Higher thymocyte selection-associated high mobility group box (TOX) expression predicts poor prognosis in patients with ovarian cancer. *BMC Cancer* 22(1):1216. <https://doi.org/10.1186/s12885-022-10336-6>
 229. Wang S, Wang C, Liu O, Hu Y, Li X, Lin B (2021) Prognostic value of immune-related cells and genes in the tumor microenvironment of ovarian cancer, especially CST4. *Life Sci* 277:119461. <https://doi.org/10.1016/j.lfs.2021.119461>
 230. Gu H, Tu H, Liu L, Liu T, Liu Z, Zhang W, Liu J (2020) RSP03 is a marker candidate for predicting tumor aggressiveness in ovarian cancer. *Ann Transl Med* 8(21):1351. <https://doi.org/10.21037/atm-20-3731>
 231. Guo Y, Yuan J, Yin S, Wang X, Shuai R, Kang J (2017) MAP2K6-FP enhances the sensitiveness of paclitaxel for ovarian cancer via inducing autophagy. *Int J Gynecol Cancer* 27(6):1082–1087. <https://doi.org/10.1097/IGC.0000000000001003>
 232. Czekierdowski A, Stachowicz N, Czekierdowska S, Łoziński T, Guryńczak G, Kluz T (2018) Prognostic significance of TEM7 and nestin expression in women with advanced high grade serous ovarian cancer. *Ginekol Pol* 89(3):135–141. <https://doi.org/10.5603/GPa2018.0023>
 233. Sun T, Bi F, Liu Z, Yang Q (2021) TMEM119 facilitates ovarian cancer cell proliferation, invasion, and migration via the PDGFRB/PI3K/AKT signaling pathway. *J Transl Med* 19(1):111. <https://doi.org/10.1186/s12967-021-02781-x>
 234. Liu L, Zhang Z, Zhang G, Wang T, Ma Y, Guo W (2020) Down-regulation of PAD12 prevents proliferation and epithelial-mesenchymal transition in ovarian cancer through inhibiting JAK2/STAT3 pathway in vitro and in vivo, alone or in combination with Olaparib. *J Transl Med* 18(1):357. <https://doi.org/10.1186/s12967-020-02528-0>
 235. Arechavaleta-Velasco F, Cuevas-Antonio R, Dominguez-Lopez P, Estrada-Moscoco I, Imani-Razavi FS, Zeferino-Toquero M, Diaz-Cueto L (2014) Matrix metalloproteinase-8 promoter gene polymorphisms in Mexican women with ovarian cancer. *Med Oncol* 31(8):132. <https://doi.org/10.1007/s12032-014-0132-3>
 236. Cybulski M, Jeleniewicz W, Nowakowski A, Stenzel-Bembenek A, Tarkowski R, Kotarski J, Stepulak A (2015) Cyclin I mRNA expression correlates with kinase insert domain receptor expression in human epithelial ovarian cancer. *Anticancer Res* 35(2):1115–1119
 237. Huang T, Tworoger SS, Hecht JL, Rice MS, Sood AK, Kubzansky LD, Poole EM (2016) Association of ovarian tumor β 2-adrenergic receptor status with ovarian cancer risk factors and survival. *Cancer Epidemiol Biomarkers Prev* 25(12):1587–1594. <https://doi.org/10.1158/1055-9965.EPI-16-0534>
 238. Li J, Xu J, Li L, Ianni A, Kumari P, Liu S, Sun P, Braun T, Tan X, Xiang R et al (2020) MGAT3-mediated glycosylation of tetraspanin CD82 at asparagine 157 suppresses ovarian cancer metastasis by inhibiting the integrin signaling pathway. *Theranostics* 10(14):6467–6482. <https://doi.org/10.7150/thno.43865>
 239. Landskron J, Kraggerud SM, Wik E, Dørum A, Bjørnslett M, Melum E, Helland Ø, Bjørge L, Lothe RA, Salvesen HB et al (2017) C77G in PTPRC (CD45) is no risk allele for ovarian cancer, but associated with less aggressive disease. *PLoS ONE* 12(7):e0182030. <https://doi.org/10.1371/journal.pone.0182030>
 240. Li Y, Jaiswal SK, Kaur R, Alsaadi D, Liang X, Drews F, DeLoia JA, Krivak T, Pietrykowska HM, Gotea V et al (2021) Differential gene expression identifies a transcriptional regulatory network involving ER-alpha and PITX1 in invasive epithelial ovarian cancer. *BMC Cancer* 21(1):768. <https://doi.org/10.1186/s12885-021-08276-8>
 241. Al-Zahrani MH, Yahya FM, Assidi M, Dallol A, Buhmeida A (2021) Klotho promoter methylation status and its prognostic value in ovarian cancer. *Mol Clin Oncol* 15(3):181. <https://doi.org/10.3892/mco.2021.2343>
 242. van Dam PA, Coelho A, Rolfo C (2017) Is there a role for urokinase-type plasminogen activator inhibitors as maintenance therapy in patients with ovarian cancer? *Eur J Surg Oncol* 43(2):252–257. <https://doi.org/10.1016/j.ejso.2016.06.002>
 243. Couch FJ, Gaudet MM, Antoniou AC, Ramus SJ, Kuchenbaecker KB, Soucy P, Beesley J, Chen X, Wang X, Kirchoff T et al (2012) Common variants at the 19p13.1 and ZNF365 loci are associated with ER subtypes of breast cancer and ovarian cancer risk in BRCA1 and BRCA2 mutation carriers. *Cancer Epidemiol Biomarkers Prev* 21(4):645–657. <https://doi.org/10.1158/1055-9965.EPI-11-0888>
 244. Sohn EJ (2022) PIK3R3, a regulatory subunit of PI3K, modulates ovarian cancer stem cells and ovarian cancer development and progression by integrative analysis. *BMC Cancer* 22(1):708. <https://doi.org/10.1186/s12885-022-09807-7>
 245. Sun H, Wang H, Wang X, Aoki Y, Wang X, Yang Y, Cheng X, Wang Z, Wang X (2020) Aurora-A/SOX8/FOXK1 signaling axis promotes chemoresistance via suppression of cell senescence and induction of glucose metabolism in ovarian cancer organoids and cells. *Theranostics* 10(15):6928–6945. <https://doi.org/10.7150/thno.43811>
 246. Zhang H, Yuan N, Che H, Cheng X (2023) MiR-188-5p inhibits cell proliferation and migration in ovarian cancer via competing for CCND2 with ELAVL1. *Cell Mol Biol (Noisy-le-grand)* 69(3):69–74. <https://doi.org/10.14715/cmb/2023.69.3.9>
 247. Fu X, Zhang Q, Wang Z, Xu Y, Dong Q (2024) CRABP2 affects chemotherapy resistance of ovarian cancer by regulating the expression of HIF1 α . *Cell Death Dis* 15(1):21. <https://doi.org/10.1038/s41419-023-06398-4>
 248. Shi C, Yang Y, Zhang L, Yu J, Qin S, Xu H, Gao Y (2019) MiR-200a-3p promoted the malignant behaviors of ovarian cancer cells through

- regulating PCDH9. *Onco Targets Ther* 12:8329–8338. <https://doi.org/10.2147/OTT.S220339>
249. Nunna S, Reinhardt R, Ragozin S, Jeltsch A (2014) Targeted methylation of the epithelial cell adhesion molecule (EpCAM) promoter to silence its expression in ovarian cancer cells. *PLoS ONE* 9(1):e87703. <https://doi.org/10.1371/journal.pone.0087703>
 250. Krishna Priya S, Kumar K, Hiran KR, Bindhu MR, Nagare RP, Vijaykumar DK, Ganesan TS (2017) Expression of a novel endothelial marker, C-type lectin 14A, in epithelial ovarian cancer and its prognostic significance. *Int J Clin Oncol* 22(1):107–117. <https://doi.org/10.1007/s10147-016-1033-6>
 251. Piotrowska-Kempisty H, Klupczyńska A, Trzybulska D, Kulcenty K, Sulej-Suchomska AM, Kucińska M, Mikstacka R, Wierchowski M, Murias M, Baer-Dubowska W, Kokot Z et al (2017) Role of CYP1A1 in the biological activity of methylated resveratrol analogue, 3,4,5,4'-tetramethoxystilbene (DMU-212) in ovarian cancer A-2780 and non-cancerous HOSE cells. *Toxicol Lett* 267:59–66. <https://doi.org/10.1016/j.toxlet.2016.12.018>
 252. Papanastasiou AD, Pampalakis G, Katsaros D, Sotiropoulou G (2011) Netrin-1 overexpression is predictive of ovarian malignancies. *Oncotarget* 2(5):363–367. <https://doi.org/10.18632/oncotarget.258>
 253. Wang Y, Hu C, Dong R, Huang X, Qiu H (2011) Platelet-derived growth factor-D promotes ovarian cancer invasion by regulating matrix metalloproteinases 2 and 9. *Asian Pac J Cancer Prev* 12(12):3367–3370
 254. Uthayanan L, El-Bahrawy M (2022) Potential roles of claudin-3 and claudin-4 in ovarian cancer management. *J Egypt Natl Canc Inst* 34(1):24. <https://doi.org/10.1186/s43046-022-00125-4>
 255. Kumar J, Fang H, McCulloch DR, Crowley T, Ward AC (2017) Leptin receptor signaling via Janus kinase 2/Signal transducer and activator of transcription 3 impacts on ovarian cancer cell phenotypes. *Oncotarget* 8(55):93530–93540. <https://doi.org/10.18632/oncotarget.19873>
 256. Felices M, Wesley E, Bendzick LE, Kodal B, Hopps R, Grzywacz B, Hinderlie P, Miller JS, Geller MA (2023) Reverse translation identifies the synergistic role of immune checkpoint blockade and IL15 to enhance immunotherapy of ovarian cancer. *Cancer Immunol Res* 11(5):674–686. <https://doi.org/10.1158/2326-6066.CIR-22-0600>
 257. Le Page C, Puiffe ML, Meunier L, Zietarska M, de Ladurantaye M, Tonin PN, Provencher D, Mes-Masson AM (2009) BMP-2 signaling in ovarian cancer and its association with poor prognosis. *J Ovarian Res* 2:4. <https://doi.org/10.1186/1757-2215-2-4>
 258. Diao B, Sun C, Yu P, Zhao Z, Yang P (2023) LAMA5 promotes cell proliferation and migration in ovarian cancer by activating Notch signaling pathway. *FASEB J* 37(9):e23109. <https://doi.org/10.1096/fj.202300306R>
 259. Fang S, Luo Y, Zhang Y, Wang H, Liu Q, Li X, Yu T (2021) NTNG1 modulates cisplatin resistance in epithelial ovarian cancer cells via the GAS6/AXL/Akt pathway. *Front Cell Dev Biol* 9:652325. <https://doi.org/10.3389/fcell.2021.652325>
 260. Sun Z, Zhou R, Dai J, Chen J, Liu Y, Wang M, Zhou R, Liu F, Zhang Q, Xu Y et al (2023) KRT19 is a promising prognostic biomarker and associates with immune infiltrates in serous ovarian cystadenocarcinoma. *Int J Gen Med* 16:4849–4862. <https://doi.org/10.2147/IJGM.S419235>
 261. Fang DD, Tao R, Wang G, Li Y, Zhang K, Xu C, Zhai G, Wang Q, Wang J, Tang C et al (2022) Discovery of a novel ALK/ROS1/FAK inhibitor, APG-2449, in preclinical non-small cell lung cancer and ovarian cancer models. *BMC Cancer* 22(1):752. <https://doi.org/10.1186/s12885-022-09799-4>
 262. Lai H, Zhao X, Qin Y, Ding Y, Chen R, Li G, Labrie M, Ding Z, Zhou J, Hu J et al (2018) FAK-ERK activation in cell/matrix adhesion induced by the loss of apolipoprotein E stimulates the malignant progression of ovarian cancer. *J Exp Clin Cancer Res* 37(1):32. <https://doi.org/10.1186/s13046-018-0696-4>
 263. Karin-Kujundzic V, Covarrubias-Pinto A, Skrtic A, Vranic S, Serman L (2022) New insight into the role of PTCH1 protein in serous ovarian carcinomas. *Int J Oncol* 61(6):145. <https://doi.org/10.3892/ijo.2022.5435>
 264. Shaosheng W, Shaochuang W, Lichun F, Na X, Xiaohong Z (2021) ITPKA induces cell senescence, inhibits ovarian cancer tumorigenesis and can be downregulated by miR-203. *Aging (Albany NY)* 13(8):11822–11832. <https://doi.org/10.18632/aging.202880>
 265. Calbay O, Padia R, Akter M, Sun L, Li B, Qian N, Guo J, Fu Z, Jin L, Huang S (2023) ASC/inflammasome-independent pyroptosis in ovarian cancer cells through translational augmentation of caspase-1. *iScience* 26(12):108408. <https://doi.org/10.1016/j.isci.2023.108408>
 266. Zhou Y, Zhu Y, Fan X, Zhang C, Wang Y, Zhang L, Zhang H, Wen T, Zhang K, Huo X et al (2017) NID1, a new regulator of EMT required for metastasis and chemoresistance of ovarian cancer cells. *Oncotarget* 8(20):33110–33121. <https://doi.org/10.18632/oncotarget.16145>
 267. Li X, Zou Z, Tang J, Zheng Y, Liu Y, Luo Y, Liu Q, Wang Y (2019) NOS1 upregulates ABCG2 expression contributing to DDP chemoresistance in ovarian cancer cells. *Oncol Lett* 17(2):1595–1602. <https://doi.org/10.3892/ol.2018.9787>
 268. Beyazit F, Ayhan S, Celik HT, Gungor T (2015) Assessment of serum angiotensin-converting enzyme in patients with epithelial ovarian cancer. *Arch Gynecol Obstet* 292(2):415–420. <https://doi.org/10.1007/s00404-015-3661-x>
 269. Kanabekova P, Al-Awadi AM, Bauyrzhanova Z, Tahtouh T, Sarray S, Almawi WY (2022) Genetic variation in progesterone receptor gene and ovarian cancer risk: a case control study. *Gene* 820:146288. <https://doi.org/10.1016/j.gene.2022.146288>
 270. Stewart J, James J, McCluggage GW, McQuaid S, Arthur K, Boyle D, Mullan P, McArt D, Yan B, Irwin G et al (2015) Analysis of wntless (WLS) expression in gastric, ovarian, and breast cancers reveals a strong association with HER2 overexpression. *Mod Pathol* 28(3):428–436. <https://doi.org/10.1038/modpathol.2014.114>
 271. O'Mara TA, Nagle CM, Batra J, Kedda MA, Clements JA, Spurdle AB (2011) Kallikrein-related peptidase 3 (KLK3/PSA) single nucleotide polymorphisms and ovarian cancer survival. *Twin Res Hum Genet* 14(4):323–327. <https://doi.org/10.1375/twin.14.4.323>
 272. Kolb S, Hoffmann I, Monjé N, Dragomir MP, Jank P, Bischoff P, Keunecke C, Pohl J, Kunze CA, Marchenko S et al (2023) LRP1B-a prognostic marker in tubo-ovarian high-grade serous carcinoma. *Hum Pathol* 141:158–168. <https://doi.org/10.1016/j.humpath.2023.09.001>
 273. Geng L, Wang Z, Tian Y (2022) Down-regulation of ZNF252P-AS1 alleviates ovarian cancer progression by binding miR-324-3p to downregulate LY6K. *J Ovarian Res* 15(1):1. <https://doi.org/10.1186/s13048-021-00933-7>
 274. Ravenni N, Weber M, Neri D (2014) A human monoclonal antibody specific to placental alkaline phosphatase, a marker of ovarian cancer. *MABS* 6(1):86–94. <https://doi.org/10.4161/mabs.27230>
 275. van Amerongen RA, Tuit S, Wouters AK, van de Meent M, Siekman SL, Meeuwswen MH, Wachsmann TLA, Remst DFG, Hagedoorn RS, van der Steen DM et al (2023) PRAME and CTCFL-reactive TCRs for the treatment of ovarian cancer. *Front Immunol* 14:1121973. <https://doi.org/10.3389/fimmu.2023.1121973>
 276. Koller S, Kandler J, Karacs J, Wolf A, Kreuzinger C, Von Der Decken I, Mungenast F, Mechtcheriakova D, Schreiner W, Gleiss A et al (2022) SLC04A1 expression is associated with activated inflammatory pathways in high-grade serous ovarian cancer. *Front Pharmacol* 13:946348. <https://doi.org/10.3389/fphar.2022.946348>
 277. Tang H, Fayomi AP, Bai S, Gupta N, Cascio S, Yang D, Buckanovich RJ (2023) Generation and characterization of humanized affinity-matured EGFL6 antibodies for ovarian cancer therapy. *Gynecol Oncol* 171:49–58. <https://doi.org/10.1016/j.ygyno.2023.02.004>
 278. Li H, Zhao J, Shi X (2021) GPBAR1 promotes proliferation of serous ovarian cancer by inducing Smad4 ubiquitination. *Appl Immunohistochem Mol Morphol* 29(7):519–526. <https://doi.org/10.1097/PAI.00000000000000917>
 279. Wang J, Dai JM, Che YL, Gao YM, Peng HJ, Liu B, Wang H, Linghu H (2014) Elmo1 helps dock180 to regulate Rac1 activity and cell migration of ovarian cancer. *Int J Gynecol Cancer* 24(5):844–850. <https://doi.org/10.1097/IGC.0000000000000137>
 280. Li F, Liang Z, Jia Y, Zhang P, Ling K, Wang Y, Liang Z (2022) microRNA-324-3p suppresses the aggressive ovarian cancer by targeting WNK2/RAS pathway. *Bioengineered* 13(5):12030–12044. <https://doi.org/10.1080/21655979.2022.2056314>
 281. Meng X, Liang X, Yang S, Wu D, Wang X (2024) A miRNA-7704/IL2RB/AKT feedback loop regulates tumorigenesis and chemoresistance in ovarian cancer. *Exp Cell Res*. <https://doi.org/10.1016/j.yexcr.2024.114012>
 282. Sutton MN, Yang H, Huang GY, Fu C, Pontikos M, Wang Y, Mao W, Pang L, Yang M, Liu J et al (2018) RAS-related GTPases DIRAS1 and DIRAS2 induce autophagic cancer cell death and are required for autophagy in murine ovarian cancer cells. *Autophagy* 14(4):637–653. <https://doi.org/10.1080/15548627.2018.1427022>

283. Li HW, Liu MB, Jiang X, Song T, Feng SX, Wu JY, Deng PF, Wang XY (2022) GALNT14 regulates ferroptosis and apoptosis of ovarian cancer through the EGFR/mTOR pathway. *Future Oncol* 18(2):149–161. <https://doi.org/10.2217/fon-2021-0883>
284. Lin Z, Li D, Cheng W, Wu J, Wang K, Hu Y (2019) MicroRNA-181 functions as an antioncogene and mediates NF- κ B pathway by targeting RTKN2 in ovarian cancers. *Reprod Sci* 26(8):1071–1081. <https://doi.org/10.1177/1933719118805865>
285. Lyu M, Li X, Shen Y, Lu J, Zhang L, Zhong S, Wang J (2022) CircATRN1 and circZNF608 inhibit ovarian cancer by sequestering miR-152-5p and encoding protein. *Front Genet* 13:784089. <https://doi.org/10.3389/fgene.2022.784089>
286. Deo AN, Thorat R, Dhavde AC, De A, Rekhi B, Ray P (2022) IGF1R- α 6 integrin-S100A4 network governs the organ-specific metastasis of chemoresistant epithelial ovarian cancer cells. *Biochim Biophys Acta Mol Basis Dis* 1868(1):166282. <https://doi.org/10.1016/j.bbadis.2021.166282>
287. Link T, Kuhlmann JD, Kobelt D, Herrmann P, Vassileva YD, Kramer M, Frank K, Göckjenan M, Wimberger P, Stein U (2019) Clinical relevance of circulating MACC1 and S100A4 transcripts for ovarian cancer. *Mol Oncol* 13(5):1268–1279. <https://doi.org/10.1002/1878-0261.12484>
288. Cymbaluk-Ploska A, Chudecka-Glaz A, Pius-Sadowska E, Machaliński B, Menkiszak J, Sompolska-Rzechuła A (2018) Suitability assessment of baseline concentration of MMP3, TIMP3, HE4 and CA125 in the serum of patients with ovarian cancer. *J Ovarian Res* 11(1):1. <https://doi.org/10.1186/s13048-017-0373-9>
289. Wu YH, Huang YF, Chang TH, Chen CC, Wu PY, Huang SC, Chou CY (2021) COL11A1 activates cancer-associated fibroblasts by modulating TGF- β 3 through the NF- κ B/IGFBP2 axis in ovarian cancer cells. *Oncogene* 40(26):4503–4519. <https://doi.org/10.1038/s41388-021-01865-8>
290. Sakamoto H, Friel AM, Wood AW, Guo L, Illic A, Seiden MV, Chung DC, Lynch MP, Serikawa T, Munro E et al (2008) Mechanisms of Cables 1 gene inactivation in human ovarian cancer development. *Cancer Biol Ther* 7(2):180–188. <https://doi.org/10.4161/cbt.7.2.5253>
291. Li M, Qian Z, Ma X, Lin X, You Y, Li Y, Chen T, Jiang H (2018) MiR-628-5p decreases the tumorigenicity of epithelial ovarian cancer cells by targeting at FGFR2. *Biochem Biophys Res Commun* 495(2):2085–2091. <https://doi.org/10.1016/j.bbrc.2017.12.049>
292. Reivan Ortiz GG, Ciongradi CI, Chaitanya MVNL, Narayanan J, Mohany M, Al-Rejaie SS, Arias-González JL, Sárbu I, Assefi M, Akram SV et al (2023) Identification of novel candidate targets for suppressing ovarian cancer progression through IL-33/ST2 axis components using the system biology approach. *Front Mol Biosci* 10:1189527. <https://doi.org/10.3389/fmolb.2023.1189527>
293. Garrido MP, Vera C, Vega M, Quest AFG, Romero C (2018) Metformin prevents nerve growth factor-dependent proliferative and proangiogenic effects in epithelial ovarian cancer cells and endothelial cells. *Ther Adv Med Oncol* 10:1758835918770984. <https://doi.org/10.1177/1758835918770984>
294. Recouvreur MS, Miao J, Gozo MC, Wu J, Walts AE, Karlan BY, Orsulic S (2022) FOXC2 promotes vasculogenic mimicry in ovarian cancer. *Cancers (Basel)* 14(19):4851. <https://doi.org/10.3390/cancers14194851>
295. Gorji-Bahri G, Krishna BM, Hagerling C, Orimo A, Jirström K, Papadakos KS, Blom AM (2024) Stromal cartilage oligomeric matrix protein as a tumorigenic driver in ovarian cancer via Notch3 signaling and epithelial-to-mesenchymal transition. *J Transl Med* 22(1):351. <https://doi.org/10.1186/s12967-024-05083-0>
296. Cole AJ, Panesso-Gómez S, Shah JS, Ebai T, Jiang Q, Gumusoglu-Acar E, Bello MG, Vlad A, Modugno F, Edwards RP et al (2023) Quiescent ovarian cancer cells secrete follistatin to induce chemotherapy resistance in surrounding cells in response to chemotherapy. *Clin Cancer Res* 29(10):1969–1983. <https://doi.org/10.1158/1078-0432.CCR-22-2254>
297. Deng Y, Dong Y, Wu L, Zhang Q, Yang L (2023) ARID5B promoted the histone demethylation of SORBS2 and hampered the metastasis of ovarian cancer. *Pathol Res Pract* 252:154911. <https://doi.org/10.1016/j.prp.2023.154911>
298. Yin X, Fang S, Wang M, Wang Q, Fang R, Chen J (2016) EFEMP1 promotes ovarian cancer cell growth, invasion and metastasis via activated the AKT pathway. *Oncotarget* 7(30):47938–47953. <https://doi.org/10.18632/oncotarget.10296>
299. Gao F, Wu Q, Lu D (2024) MicroRNA-10a-5p-mediated downregulation of GATA6 inhibits tumor progression in ovarian cancer. *Hum Cell* 37(1):271–284. <https://doi.org/10.1007/s13577-023-00987-3>
300. Hussain A, Voisin V, Poon S, Karamboulas C, Bui NHB, Meens J, Dmytryshyn J, Ho VW, Tang KH, Paterson J et al (2020) Distinct fibroblast functional states drive clinical outcomes in ovarian cancer and are regulated by TCF21. *J Exp Med* 217(8):e20191094. <https://doi.org/10.1084/jem.20191094>
301. Lurie G, Terry KL, Wilkens LR, Thompson PJ, McDuffie KE, Carney ME, Palmieri RT, Cramer DW, Goodman MT (2010) Pooled analysis of the association of PTGS2 rs5275 polymorphism and NSAID use with invasive ovarian carcinoma risk. *Cancer Causes Control* 21(10):1731–1741. <https://doi.org/10.1007/s10552-010-9602-x>
302. Wu M, Qiu Q, Zhou Q, Li J, Yang J, Zheng C, Luo A, Li X, Zhang H, Cheng X et al (2022) circFBXO7/miR-96-5p/MTSS1 axis is an important regulator in the Wnt signaling pathway in ovarian cancer. *Mol Cancer* 21(1):137. <https://doi.org/10.1186/s12943-022-01611-y>
303. Homayoun M, Sajedi N, Soleimani M (2022) In vitro evaluation of the pogostone effects on the expression of PTEN and DACT1 tumor suppressor genes, cell cycle, and apoptosis in ovarian cancer cell line. *Res Pharm Sci* 17(2):164–175. <https://doi.org/10.4103/1735-5362.335175>
304. Li M, Cai J, Han X, Ren Y (2020) Downregulation of circNRP1 suppresses the paclitaxel resistance of ovarian cancer via regulating the miR-211-5p/HOXC8 axis. *Cancer Manag Res* 12:9159–9171. <https://doi.org/10.2147/CMAR.S268872>
305. Basu M, Roy SS (2013) Wnt/ β -catenin pathway is regulated by PITX2 homeodomain protein and thus contributes to the proliferation of human ovarian adenocarcinoma cell. *SKOV-3. J Biol Chem* 288(6):4355–4367. <https://doi.org/10.1074/jbc.M112.409102>
306. Charbonneau B, Block MS, Bamlet WR, Vierkant RA, Kalli KR, Fogarty Z, Rider DN, Sellers TA, Tworoger SS, Poole E et al (2014) Risk of ovarian cancer and the NF- κ B pathway: genetic association with IL1A and TNFSF10. *Cancer Res* 74(3):852–861. <https://doi.org/10.1158/0008-5472.CAN-13-1051>
307. Cartmel B, Hughes M, Ercolano EA, Gottlieb L, Li F, Zhou Y, Harrigan M, Ligibel JA, von Gruenigen VE, Gogoi R et al (2021) Randomized trial of exercise on depressive symptomatology and brain derived neurotrophic factor (BDNF) in ovarian cancer survivors: the women's activity and lifestyle study in connecticut (WALC). *Gynecol Oncol* 161(2):587–594. <https://doi.org/10.1016/j.ygyno.2021.02.036>
308. Lin M, Wang J, Wang S, Huang Y (2022) Expression level of keratin 7 in epithelial ovarian cancer and malignant metastasis of benign epithelial ovarian tumors. *Cell Mol Biol (Noisy-le-grand)* 68(2):153–161. <https://doi.org/10.14715/cmb/2022.68.2.22>
309. Cui Y, Shen G, Ma L, Lv Q (2021) Overexpression of NDRG2 promotes the therapeutic effect of pazopanib on ovarian cancer. *J Recept Signal Transduct Res* 41(6):546–552. <https://doi.org/10.1080/10799893.2020.1831536>
310. Dawoud MM, Aiad HAES, Tawfiq EAE, Al-Qalashy FSA, Eissa N, El-Rebey HS (2021) Role of SIX1, EYA2, and E-cadherin in ovarian carcinoma. Evidence on epithelial-mesenchymal transition from an immunohistochemical study. *Ann Diagn Pathol* 55:151815. <https://doi.org/10.1016/j.anndiagpath.2021.151815>
311. Li X, Yang Z, Xu S, Wang Z, Jin P, Yang X, Zhang Z, Wang Y, Wei X, Fang T et al (2019) Targeting INHBA in ovarian cancer cells suppresses cancer xenograft growth by attenuating stromal fibroblast activation. *Dis Markers* 2019:7275289. <https://doi.org/10.1155/2019/7275289>
312. D'Antona L, Dattilo V, Catalogna G, Scumaci D, Fiumara CV, Musumeci F, Perrotti G, Schenone S, Talerico R, Spoletti CB et al (2019) Preclinical model of ovarian cancer, the SGK1 inhibitor SI113 counteracts the development of paclitaxel resistance and restores drug sensitivity. *Transl Oncol* 12(8):1045–1055. <https://doi.org/10.1016/j.tranon.2019.05.008>
313. Li M, Li L, Cheng X, Li L, Tu K (2023) Hypoxia promotes the growth and metastasis of ovarian cancer cells by suppressing ferroptosis via upregulating SLC2A12. *Exp Cell Res* 433(2):113851. <https://doi.org/10.1016/j.yexcr.2023.113851>
314. Moskovich D, Finkelshtein Y, Alfandari A, Rosemarin A, Lifschytz T, Weisz A, Mondal S, Ungati H, Katzav A, Kidron D et al (2021) Targeting the DIO3 enzyme using first-in-class inhibitors effectively suppresses

- tumor growth: a new paradigm in ovarian cancer treatment. *Oncogene* 40(44):6248–6257. <https://doi.org/10.1038/s41388-021-02020-z>
315. Dafou D, Grun B, Sinclair J, Lawrenson K, Benjamin EC, Hogdall E, Kruger-Kjaer S, Christensen L, Sowter HM, Al-Attar A et al (2010) Microcell-mediated chromosome transfer identifies EPB41L3 as a functional suppressor of epithelial ovarian cancers. *Neoplasia* 12(7):579–589. <https://doi.org/10.1593/neo.10340>
 316. An Y, Wang X, Chen L, Yang Y, Lin X, Wang N, Duan J, Long S, Zhao X (2021) Oncolytic reovirus induces ovarian cancer cell apoptosis in a TLR3-dependent manner. *Virus Res* 301:198440. <https://doi.org/10.1016/j.virusres.2021.198440>
 317. Xu J, Wu F, Zhu Y, Wu T, Cao T, Gao W, Liu M, Qian W, Feng G, Xi X et al (2024) ANGPTL4 regulates ovarian cancer progression by activating the ERK1/2 pathway. *Cancer Cell Int* 24(1):54. <https://doi.org/10.1186/s12935-024-03246-z>
 318. Wu Q, Suo Z, Kristensen GB, Baekelandt M, Nesland JM (2006) The prognostic impact of EphB2/B4 expression on patients with advanced ovarian carcinoma. *Gynecol Oncol* 102(1):15–21. <https://doi.org/10.1016/j.ygyno.2005.11.034>
 319. Song W, Hu L, Li W, Wang G, Li Y, Yan L, Li A, Cui J (2014) Oncogenic Fli-1 is a potential prognostic marker for the progression of epithelial ovarian cancer. *BMC Cancer* 14:424. <https://doi.org/10.1186/1471-2407-14-424>
 320. Zeng Z, Lin C, Zhang MC, Kossinna P, Wang P, Cao D, Wang J, Xu M, Wang X, Li Q et al (2023) Enterolactone and trabectedin suppress epithelial ovarian cancer synergistically via upregulating THBS1. *Phytother Res* 37(10):4722–4739. <https://doi.org/10.1002/ptr.7942>
 321. Shepherd TG, Thériault BL, Nachtigal MW (2008) Autocrine BMP4 signaling regulates ID3 proto-oncogene expression in human ovarian cancer cells. *Gene* 414(1–2):95–105. <https://doi.org/10.1016/j.gene.2008.02.015>
 322. Liu WJ, Huang YX, Wang W, Zhang Y, Liu BJ, Qiu JG, Jiang BH, Liu LZ (2021) NOX4 signaling mediates cancer development and therapeutic resistance through HER3 in ovarian cancer cells. *Cells* 10(7):1647. <https://doi.org/10.3390/cells10071647>
 323. Lecker LSM, Berlato C, Maniati E, Delaine-Smith R, Pearce OMT, Heath O, Nichols SJ, Trevisan C, Novak M, McDermott J et al (2021) TGF β 1 production by macrophages contributes to an immunosuppressive microenvironment in ovarian cancer. *Cancer Res* 81(22):5706–5719. <https://doi.org/10.1158/0008-5472.CAN-21-0536>
 324. Zhu Y, Yang L, Wang J, Li Y, Chen Y (2022) SP1-induced lncRNA MCF2L-AS1 promotes cisplatin resistance in ovarian cancer by regulating IGF2BP1/IGF2/MEK/ERK axis. *J Gynecol Oncol* 33(6):e75. <https://doi.org/10.3802/jgo.2022.33.e75>
 325. Yang M, Xie X, Ding Y (2016) SALL4 is a marker of poor prognosis in serous ovarian carcinoma promoting invasion and metastasis. *Oncol Rep* 35(3):1796–1806. <https://doi.org/10.3892/or.2016.4545>
 326. Tan Y, Cheung M, Pei J, Menges CW, Godwin AK, Testa JR (2010) Upregulation of DLX5 promotes ovarian cancer cell proliferation by enhancing IRS-2-AKT signaling. *Cancer Res* 70(22):9197–9206. <https://doi.org/10.1158/0008-5472.CAN-10-1568>
 327. Huang YF, Wu YH, Cheng WF, Peng SL, Shen WL, Chou CY (2018) Vitamin D-binding protein enhances epithelial ovarian cancer progression by regulating the insulin-like growth factor-1/Akt pathway and vitamin D receptor transcription. *Clin Cancer Res* 24(13):3217–3228. <https://doi.org/10.1158/1078-0432.CCR-17-2943>
 328. Califano D, Pignata S, Pisano C, Gregg S, Laurelli G, Losito NS, Ottaiano A, Gallipoli A, Pasquinelli R, De Simone V et al (2010) FEZ1/LZTS1 protein expression in ovarian cancer. *J Cell Physiol* 222(2):382–386. <https://doi.org/10.1002/jcp.21962>
 329. Asad M, Wong MK, Tan TZ, Choolani M, Low J, Mori S, Virshup D, Thiery JP, Huang RY (2014) FZD7 drives in vitro aggressiveness in Stem-A subtype of ovarian cancer via regulation of non-canonical Wnt/PCP pathway. *Cell Death Dis* 5(7):e1346. <https://doi.org/10.1038/cddis.2014.302>
 330. McGrath SE, Annels N, Madhuri TK, Taylor A, Butler-Manuel SA, Morgan R, Pandha H, Michael A (2018) Engrailed-2 (EN2) - a novel biomarker in epithelial ovarian cancer. *BMC Cancer* 18(1):943. <https://doi.org/10.1186/s12885-018-4816-5>
 331. Fan S, Wang Y, Sheng N, Xie Y, Lu J, Zhang Z, Shan Q, Wu D, Sun C, Li M et al (2019) Low expression of EN1 predicts a favorable prognosis in patients with ovarian cancer. *J Cell Biochem* 120(1):861–871. <https://doi.org/10.1002/jcb.27447>
 332. Marks ZRC, Campbell NK, Mangan NE, Vandenberg CJ, Gearing LJ, Matthews AY, Gould JA, Tate MD, Wray-McCann G, Ying L et al (2023) Interferon- ϵ is a tumour suppressor and restricts ovarian cancer. *Nature* 620(7976):1063–1070. <https://doi.org/10.1038/s41586-023-06421-w>
 333. Li Y, Qu J, Sun Y, Chang C (2023) Troponin T1 promotes the proliferation of ovarian cancer by regulating cell cycle and apoptosis. *Iran J Biotechnol* 21(1):e3405. <https://doi.org/10.30498/ijb.2022.344921.3405>
 334. Lei Y, Henderson BR, Emmanuel C, Harnett PR, deFazio A (2015) Inhibition of ANKRD1 sensitizes human ovarian cancer cells to endoplasmic reticulum stress-induced apoptosis. *Oncogene* 34(4):485–495. <https://doi.org/10.1038/ncr.2013.566>
 335. Lu R, Tang P, Zhang D, Lin S, Li H, Feng X, Sun M, Zhang H (2023) SOX9/NFIA promotes human ovarian cancer metastasis through the Wnt/ β -catenin signaling pathway. *Pathol Res Pract* 248:154602. <https://doi.org/10.1016/j.prp.2023.154602>
 336. Nieddu V, Melocchi V, Battistini C, Franciosa G, Lupia M, Stellato C, Bertalot G, Olsen JV, Colombo N, Bianchi F et al (2023) Matrix Gla Protein drives stemness and tumor initiation in ovarian cancer. *Cell Death Dis* 14(3):220. <https://doi.org/10.1038/s41419-023-05760-w>
 337. Yi X, Liu Y, Zhou B, Xiang W, Deng A, Fu Y, Zhao Y, Ouyang Q, Liu Y, Sun Z et al (2021) Incorporating SULF1 polymorphisms in a pretreatment CT-based radiomic model for predicting platinum resistance in ovarian cancer treatment. *Biomed Pharmacother* 133:111013. <https://doi.org/10.1016/j.biopha.2020.111013>
 338. Paucarmayta A, Taitz H, Casablanca Y, Rodriguez GC, Maxwell GL, Darcy KM, Syed V (2019) TGF- β signaling proteins and CYP24A1 may serve as surrogate markers for progesterone calcium treatment in ovarian and endometrial cancers of different histological types. *Transl Cancer Res* 8(4):1423–1437. <https://doi.org/10.21037/tcr.2019.07.36>
 339. Verma S, Bakshi D, Sharma V, Sharma I, Shah R, Bhat A, Bhat GR, Sharma B, Wakhloo A, Kaul S et al (2020) Genetic variants of DNAH11 and LRFN2 genes and their association with ovarian and breast cancer. *Int J Gynaecol Obstet* 148(1):118–122. <https://doi.org/10.1002/ijgo.12997>
 340. Sobstyl M, Niedźwiedzka-Rystwej P, Hryniewicz R, Bębnowska D, Korona-Główniak I, Pasiarski M, Sosnowska-Pasiarska B, Smok-Kalwat J, Gózdź S, Sobstyl A et al (2021) Toll-like receptor 2 as a marker molecule of advanced ovarian cancer. *Biomolecules* 11(8):1205. <https://doi.org/10.3390/biom11081205>
 341. Yousefi H, Momeny M, Ghaffari SH, Parsanejad N, Poursheikhani A, Javadkooshesh S, Zarrinrad G, Esmaili F, Alishahi Z, Sabourinejad Z et al (2019) IL-6/IL-6R pathway is a therapeutic target in chemoresistant ovarian cancer. *Tumori* 105(1):84–91. <https://doi.org/10.1177/0300891618784790>
 342. Lawrenson K, Grun B, Lee N, Mhawech-Fauceglia P, Kan J, Swenson S, Lin YG, Pejovic T, Millstein J, Gayther SA (2015) NPPB is a novel candidate biomarker expressed by cancer-associated fibroblasts in epithelial ovarian cancer. *Int J Cancer* 136(6):1390–1401. <https://doi.org/10.1002/ijc.2909>
 343. Mehner C, Oberg AL, Kalli KR, Nassar A, Hockla A, Pendlebury D, Cichon MA, Goergen KM, Maurer MJ, Goode EL et al (2015) Serine protease inhibitor Kazal type 1 (SPINK1) drives proliferation and anoikis resistance in a subset of ovarian cancers. *Oncotarget* 6(34):35737–35754. <https://doi.org/10.18632/oncotarget.5927>
 344. Wiedemeyer K, Köbel M, Koelkebeck H, Xiao Z, Vashisht K (2020) High glypican-3 expression characterizes a distinct subset of ovarian clear cell carcinomas in Canadian patients: an opportunity for targeted therapy. *Hum Pathol* 98:56–63. <https://doi.org/10.1016/j.humpath.2020.01.002>
 345. Beck OG, Hardesty MM (2023) Entrectinib use in a platinum-refractory mucinous ovarian cancer harboring a NTRK3 gene fusion. *Gynecol Oncol Rep* 47:101187. <https://doi.org/10.1016/j.gore.2023.101187>
 346. Liu Y, Yang J, Shi Z, Tan X, Jin N, O'Brien C, Ott C, Grisoli A, Lee E, Volk K et al (2021) In vivo selection of highly metastatic human ovarian cancer sublines reveals role for AMIGO2 in intra-peritoneal metastatic regulation. *Cancer Lett* 503:163–173. <https://doi.org/10.1016/j.canlet.2021.01.024>
 347. Wang Y, Qiu C, Lu N, Liu Z, Jin C, Sun C, Bu H, Yu H, Dongol S, Kong B (2018) FOXD1 is targeted by miR-30a-5p and miR-200a-5p and suppresses the proliferation of human ovarian carcinoma cells by promoting p21 expression in a p53-independent manner. *Int J Oncol* 52(6):2130–2142. <https://doi.org/10.3892/ijco.2018.4359>

348. Vlad C, Kubelac P, Onisim A, Fetica B, Fulop A, Irimie A, Achimas-Cadariu P (2016) Expression of CDCP1 and ADAM12 in the ovarian cancer microenvironment. *J BUON* 21(4):973–978
349. Liu W, Tian X, Ding X, Zhang L (2019) Expression of dual-specificity phosphatase 2 (DUSP2) in patients with serous ovarian carcinoma and in SKOV3 and OVCAR3 cells in vitro. *Med Sci Monit* 25:10180–10189. <https://doi.org/10.12659/MSM.919089>
350. Guo B, Yu L, Sun Y, Yao N, Ma L (2020) Long non-coding RNA USP2-AS1 accelerates cell proliferation and migration in ovarian cancer by sponging miR-520d-3p and up-regulating KIAA1522. *Cancer Manag Res* 12:10541–10550. <https://doi.org/10.2147/CMAR.S268863>
351. Wang L, Ye TY, Wu H, Chen SY, Weng JR, Xi XW (2019) LINC00702 accelerates the progression of ovarian cancer through interacting with EZH2 to inhibit the transcription of KLF2. *Eur Rev Med Pharmacol Sci* 23(3 Suppl):201–208. https://doi.org/10.26355/eurrev_201908_18648
352. Jin Y, Wang H (2022) Circ_0078607 inhibits the progression of ovarian cancer via regulating the miR-32-5p/SIK1 network. *J Ovarian Res* 15(1):3. <https://doi.org/10.1186/s13048-021-00931-9>
353. Feng S, Ding B, Dai Z, Yin H, Ding Y, Liu S, Zhang K, Lin H, Xiao Z, Shen Y (2024) Cancer-associated fibroblast-secreted FGF7 as an ovarian cancer progression promoter. *J Transl Med* 22(1):280. <https://doi.org/10.1186/s12967-024-05085-y>
354. Liu L, Chen C, Liu P, Li J, Pang Z, Zhu J, Lin Z, Zhou H, Xie Y, Lan T et al (2023) MYH10 combines with MYH9 to recruit USP45 by deubiquitinating snail and promotes serous ovarian cancer carcinogenesis, progression, and cisplatin resistance. *Adv Sci (Weinh)* 10(14):e2203423. <https://doi.org/10.1002/advs.202203423>
355. Wang Q, Bian CE, Peng H, He L, Zhao X (2015) Association of circulating insulin-like growth factor 1 and insulin-like growth factor binding protein 3 with the risk of ovarian cancer: a systematic review and meta-analysis. *Mol Clin Oncol* 3(3):623–628. <https://doi.org/10.3892/mco.2015.516>
356. Zhang N, Kim SH, Gainullina A, Erlich EC, Onufer EJ, Kim J, Czepielewski RS, Helmink BA, Dominguez JR, Saunders BT et al (2021) LYVE1+ macrophages of murine peritoneal mesothelium promote omentum-independent ovarian tumor growth. *J Exp Med* 218(12):e20210924. <https://doi.org/10.1084/jem.20210924>
357. Topalov NE, Mayr D, Kuhn C, Leutbecher A, Scherer C, Kraus FBT, Tauber CV, Beyer S, Meister S, Hester A et al (2023) Characterization and prognostic impact of ACTBL2-positive tumor-infiltrating leukocytes in epithelial ovarian cancer. *Sci Rep* 13(1):22620. <https://doi.org/10.1038/s41598-023-49286-9>
358. Lin CJ, Huang WR, Wu CZ, Tseng RC (2021) Changes in SLIT2 expression are associated with the migration of human ovarian clear cell carcinoma cells. *Oncol Lett* 22(1):551. <https://doi.org/10.3892/ol.2021.12812>
359. Liu L, Lv J, Lin Z, Ning Y, Li J, Liu P, Chen C (2022) Co-overexpression of GRK5/ACTC1 correlates with the clinical parameters and poor prognosis of epithelial ovarian cancer. *Front Mol Biosci* 8:785922. <https://doi.org/10.3389/fmolb.2021.785922>
360. Harmankaya İ, Akar S, Uğraş S, Güler AH, Ezveci H, Aydoğdu M, Çelik Ç (2021) Nicotinamide N-methyltransferase overexpression may be associated with poor prognosis in ovarian cancer. *J Obstet Gynaecol* 41(2):248–253. <https://doi.org/10.1080/01443615.2020.1732891>
361. Lin HW, Chiang YC, Sun NY, Chen YL, Chang CF, Tai YJ, Chen CA, Cheng WF (2019) CHI3L1 results in poor outcome of ovarian cancer by promoting properties of stem-like cells. *Endocr Relat Cancer* 26(1):73–88. <https://doi.org/10.1530/ERC-18-0300>
362. Chen Y, He Y, Liu S (2023) RUNX1-regulated signaling pathways in ovarian cancer. *Biomedicines* 11(9):2357. <https://doi.org/10.3390/biomedicines11092357>
363. Gao Y, Huang Y (2023) Circ_0007841 knockdown confers cisplatin sensitivity to ovarian cancer cells by down-regulation of NFIB expression in a miR-532-5p-dependent manner. *J Chemother* 35(2):117–130. <https://doi.org/10.1080/1120009X.2022.2056995>
364. Xu X, Wang Q, Shen L, Shen Y, Liu H, Liu Y, Yang Z, Hoffman RM, Feng W (2024) Anlotinib inhibits ovarian cancer and enhances cisplatin sensitivity via suppressing NOTCH2 expression and stemness. *Anticancer Res* 44(4):1399–1407. <https://doi.org/10.21873/anticancerres.16936>
365. Meng Q, Duan P, Li L, Miao Y (2018) Expression of placenta growth factor is associated with unfavorable prognosis of advanced-stage serous ovarian cancer. *Tohoku J Exp Med* 244(4):291–296. <https://doi.org/10.1620/tjem.244.291>
366. Parra-Herran C, Dundr P, McCluggage WG (2024) Editorial: infiltrative pattern of invasion is independently associated with shorter survival and desmoplastic stroma markers FAP and THBS2 in mucinous ovarian carcinoma. *Histopathology* 84(7):1092–1094. <https://doi.org/10.1111/his.15181>
367. Dansonka-Mieszkowska A, Szafron LA, Kulesza M, Stachurska A, Leszczynski P, Tomczyk-Szatkowska A, Sobiczewski P, Parada J, Kulinczak M, Moes-Sosnowska J, Pienkowska-Grela B et al (2022) PROM1, CXCL8, RUNX1, NAV1 and TP73 genes as independent markers predictive of prognosis or response to treatment in two cohorts of high-grade serous ovarian cancer patients. *PLoS ONE* 17(7):e0271539. <https://doi.org/10.1371/journal.pone.0271539>
368. Sheng Q, Liu X, Fleming E, Yuan K, Piao H, Chen J, Moustafa Z, Thomas RK, Greulich H, Schinzel A et al (2010) An activated ErbB3/NRG1 autocrine loop supports in vivo proliferation in ovarian cancer cells. *Cancer Cell* 17(3):298–310. <https://doi.org/10.1016/j.ccr.2009.12.047>
369. Syed N, Coley HM, Sehouli J, Koensgen D, Mustea A, Szlosarek P, McNeish I, Blagden SP, Schmid P, Lovell DP et al (2011) Polo-like kinase Plk2 is an epigenetic determinant of chemosensitivity and clinical outcomes in ovarian cancer. *Cancer Res* 71(9):3317–3327. <https://doi.org/10.1158/0008-5472.CAN-10-2048>
370. Cortez AJ, Kujawa KA, Wilk AM, Sojka DR, Syrakis JP, Olbryt M, Lisowska KM (2020) Evaluation of the role of ITGBL1 in ovarian cancer. *Cancers (Basel)* 12(9):2676. <https://doi.org/10.3390/cancers12092676>
371. Siva A, Xin H, Qin F, Oltean D, Bowdish KS, Kretz-Rommel A (2008) Immune modulation by melanoma and ovarian tumor cells through expression of the immunosuppressive molecule CD200. *Cancer Immunol Immunother* 57(7):987–996. <https://doi.org/10.1007/s00262-007-0429-6>
372. Yang LQ, Hu HY, Han Y, Tang ZY, Gao J, Zhou QY, Liu YX, Chen HS, Xu TN, Ao L et al (2022) CpG-binding protein CFP1 promotes ovarian cancer cell proliferation by regulating BST2 transcription. *Cancer Gene Ther* 29(12):1895–1907. <https://doi.org/10.1038/s41417-022-00503-z>
373. Liu X, Wei L, Zhao B, Cai X, Dong C, Yin F (2018) Low expression of KCNN3 may affect drug resistance in ovarian cancer. *Mol Med Rep* 18(2):1377–1386. <https://doi.org/10.3892/mmr.2018.9107>
374. Liu CL, Pan HW, Torng PL, Fan MH, Mao TL (2019) SRPX and HMCN1 regulate cancer-associated fibroblasts to promote the invasiveness of ovarian carcinoma. *Oncol Rep* 42(6):2706–2715. <https://doi.org/10.3892/or.2019.7379>
375. Kollara A, Burt BD, Ringuette MJ, Brown TJ (2023) The adaptor protein VEPH1 interacts with the kinase domain of ERBB2 and impacts EGF signaling in ovarian cancer cells. *Cell Signal* 106:110634. <https://doi.org/10.1016/j.cellsig.2023.110634>
376. Arakawa N, Kobayashi H, Yonemoto N, Masuishi Y, Ino Y, Shigetomi H, Furukawa N, Ohtake N, Miyagi Y, Hirahara F et al (2016) Clinical significance of tissue factor pathway inhibitor 2, a serum biomarker candidate for ovarian clear cell carcinoma. *PLoS ONE* 11(10):e0165609. <https://doi.org/10.1371/journal.pone.0165609>
377. Sung HY, Han J, Ju W, Ahn JH (2016) Synaptotagmin-like protein 2 gene promotes the metastatic potential in ovarian cancer. *Oncol Rep* 36(1):535–541. <https://doi.org/10.3892/or.2016.4835>
378. Liang ZQ, Gao L, Chen JH, Dai WB, Su YS, Chen G (2021) Downregulation of the coiled-coil domain containing 80 and its perspective mechanisms in ovarian carcinoma: a comprehensive study. *Int J Genom* 2021:3752871. <https://doi.org/10.1155/2021/3752871>
379. Zuberi M, Dholariya S, Khan I, Mir R, Guru S, Bhat M, Sumi M, Saxena A (2021) Epigenetic silencing of DAPK1 and p16INK4a genes by CpG island hypermethylation in epithelial ovarian cancer patients. *Indian J Clin Biochem* 36(2):200–207. <https://doi.org/10.1007/s12291-020-00888-4>
380. Singh N, Hutson R, Milton NGN, Javid FA (2022) Ovarian cancer and KISS-1 gene expression: a consideration of the use of Kisspeptin plus Kisspeptin aptamers in diagnostics and therapy. *Eur J Pharmacol* 917:174752. <https://doi.org/10.1016/j.ejphar.2022.174752>
381. Li J, Qin X, Shi J, Wang X, Li T, Xu M, Chen X, Zhao Y, Han J, Piao Y et al (2021) A systematic CRISPR screen reveals an IL-20/IL20RA-mediated immune crosstalk to prevent the ovarian cancer metastasis. *Elife* 10:e66222. <https://doi.org/10.7554/eLife.66222>

382. Nykopp TK, Rilla K, Sironen R, Tammi MI, Tammi RH, Hämäläinen K, Heikkinen AM, Komulainen M, Kosma VM, Anttila M (2009) Expression of hyaluronan synthases (HAS1-3) and hyaluronidases (HYAL1-2) in serous ovarian carcinomas: inverse correlation between HYAL1 and hyaluronan content. *BMC Cancer* 9:143. <https://doi.org/10.1186/1471-2407-9-143>
383. Hetland TE, Nymoan DA, Emilsen E, Kærn J, Tropé CG, Flørenes VA, Davidson B (2012) MGMT1 expression in serous ovarian carcinoma differs at various anatomic sites, but is unrelated to chemoresistance or survival. *Gynecol Oncol* 126(3):460–465. <https://doi.org/10.1016/j.ygyno.2012.05.029>
384. Minopoli M, Botti G, Gigantino V, Ragone C, Sarno S, Motti ML, Scognamiglio G, Greggi S, Scaffa C, Roca MS et al (2019) Targeting the formyl peptide receptor type 1 to prevent the adhesion of ovarian cancer cells onto mesothelium and subsequent invasion. *J Exp Clin Cancer Res* 38(1):459. <https://doi.org/10.1186/s13046-019-1465-8>
385. Gong TT, Liu FH, Xiao Q, Li YZ, Wei YF, Xu HL, Cao F, Sun ML, Jiang FL, Tao T et al (2024) SH3RF2 contributes to cisplatin resistance in ovarian cancer cells by promoting RBPMS degradation. *Commun Biol* 7(1):67. <https://doi.org/10.1038/s42003-023-05721-1>
386. Giannini A, D'Orta O, Corrado G, Bruno V, Sperduti I, Bogani G, Laganà AS, Chiantera V, Caserta D, Vizza E (2024) The role of L1CAM as predictor of poor prognosis in stage I endometrial cancer: a systematic review and meta-analysis. *Arch Gynecol Obstet* 309(3):789–799. <https://doi.org/10.1007/s00404-023-07149-8>
387. Karageorgi S, McGrath M, Lee IM, Buring J, Kraft P, De Vivo I (2011) Polymorphisms in genes hydroxysteroid-dehydrogenase-17b type 2 and type 4 and endometrial cancer risk. *Gynecol Oncol* 121(1):54–58. <https://doi.org/10.1016/j.ygyno.2010.11.014>
388. Kiseli M, Caglar GS, Yarci Gursoy A, Tasci T, Candar T, Akincioglu E, Pabuccu EG, Boran N, Tulunay G, Umudum H (2018) Pro-gastrin releasing peptide: a new serum marker for endometrioid adenocarcinoma. *Gynecol Obstet Invest* 83(6):540–545. <https://doi.org/10.1159/000488854>
389. Wu Z, Jeong JH, Ren C, Yang L, Ding L, Li F, Jiang D, Zhu Y, Lu J (2021) Fatty acid-binding protein 4 (FABP4) suppresses proliferation and migration of endometrial cancer cells via PI3K/Akt pathway. *Onco Targets Ther* 14:3929–3942. <https://doi.org/10.2147/OTT.S311792>
390. Lin M, Lei T, Zheng J, Chen S, Du L, Xie H (2019) UBE2S mediates tumor progression via SOX6/ β -Catenin signaling in endometrial cancer. *Int J Biochem Cell Biol* 109:17–22. <https://doi.org/10.1016/j.biocel.2019.01.014>
391. Yang X, Dong Y, Zhao J, Sun H, Deng Y, Fan J, Yan Q (2007) Increased expression of human macrophage metalloelastase (MMP-12) is associated with the invasion of endometrial adenocarcinoma. *Pathol Res Pract* 203(7):499–505. <https://doi.org/10.1016/j.prp.2007.03.008>
392. Rojo JV, González LO, Lamelas ML, Merino A, Vizoso F (2001) Apolipoprotein D expression in endometrial carcinomas. *Acta Obstet Gynecol Scand* 80(2):158–161. <https://doi.org/10.1034/j.1600-0412.2001.080002158.x>
393. Hong JH, Cho HW, Ouh YT, Lee JK, Chun Y (2023) Lymphocyte activation gene (LAG)-3 is a potential immunotherapeutic target for microsatellite stable, programmed death-ligand 1 (PD-L1)-positive endometrioid endometrial cancer. *J Gynecol Oncol* 34(2):e18. <https://doi.org/10.3802/jgo.2023.34.e18>
394. Zhong W, Liu Y, Zhang L, Zhuang W, Chen J, Huang Z, Zheng Y, Huang Y (2023) Combination of serum CST1 and HE4 for early diagnosis of endometrial cancer. *PeerJ* 11:e16424. <https://doi.org/10.7717/peerj.16424>
395. Panda H, Pelakh L, Chuang TD, Luo X, Bukulmez O, Chegini N (2012) Endometrial miR-200c is altered during transformation into cancerous states and targets the expression of ZEBs, VEGFA, FLT1, IKK β , KLF9, and FBLN5. *Reprod Sci* 19(8):786–796. <https://doi.org/10.1177/1933719112438448>
396. Chen CH, Weng TH, Huang KY, Kao HJ, Liao KW, Weng SL (2022) Anti-cancer peptide Q7 suppresses the growth and migration of human endometrial cancer by inhibiting DHCR24 expression and modulating the AKT-mediated pathway. *Int J Med Sci* 19(14):2008–2021. <https://doi.org/10.7150/ijms.78349>
397. Gu W, Mitsuhashi A, Kobayashi T, Shozu M (2022) Metformin attenuates the production and proliferative effects of prolactin induced by medroxyprogesterone acetate during fertility-sparing treatment for endometrial cancer. *BMC Cancer* 22(1):753. <https://doi.org/10.1186/s12885-022-09858-w>
398. Wasniewski T, Kiezun J, Krazinski BE, Kowalczyk AE, Szostak B, Wierzbicki PM, Kiewisz J (2019) WNT5A gene and protein expression in endometrial cancer. *Folia Histochem Cytobiol* 57(2):84–93. <https://doi.org/10.5603/FHC.a2019.0010>
399. Catusus L, Pons C, Muñoz J, Espinosa I, Prat J (2013) Promoter hypermethylation contributes to TIMP3 down-regulation in high stage endometrioid endometrial carcinomas. *Histopathology* 62(4):632–641. <https://doi.org/10.1111/his.12047>
400. Guan L, Wang Y, Cheng J, Zhang J, Kang S (2023) Expression and clinical significance of HER2/neu, aromatase P450 and adhesion molecule CD24 in endometrial cancer. *Eur J Histochem* 67(3):3655. <https://doi.org/10.4081/ejh.2023.3655>
401. Kölbl AC, Birk AE, Kuhn C, Jeschke U, Andergassen U (2016) Influence of VEGFR and LHCGR on endometrial adenocarcinoma. *Oncol Lett* 12(3):2092–2098. <https://doi.org/10.3892/ol.2016.4906>
402. Yue M, Hu J, Min X, Xu H (2023) Uterine tumor resembling high-grade endometrial mesenchymal sarcoma with GATAD2B-MMRN1 fusion. *Int J Clin Exp Pathol* 16(9):252–258
403. Palomero J, Panisello C, Lozano-Rabella M, Tirtakusuma R, Díaz-Gómez J, Grases D, Pasamar H, Arregui L, Dorca Duch E, Guerra Fernández E et al (2022) Biomarkers of tumor-reactive CD4+ and CD8+ TILs associate with improved prognosis in endometrial cancer. *J Immunother Cancer* 10(12):e005443. <https://doi.org/10.1136/jitc-2022-005443>
404. Yilmaz E, Melekoglu R, Taskapan C, Olmez Budak F, Toprak S (2020) The investigation of serum levels of ADAMTS 5 and 8 (the A disintegrin and metalloproteinase with thrombospondin motifs) in the etiology of endometrial cancer. *J Obstet Gynaecol* 40(6):856–859. <https://doi.org/10.1080/01443615.2019.1674265>
405. Keightley MC, Sales KJ, Jabbour HN (2010) PGF2 α -F-prostanoid receptor signalling via ADAMTS1 modulates epithelial cell invasion and endothelial cell function in endometrial cancer. *BMC Cancer* 10:488. <https://doi.org/10.1186/1471-2407-10-488>
406. Xue T, Liu X, Zhang M (2021) PAD12-catalyzed MEK1 citrullination activates ERK1/2 and promotes IGF2BP1-mediated SOX2 mRNA stability in endometrial cancer. *Adv Sci (Weinh)* 8(6):2002831. <https://doi.org/10.1002/adv.202002831>
407. Zheng X, Xu K, Zhu L, Mao M, Zhang F, Cui L (2020) MIR-486-5p act as a biomarker in endometrial carcinoma: promotes cell proliferation, migration, invasion by targeting MARK1. *Onco Targets Ther* 13:4843–4853. <https://doi.org/10.2147/OTT.S246841>
408. Wójcik-Krowiranda KM, Szczepaniec S, Bieńkiewicz A (2018) The role of the β klotho gene in uterine endometrial cancer. *Ginekol Pol* 89(10):563–567. <https://doi.org/10.5603/GPa.2018.0096>
409. Nordengren J, Casslén B, Gustavsson B, Einarsdottir M, Willén R (1998) Discordant expression of mRNA and protein for urokinase and tissue plasminogen activators (u-PA, t-PA) in endometrial carcinoma. *Int J Cancer* 79(2):195–201. [https://doi.org/10.1002/\(sici\)1097-0215\(19980417\)79:2%3c195::aid-ijc16%3e3.0.co;2-9](https://doi.org/10.1002/(sici)1097-0215(19980417)79:2%3c195::aid-ijc16%3e3.0.co;2-9)
410. Tian W, Li Z, Bai L, Chen L, Yan Y, Li H, Han Y, Teng F, Gao C, Xue F et al (2020) The oncogenic role of SOX8 in endometrial carcinoma. *Cancer Biol Ther* 21(12):1136–1144. <https://doi.org/10.1080/15384047.2020.1840318>
411. Egan D, Moran B, Wilkinson M, Pinyol M, Guerra E, Gatius S, Matias-Guiu X, Kolch W, le Roux CW, Brennan DJ (2022) CRABP2 - A novel biomarker for high-risk endometrial cancer. *Gynecol Oncol* 167(2):314–322. <https://doi.org/10.1016/j.ygyno.2022.09.020>
412. Orton S, Karkia R, Mustafov D, Gharanej S, Braoudaki M, Filipe A, Panfilov S, Saravi S, Khan N, Kyrou I et al (2024) In silico and in vitro mapping of receptor-type protein tyrosine phosphatase receptor type D in health and disease: implications for asprosin signalling in endometrial cancer and neuroblastoma. *Cancers (Basel)* 16(3):582. <https://doi.org/10.3390/cancers16030582>
413. Wen KC, Sung PL, Chou YT, Pan CM, Wang PH, Lee OK, Wu CW (2018) The role of EpCAM in tumor progression and the clinical prognosis of endometrial carcinoma. *Gynecol Oncol* 148(2):383–392. <https://doi.org/10.1016/j.ygyno.2017.11.033>
414. Xu Q, Zhou W, Zhou Y, Zhang X, Jiang R, Ai Z, Chen J, Ma L (2024) IRX2 regulates endometrial carcinoma oncogenesis by transcriptional

- repressing RUVBL1. *Exp Cell Res* 434(1):113866. <https://doi.org/10.1016/j.yexcr.2023.113866>
415. Peszek W, Kras P, Grabarek BO, Boroń D, Oplawski M (2020) Cisplatin changes expression of SEMA3B in endometrial cancer. *Curr Pharm Biotechnol* 21(13):1368–1376. <https://doi.org/10.2174/1389201021666200514215839>
 416. Spyrou I, Sifakis S, Ploumidis A, Papalampros AE, Felekouras E, Tsatsakis AM, Spandidos DA, Androutsopoulos VP (2014) Expression profile of CYP1A1 and CYP1B1 enzymes in endometrial tumors. *Tumour Biol* 35(10):9549–9556. <https://doi.org/10.1007/s13277-014-2240-2>
 417. Ding J, Li XM, Liu SL, Zhang Y, Li T (2014) Overexpression of platelet-derived growth factor-D as a poor prognosticator in endometrial cancer. *Asian Pac J Cancer Prev* 15(8):3741–3745. <https://doi.org/10.7314/apjcp.2014.15.8.3741>
 418. Boroń D, Nowakowski R, Grabarek BO, Zmarzły N, Oplawski M (2021) Expression pattern of leptin and its receptors in endometrioid endometrial cancer. *J Clin Med* 10(13):2787. <https://doi.org/10.3390/jcm10132787>
 419. Wu C, Li T, Cheng W (2022) The correlation between APOE expression and the clinical characteristics and prognosis of patients with endometrial cancer. *Medicine (Baltimore)* 101(37):e30536. <https://doi.org/10.1097/MD.00000000000030536>
 420. Yang Y, Liu PY, Bao W, Chen SJ, Wu FS, Zhu PY (2020) Hydrogen inhibits endometrial cancer growth via a ROS/NLRP3/caspase-1/GSDMD-mediated pyroptotic pathway. *BMC Cancer* 20(1):28. <https://doi.org/10.1186/s12885-019-6491-6>
 421. Ma X, Xia M, Wei L, Guo K, Sun R, Liu Y, Qiu C, Jiang J (2022) ABX-1431 inhibits the development of endometrial adenocarcinoma and reverses progesterone resistance by targeting MGLL. *Cell Death Dis* 13(12):1067. <https://doi.org/10.1038/s41419-022-05507-z>
 422. Bayramoglu Z, Kilinc ANU, Omeroglu E, Yilmaz F, Bayramoglu D, Unlu Y, Aydin HA (2022) Expression of extracellular matrix proteins nidogen-1 and legumain in endometrial carcinomas. *J Obstet Gynaecol Res* 48(4):1019–1025. <https://doi.org/10.1111/jog.15158>
 423. Pavlič R, Vidic S, Anko M, Knific T, Büdefeld T, Marton K, Sinreich M, Poschner S, Jäger W, Frković-Grazio S et al (2021) Altered profile of E1-S transporters in endometrial cancer: lower protein levels of ABCG2 and OSTβ and up-regulation of SLC01B3 expression. *Int J Mol Sci* 22(8):3819. <https://doi.org/10.3390/ijms22083819>
 424. Raba G, Zawlik I, Braun M, Paszek S, Potocka N, Skrzypa M, Obrzut B, Kluza M, Kluza K, Zych B et al (2020) Evaluation of the association between angiotensin converting enzyme insertion/deletion polymorphism and the risk of endometrial cancer in and characteristics of Polish women. *Adv Clin Exp Med* 29(5):581–585. <https://doi.org/10.17219/acem/118843>
 425. Przewoźny S, Rogaliński J, de Mezer M, Markowska A, Markowska J, Żurawski J (2024) Estrogen receptor (ER) and progesterone receptor (PgR) expression in endometrial cancer—an immunohistochemical assessment. *Diagnostics (Basel)* 14(3):322. <https://doi.org/10.3390/diagnostics14030322>
 426. Signorini Filho RC, de Azevedo Focchi GR, Theodoro TR, Pinhal MA, Nicolau SM (2015) Immunohistochemical expression of heparanases 1 and 2 in benign tissue and in invasive neoplasia of the endometrium: a case-control study. *Int J Gynecol Cancer* 25(2):269–278. <https://doi.org/10.1097/IGC.0000000000000329>
 427. Cai G, Sun W, Bi F, Wang D, Yang Q (2021) Knockdown of LMTK3 in the endometrioid adenocarcinoma cell line ishikawa: inhibition of growth and estrogen receptor α. *Front Oncol* 11:692282. <https://doi.org/10.3389/fonc.2021.692282>
 428. Miggiano GA, Martorana GE, Mordente A, Castelli A (1985) Biochemical properties of alkaline phosphatase from endometrial cancer cells. *Enzyme* 34(3):113–121. <https://doi.org/10.1159/000469373>
 429. Garrett AA, Bai S, Cascio S, Gupta N, Yang D, Buckanovich RJ (2024) EGFL6 promotes endometrial cancer cell migration and proliferation. *Gynecol Oncol*. <https://doi.org/10.1016/j.ygyno.2024.02.016>
 430. Kong X, Li M, Shao K, Yang Y, Wang Q, Cai M (2020) Progesterone induces cell apoptosis via the CACNA2D3/Ca2+/p38 MAPK pathway in endometrial cancer. *Oncol Rep* 43(1):121–132. <https://doi.org/10.3892/or.2019.7396>
 431. Gao Q, Huang Q, Li F, Luo F (2021) LncRNA MCTP1-AS1 regulates EMT process in endometrial cancer by targeting the miR-650/SMAD7 axis. *Oncotargets Ther* 14:751–761. <https://doi.org/10.2147/OTT.S240010>
 432. Guo J, Ye F, Xie W, Zhang X, Zeng R, Sheng W, Mi Y, Sheng X (2022) The HOXC-AS2/miR-876-5p/HKDC1 axis regulates endometrial cancer progression in a high glucose-related tumor microenvironment. *Cancer Sci* 113(7):2297–2310. <https://doi.org/10.1111/cas.15384>
 433. Ren W, Chi YB, Sun JL (2021) Effect of shRNA-mediated regulation of S100A4 gene expression on proliferation and apoptosis of KLE endometrial cancer cells. *Clin Transl Oncol* 23(1):148–154. <https://doi.org/10.1007/s12094-020-02406-7>
 434. Wang W, Jin W, Liu X, Zheng L (2022) Circ_0002577/miR-126-5p/MACC1 axis promotes endometrial carcinoma progression by regulation of proliferation, migration, invasion, and apoptosis of endometrial carcinoma cells. *Arch Gynecol Obstet* 306(2):481–491. <https://doi.org/10.1007/s00404-022-06412-8>
 435. Beeghly-Fadiel A, Xiang YB, Deming SL, Long JR, Xu WH, Cai Q, Zheng W, Shu XO (2009) No association between matrix metalloproteinase (MMP)-1, MMP-3, and MMP-7 SNPs and endometrial cancer risk. *Cancer Epidemiol Biomarkers Prev* 18(6):1925–1928. <https://doi.org/10.1158/1055-9965.EPI-09-0244>
 436. Stehbens SJ, Ju RJ, Adams MN, Perry SR, Haass NK, Bryant DM, Pollock PM (2018) FGFR2-activating mutations disrupt cell polarity to potentiate migration and invasion in endometrial cancer cell models. *J Cell Sci* 131(15):jcs213678. <https://doi.org/10.1242/jcs.213678>
 437. Zeng X, Li J, Kang LN, Xi MR, Liao GD (2020) Potential clinical value of interleukin-31 and interleukin-33 with their receptors expression as diagnostic and predictive factors in endometrial cancer: a case-control study. *Int J Clin Exp Pathol* 13(6):1324–1332
 438. Zhu Q, Tang M, Wu L (2020) Expression of combined interference of slug and FoxC2 in endometrial carcinoma and its clinicopathological relationship. *Transl Cancer Res* 9(9):5268–5280. <https://doi.org/10.21037/tcr-20-809>
 439. Liang M, Liu C, Lei T, Guo S, Min J (2022) Effect of integrin α7 on cell proliferation, invasion, apoptosis and the PI3K/AKT pathway, and its association with clinicopathological features in endometrial cancer. *Oncol Lett* 25(1):26. <https://doi.org/10.3892/ol.2022.13612>
 440. Yang T, Zhang H, Qiu H, Li B, Wang J, Du G, Ren C, Wan X (2016) EFEMP1 is repressed by estrogen and inhibits the epithelial-mesenchymal transition via Wnt/β-catenin signaling in endometrial carcinoma. *Oncotarget* 7(18):25712–25725. <https://doi.org/10.18632/oncotarget.8263>
 441. Zhao Y, Zou X, Wang G, Liu Y, Zhang C, Lu W, Li Q (2021) Effects of GATA6-AS/MMP9 on malignant progression of endometrial carcinoma. *J BUON* 26(5):1789–1795
 442. Asanoma K, Hori E, Yoshida S, Yagi H, Onoyama I, Kodama K, Yasunaga M, Ohgami T, Kaneki E, Okugawa K et al (2019) Mutual suppression between BHLHE40/BHLHE41 and the MIR301B-MIR130B cluster is involved in epithelial-to-mesenchymal transition of endometrial cancer cells. *Oncotarget* 10(45):4640–4654. <https://doi.org/10.18632/oncotarget.27061>
 443. Li Q, Lei Y, Du W (2018) A novel target of p53, TCF21, can respond to hypoxia by MAPK pathway inactivation in uterine corpus endometrial carcinoma. *DNA Cell Biol* 37(5):473–480. <https://doi.org/10.1089/dna.2017.4062>
 444. Fan J, Zhou H (2022) Comprehensive analysis of GDF10 methylation site-associated genes as prognostic markers for endometrial cancer. *J Oncol* 2022:7117083. <https://doi.org/10.1155/2022/7117083>
 445. Yoshida H, Uno M, Ogimoto K, Kobayashi-Kato M, Tanase Y, Ishikawa M, Kato T (2023) Endometrioid endometrial carcinoma with NKX31 expression in a transgender man: a case report. *Int J Gynecol Pathol* 42(3):308–314. <https://doi.org/10.1097/PGP.0000000000000869>
 446. Hojnik M, Kenda Šuster N, Smrkolj Š, Frković Grazio S, Verdenik I, Rižner TL (2020) AKR1C3 is associated with better survival of patients with endometrial carcinomas. *J Clin Med* 9(12):4105. <https://doi.org/10.3390/jcm9124105>
 447. Peng TF, Zhou YJ, Zhou J, Zhou Y, Li XC, Ouyang Q (2022) Long non-coding RNA VPS9D1-AS1 enhances proliferation, invasion, and epithelial-mesenchymal transition in endometrial cancer via miR-377-3p/SGK1. *Kaohsiung J Med Sci* 38(11):1048–1059. <https://doi.org/10.1002/kjm2.12606>

448. Tamate M, Tanaka R, Osogami H, Matsuura M, Satohisa S, Iwasaki M, Saito T (2017) Rap1GAP inhibits tumor progression in endometrial cancer. *Biochem Biophys Res Commun* 485(2):476–483. <https://doi.org/10.1016/j.bbrc.2017.02.044>
449. Song W, Zhang T, Li W, Mu R, Zhang L, Li Y, Jin B, Wang N, Li A, Cui J (2015) Overexpression of FLI-1 is associated with adverse prognosis of endometrial cancer. *Cancer Invest* 33(9):469–475. <https://doi.org/10.3109/07357907.2015.1069831>
450. Degasper C, Brunner A, Sampson N, Tsubulak I, Wieser V, Welpöner H, Marth C, Fiegl H, Zeimet AG (2019) NADPH oxidase 4 expression in the normal endometrium and in endometrial cancer. *Tumour Biol* 41(2):1010428319830002. <https://doi.org/10.1177/1010428319830002>
451. Shen Y, Wang X, Xu J, Lu L (2017) SerpinE2, a poor biomarker of endometrial cancer, promotes the proliferation and mobility of EC cells. *Cancer Biomark* 19(3):271–278. <https://doi.org/10.3233/CBM-160442>
452. Shi Z, Li C, Tarwater L, Li J, Li Y, Kaliney W, Chandrashekar DS, Stack MS (2018) RNA-seq reveals the overexpression of IGSF9 in endometrial cancer. *J Oncol* 2018:2439527. <https://doi.org/10.1155/2018/2439527>
453. Huang Z, Shen F, Chen J, Xie B, Chen X, Zhao Y, Chen S (2024) LncRNA linc01194 promotes the progress of endometrial carcinoma by up-regulating SOX2 through binding to IGF2BP1. *J Gynecol Oncol* 35(2):e21. <https://doi.org/10.3802/jgo.2024.35.e21>
454. Dyhdalo KS, Ababneh E, Lanigan C, Bowers K, Zhang S, McKenney JK, Joehlin-Price AS (2023) Evaluation of lineage/site-specific nuclear immunohistochemical markers SATB2, Cyclin D1, SALL4, and BCOR in high-grade endometrial carcinomas. *Int J Gynecol Pathol* 42(5):443–450. <https://doi.org/10.1097/PGP.0000000000000922>
455. An HJ, Song DH (2019) Displacement of vitamin D receptor is related to lower histological grade of endometrioid carcinoma. *Anticancer Res* 39(8):4143–4147. <https://doi.org/10.21873/anticancer.13573>
456. Qiao Z, Jiang Y, Wang L, Wang L, Jiang J, Zhang J (2019) Mutations in KIAA1109, CACNA1C, BSN, AKAP13, CELSR2, and HELZ2 are associated with the prognosis in endometrial cancer. *Front Genet* 10:909. <https://doi.org/10.3389/fgene.2019.00909>
457. He L, He W, Luo J, Xu M (2022) Upregulated ENC1 predicts unfavorable prognosis and correlates with immune infiltration in endometrial cancer. *Front Cell Dev Biol* 10:919637. <https://doi.org/10.3389/fcell.2022.919637>
458. Li Y, Liu J, Piao J, Ou J, Zhu X (2021) Circ_0109046 promotes the malignancy of endometrial carcinoma cells through the microRNA-105/SOX9/Wnt/ β -catenin axis. *IUBMB Life* 73(1):159–176. <https://doi.org/10.1002/iub.2415>
459. Wang J, Song T, Zhou S, Kong X (2019) YAP promotes the malignancy of endometrial cancer cells via regulation of IL-6 and IL-11. *Mol Med* 25(1):32. <https://doi.org/10.1186/s10020-019-0103-4>
460. Bruce SF, Cho K, Noia H, Lomonosova E, Stock EC, Opl A, Blachut B, Mullen MM, Kuroki LM, Hagemann AR et al (2022) GAS6-AXL inhibition by AVB-500 overcomes resistance to paclitaxel in endometrial cancer by decreasing tumor cell glycolysis. *Mol Cancer Ther* 21(8):1348–1359. <https://doi.org/10.1158/1535-7163.MCT-21-0704>
461. Cheng X, Shen C, Liao Z (2023) KLF2 transcription suppresses endometrial cancer cell proliferation, invasion, and migration through the inhibition of NPM1. *J Obstet Gynaecol* 43(2):2238827. <https://doi.org/10.1080/01443615.2023.2238827>
462. Zhang Z, Li B, Wang Z, Yang L, Peng J, Wang H, Wang Y, Hong L (2024) Novel LncRNA LINC02936 suppresses ferroptosis and promotes tumor progression by interacting with SIX1/CP axis in endometrial cancer. *Int J Biol Sci* 20(4):1356–1374. <https://doi.org/10.7150/ijbs.86256>
463. Gribben L, Baxter RC, Marsh DJ (2012) Insulin-like growth factor binding protein-3 inhibits migration of endometrial cancer cells. *Cancer Lett* 317(1):41–48. <https://doi.org/10.1016/j.canlet.2011.11.011>
464. Koukourakis MI, Giatromanolaki A, Sivridis E, Simopoulos C, Gatter KC, Harris AL, Jackson DG (2005) LYVE-1 immunohistochemical assessment of lymphangiogenesis in endometrial and lung cancer. *J Clin Pathol* 58(2):202–206. <https://doi.org/10.1136/jcp.2004.019174>
465. Shi Y, Zha J, Zuo M, Yan Q, Song H (2020) Long noncoding RNA CHL1-AS1 promotes cell proliferation and migration by sponging miR-6076 to regulate CHL1 expression in endometrial cancer. *J Cell Biochem* 121(3):2655–2663. <https://doi.org/10.1002/jcb.29486>
466. Sun H, Wang X, Zhang Y, Che X, Liu Z, Zhang L, Qiu C, Lv Q, Jiang J (2016) Biglycan enhances the ability of migration and invasion in endometrial cancer. *Arch Gynecol Obstet* 293(2):429–438. <https://doi.org/10.1007/s00404-015-3844-5>
467. Sirohi VK, Popli P, Sankhwar P, Kaushal JB, Gupta K, Manohar M, Dwivedi A (2017) Curcumin exhibits anti-tumor effect and attenuates cellular migration via Slit-2 mediated down-regulation of SDF-1 and CXCR4 in endometrial adenocarcinoma cells. *J Nutr Biochem* 44:60–70. <https://doi.org/10.1016/j.jnutbio.2016.12.021>
468. Oplawski M, Dziobek K, Grabarek B, Zmarzły N, Dąbrus D, Januszyk P, Brus R, Tomala B, Boroń D (2019) Expression of NRP-1 and NRP-2 in endometrial cancer. *Curr Pharm Biotechnol* 20(3):254–260. <https://doi.org/10.2174/1389201020666190219121602>
469. Wen Q, Xie X, Chen C, Wen B, Liu Y, Zhou J, Lin X, Jin H, Shi K (2023) Lipid reprogramming induced by the NNMT-ABCA1 axis enhanced membrane fluidity to promote endometrial cancer progression. *Aging (Albany NY)* 15(21):11860–11874. <https://doi.org/10.18632/aging.205142>
470. Liang M, Wang H, Liu C, Lei T, Min J (2020) LncRNA RUNX1-IT1 is downregulated in endometrial cancer and binds to miR-21 precursor to suppress its maturation. *Cancer Manag Res* 12:13451–13459. <https://doi.org/10.2147/CMAR.S272165>
471. Whitcomb BP, Mutch DG, Herzog TJ, Rader JS, Gibb RK, Goodfellow PJ (2003) Frequent HOXA11 and THBS2 promoter methylation, and a methylator phenotype in endometrial adenocarcinoma. *Clin Cancer Res* 9(6):2277–2287
472. Xing M, Wu B, Wang S (2023) Heat shock protein B7 inhibits the progression of endometrial carcinoma by inhibiting PI3K/AKT/mTOR pathway. *Reprod Sci* 30(2):590–600. <https://doi.org/10.1007/s43032-022-01041-7>
473. Srinivasan R, Benton E, McCormick F, Thomas H, Gullick WJ (1999) Expression of the c-erbB-3/HER-3 and c-erbB-4/HER-4 growth factor receptors and their ligands, neuregulin-1 alpha, neuregulin-1 beta, and betacellulin, in normal endometrium and endometrial cancer. *Clin Cancer Res* 5(10):2877–2883
474. Kawaguchi R, Maehana T, Sugimoto S, Kawahara N, Iwai K, Yamada Y, Kimura F (2024) Immunohistochemical analysis of the tissue factor pathway inhibitor-2 in endometrial clear cell carcinoma: a single-center retrospective study. *Int J Gynecol Pathol* 43(1):25–32. <https://doi.org/10.1097/PGP.0000000000000956>
475. Nykopp TK, Rilla K, Tammi MI, Tammi RH, Sironen R, Hämäläinen K, Kosma VM, Heinonen S, Anttila M (2010) Hyaluronan synthases (HAS1-3) and hyaluronidases (HYAL1-2) in the accumulation of hyaluronan in endometrioid endometrial carcinoma. *BMC Cancer* 10:512. <https://doi.org/10.1186/1471-2407-10-512>
476. Sun J, Ji G, Xie J, Jiao Z, Zhang H, Chen J (2019) Six-transmembrane epithelial antigen of the prostate 1 is associated with tumor invasion and migration in endometrial carcinomas. *J Cell Biochem* 120(7):11172–11189. <https://doi.org/10.1002/jcb.28393>
477. Yan J, Ye G, Shao Y (2022) High expression of the ferroptosis-associated MGST1 gene in relation to poor outcome and maladjusted immune cell infiltration in uterine corpus endometrial carcinoma. *J Clin Lab Anal* 36(4):e24317. <https://doi.org/10.1002/jcla.24317>
478. Feng S, Lu Y, Sun L, Hao S, Liu Z, Yang F, Zhang L, Wang T, Jiang L, Zhang J et al (2022) MiR-95-3p acts as a prognostic marker and promotes cervical cancer progression by targeting VCAM1. *Ann Transl Med* 10(21):1171. <https://doi.org/10.21037/atm-22-5184>
479. Li W, Song Y, Pan C, Yu J, Zhang J, Zhu X (2021) Aquaporin-8 is a novel marker for progression of human cervical cancer cells. *Cancer Biomark* 32(3):391–400. <https://doi.org/10.3233/CBM-203251>
480. Romanová M, Židlický V, Javůrková V, Kondé A, Šimětka O, Klát J (2023) L1CAM is not a predictive factor in early-stage squamous-cell cervical cancer. *In Vivo* 37(5):2334–2339. <https://doi.org/10.21873/in vivo.13337>
481. Li G, Wu Q, Gong L, Xu X, Cai J, Xu L, Zeng Y, He X, Wang Z (2021) FABP4 is an independent risk factor for lymph node metastasis and poor prognosis in patients with cervical cancer. *Cancer Cell Int* 21(1):568. <https://doi.org/10.1186/s12935-021-02273-4>
482. Rodríguez-Esquivel M, Romero-Morelos P, Taniguchi-Ponciano K, Mendoza-Rodríguez M, Marrero-Rodríguez D, Bandera-Delgado A, Huerta-Padilla V, Serna-Reyna L, Gómez-Gutiérrez G, Gómez-Virgilio L et al (2020) Expression of pregnancy specific β -1 glycoprotein 1 in cervical cancer cells. *Arch Med Res* 51(6):504–514. <https://doi.org/10.1016/j.jarmed.2020.05.025>

483. Zheng H, Liu M, Shi S, Huang H, Yang X, Luo Z, Song Y, Xu Q, Li T, Xue L et al (2024) MAP4K4 and WT1 mediate SOX6-induced cellular senescence by synergistically activating the ATF2-TGF β 2-Smad2/3 signaling pathway in cervical cancer. *Mol Oncol*. <https://doi.org/10.1002/1878-0261.13613>
484. Lin CL, Ying TH, Yang SF, Chiou HL, Chen YS, Kao SH, Hsieh YH (2021) MTA2 silencing attenuates the metastatic potential of cervical cancer cells by inhibiting AP1-mediated MMP12 expression via the ASK1/MEK3/p38/YB1 axis. *Cell Death Dis* 12(5):451. <https://doi.org/10.1038/s41419-021-03729-1>
485. Zhang Y, Qin Y, Li D, Yang Y (2022) A risk prediction model mediated by genes of APOD/APOC1/SQLE associates with prognosis in cervical cancer. *BMC Womens Health* 22(1):534. <https://doi.org/10.1186/s12905-022-02083-4>
486. Li Y, Wang W, Tian J, Zhou Y, Shen Y, Wang M, Tang L, Liu C, Zhang X, Shen F et al (2023) Clinical significance of soluble LAG-3 (sLAG-3) in patients with cervical cancer determined via an enzyme-linked immunosorbent assay with monoclonal antibodies. *Technol Cancer Res Treat* 22:15330338231202650. <https://doi.org/10.1177/15330338231202650>
487. Petrovic I, Milivojevic M, Popovic J, Schwirtlich M, Rankovic B, Stevanovic M (2015) SOX18 is a novel target gene of hedgehog signaling in cervical carcinoma cell lines. *PLoS ONE* 10(11):e0143591. <https://doi.org/10.1371/journal.pone.0143591>
488. Yang SH, Wang XL, Cai J, Wang SH (2020) Diagnostic value of circulating PIGF in combination with Flt-1 in early cervical cancer. *Curr Med Sci* 40(5):973–978. <https://doi.org/10.1007/s11596-020-2269-y>
489. Zhang C, Liao Y, Liu P, Du Q, Liang Y, Ooi S, Qin S, He S, Yao S, Wang W (2020) FABP5 promotes lymph node metastasis in cervical cancer by reprogramming fatty acid metabolism. *Theranostics* 10(15):6561–6580. <https://doi.org/10.7150/thno.44868>
490. Huang J, Yang J, Zhang Y, Lu D, Dai Y (2023) FTO promotes cervical cancer cell proliferation, colony formation, migration and invasion via the regulation of the BMP4/Hippo/YAP1/TAZ pathway. *Exp Cell Res* 427(1):113585. <https://doi.org/10.1016/j.yexcr.2023.113585>
491. Ramírez De Arellano A, Riera Leal A, Lopez-Pulido EI, González-Lucano LR, Macías Barragan J, Del Toro AS, García-Chagollan M, Palafox-Sánchez CA, Muñoz-Valle JF, Pereira-Suárez AL (2018) A 60 kDa prolactin variant secreted by cervical cancer cells modulates apoptosis and cytokine production. *Oncol Rep* 39(3):1253–1260. <https://doi.org/10.3892/or.2018.6222>
492. Zhang M, Xu Q, Yan S, Li Z, Yan W, Jia X (2016) Suppression of forkhead box Q1 by microRNA-506 represses the proliferation and epithelial-mesenchymal transition of cervical cancer cells. *Oncol Rep* 35(5):3106–3114. <https://doi.org/10.3892/or.2016.4651>
493. Shao Y, Zhu F, Zhu S, Bai L (2021) HDAC6 suppresses microRNA-199a transcription and augments HPV-positive cervical cancer progression through Wnt5a upregulation. *Int J Biochem Cell Biol* 136:106000. <https://doi.org/10.1016/j.biocel.2021.106000>
494. Ko J, Ryu KS, Lee YH, Na DS, Kim YS, Oh YM, Kim IS, Kim JW (2002) Human secreted frizzled-related protein is down-regulated and induces apoptosis in human cervical cancer. *Exp Cell Res* 280(2):280–287. <https://doi.org/10.1006/excr.2002.5649>
495. Shen HW, Tan JF, Shang JH, Hou MZ, Liu J, He L, Yao SZ, He SY (2016) CPE overexpression is correlated with pelvic lymph node metastasis and poor prognosis in patients with early-stage cervical cancer. *Arch Gynecol Obstet* 294(2):333–342. <https://doi.org/10.1007/s00404-015-3985-6>
496. Li T, Feng R, Chen B, Zhou J (2023) EREG is a risk factor for the prognosis of patients with cervical cancer. *Front Med (Lausanne)* 10:1161835. <https://doi.org/10.3389/fmed.2023.1161835>
497. Yuan Y, Ye HQ, Ren QC (2018) Upregulation of the BDNF/TrkB pathway promotes epithelial-mesenchymal transition, as well as the migration and invasion of cervical cancer. *Int J Oncol* 52(2):461–472. <https://doi.org/10.3892/ijo.2017.4230>
498. Zhang J, Chen X, Bian L, Wang Y, Liu H (2019) CD44+/CD24+-expressing cervical cancer cells and radioresistant cervical cancer cells exhibit cancer stem cell characteristics. *Gynecol Obstet Invest* 84(2):174–182. <https://doi.org/10.1159/000493129>
499. Han C, Hu C, Liu T, Sun Y, Hu F, He Y, Zhang J, Chen J, Ding J, Fan J et al (2024) GF2BP3 enhances lipid metabolism in cervical cancer by upregulating the expression of SCD. *Cell Death Dis* 15(2):138. <https://doi.org/10.1038/s41419-024-06520-0>
500. Sun Y, Feng Y, Zhang G, Xu Y (2019) The endonuclease APE1 processes miR-92b formation, thereby regulating expression of the tumor suppressor LDLR in cervical cancer cells. *Ther Adv Med Oncol* 11:1758835919855859. <https://doi.org/10.1177/1758835919855859>
501. Grover S, Mehta P, Wang Q, Bhatia R, Bvochora-Nsingo M, Davey S, Iyengar M, Shah S, Shin SS, Zetola NM (2020) Association between CD4 count and chemoradiation therapy outcomes among cervical cancer patients with HIV. *J Acquir Immune Defic Syndr* 85(2):201–208. <https://doi.org/10.1097/QAI.0000000000002420>
502. Liu XL, Meng YH, Wang JL, Yang BB, Zhang F, Tang SJ (2014) FOXL2 suppresses proliferation, invasion and promotes apoptosis of cervical cancer cells. *Int J Clin Exp Pathol* 7(4):1534–1543
503. Dong M, Dong Z, Zhu X, Zhang Y, Song L (2019) Long non-coding RNA MIR205HG regulates KRT17 and tumor processes in cervical cancer via interaction with SRSF1. *Exp Mol Pathol* 111:104322. <https://doi.org/10.1016/j.yexmp.2019.104322>
504. Sato A, Ishiwata T, Matsuda Y, Yamamoto T, Asakura H, Takeshita T, Naito Z (2012) Expression and role of nestin in human cervical intraepithelial neoplasia and cervical cancer. *Int J Oncol* 41(2):441–448. <https://doi.org/10.3892/ijo.2012.147>
505. You X, Wang Y, Meng J, Han S, Liu L, Sun Y, Zhang J, Sun S, Li X, Sun W et al (2021) Exosomal miR-663b exposed to TGF- β 1 promotes cervical cancer metastasis and epithelial-mesenchymal transition by targeting MGAT3. *Oncol Rep* 45(4):12. <https://doi.org/10.3892/or.2021.7963>
506. Natalia MA, Alejandro GT, Virginia TJ, Alvarez-Salas LM (2018) MARK1 is a novel target for miR-125a-5p: implications for cell migration in cervical tumor cells. *Microna* 7(1):54–61. <https://doi.org/10.2174/2211536606666171024160244>
507. Aviel-Ronen S, Rubinek T, Zadok O, Vituri A, Avivi C, Wolf I, Barshack I (2016) Klotho expression in cervical cancer: differential expression in adenocarcinoma and squamous cell carcinoma. *J Clin Pathol* 69(1):53–57. <https://doi.org/10.1136/jclinpath-2015-202929>
508. Hu X, Mandika C, He L, You Y, Chang Y, Wang J, Chen T, Zhu X (2021) Correction to “construction of urokinase-type plasminogen activator receptor-targeted heterostructures for efficient photothermal chemotherapy against cervical cancer to achieve simultaneous anticancer and antiangiogenesis.” *ACS Appl Mater Interfaces* 13(44):53300. <https://doi.org/10.1021/acsami.1c19496>
509. Zhang W, Cao H, Yang J, Zhao J, Liang Z, Kang X, Wang R (2022) The identification and validation of EphA7 hypermethylation, a novel biomarker, in cervical cancer. *BMC Cancer* 22(1):636. <https://doi.org/10.1186/s12885-022-09653-7>
510. Sun Q, Yang Z, Li P, Wang X, Sun L, Wang S, Liu M, Tang H (2019) A novel miRNA identified in GRSF1 complex drives the metastasis via the PIK3R3/AKT/NF- κ B and TIMP3/MMP9 pathways in cervical cancer cells. *Cell Death Dis* 10(9):636. <https://doi.org/10.1038/s41419-019-1>
511. Zhao C, Liu J, Wu H, Hu J, Chen J, Chen J, Qiao F (2020) Aberrant methylation-mediated downregulation of lncRNA CCND2 AS1 promotes cell proliferation in cervical cancer. *J Biol Res (Thessalon)* 27:11. <https://doi.org/10.1186/s40709-020-00122-5>
512. Xu Z, Guo Y, Wang L, Cui J (2024) HECW1 restrains cervical cancer cell growth by promoting DVL1 ubiquitination and downregulating the activation of Wnt/ β -catenin signaling. *Exp Cell Res* 435(2):113949. <https://doi.org/10.1016/j.yexcr.2024.113949>
513. Chantima W, Thepthai C, Cheunsuchon P, Dharakul T (2017) EpCAM expression in squamous cell carcinoma of the uterine cervix detected by monoclonal antibody to the membrane-proximal part of EpCAM. *BMC Cancer* 17(1):811. <https://doi.org/10.1186/s12885-017-3798-z>
514. Zong Y, Chang Y, Huang K, Liu J, Zhao Y (2024) The role of BATF2 deficiency in immune microenvironment rearrangement in cervical cancer - New biomarker benefiting from combination of radiotherapy and immunotherapy. *Int Immunopharmacol* 126:111199. <https://doi.org/10.1016/j.intimp.2023.111199>
515. Wongprate M, Ishida W, Phuthong S, Natphopsuk S, Ishida T (2020) Genetic polymorphisms of the human cytochrome P450 1A1 (CYP1A1) and cervical cancer susceptibility among Northeast Thai women. *Asian Pac J Cancer Prev* 21(1):243–248. <https://doi.org/10.31557/APJCP.2020.21.1.243>

516. Han YQ, Ming SL, Wu HT, Zeng L, Ba G, Li J, Lu WF, Han J, Du QJ, Sun MM et al (2018) Myostatin knockout induces apoptosis in human cervical cancer cells via elevated reactive oxygen species generation. *Redox Biol* 19:412–428. <https://doi.org/10.1016/j.redox.2018.09.009>
517. Vidal AC, Skaar D, Maguire R, Dodor S, Musselwhite LW, Bartlett JA, Oneko O, Obure J, Mlay P, Murphy SK et al (2015) IL-10, IL-15, IL-17, and GM-CSF levels in cervical cancer tissue of Tanzanian women infected with HPV16/18 vs. non-HPV16/18 genotypes. *Infect Agent Cancer* 10:10. <https://doi.org/10.1186/s13027-015-0005-1>
518. Huang J, Xu W, Huang Q, Chen E, Chen J (2024) SYT7 (synaptotagmin 7) promotes cervical squamous cell carcinoma. *Heliyon* 10(3):e24806. <https://doi.org/10.1016/j.heliyon.2024.e24806>
519. Zhou N, Ding B, Agler M, Cockett M, McPhee F (2015) Lethality of PAK3 and SGK2 shRNAs to human papillomavirus positive cervical cancer cells is independent of PAK3 and SGK2 knockdown. *PLoS ONE* 10(1):e0117357. <https://doi.org/10.1371/journal.pone.0117357>
520. Okamoto S, Niikura H, Nakabayashi K, Hiyama K, Matoda M, Takeshima N, Watanabe M, Nagase S, Otsuki T, Yaegashi N (2013) Detection of sentinel lymph node metastases in cervical cancer: assessment of KRT19 mRNA in the one-step nucleic acid amplification (OSNA) method. *Gynecol Oncol* 130(3):530–536. <https://doi.org/10.1016/j.ygyno.2013.06.027>
521. Machida H, Matsuo K, Tanaka M, Kitatani K, Takase A, Yokoyama K, Kajiwara H, Yasaka M, Ikeda M, Yoshida H et al (2023) ROS1 as a possible prognostic biomarker of cervical adenocarcinoma: an exploratory analysis with next-generation sequencing. *Gynecol Oncol* 171:59–66. <https://doi.org/10.1016/j.ygyno.2023.02.006>
522. Xu J, Lu W (2021) CircSPIDR acts as a tumour suppressor in cervical adenocarcinoma by sponging miR-431-5p and regulating SORCS1 and CUBN expression. *Aging (Albany NY)* 13(14):18340–18359. <https://doi.org/10.18632/aging.203283>
523. Chakraborty C, Dutta S, Mukherjee N, Samadder S, Roychowdhury A, Roy A, Mondal RK, Basu P, Roychowdhury S, Panda CK (2015) Inactivation of PTCH1 is associated with the development of cervical carcinoma: clinical and prognostic implication. *Tumour Biol* 36(2):1143–1154. <https://doi.org/10.1007/s13277-014-2707-1>
524. Zeng L, Zhen Y, Chen Y, Zou L, Zhang Y, Hu F, Feng J, Shen J, Wei B (2014) Naringin inhibits growth and induces apoptosis by a mechanism dependent on reduced activation of NF- κ B/COX-2-caspase-1 pathway in HeLa cervical cancer cells. *Int J Oncol* 45(5):1929–1936. <https://doi.org/10.3892/ijo.2014.2617>
525. Ding M, Zhang H, Liu L, Liang R (2019) Effect of NOS1 regulating ABCG2 expression on proliferation and apoptosis of cervical cancer cells. *Oncol Lett* 17(2):1531–1536. <https://doi.org/10.3892/ol.2018.9786>
526. Baik S, Mehta FF, Unsal E, Park Y, Chung SH (2022) Estrogen inhibits epithelial progesterone receptor-dependent progestin therapy efficacy in a mouse model of cervical cancer. *Am J Pathol* 192(2):353–360. <https://doi.org/10.1016/j.ajpath.2021.10.008>
527. Marques RM, Focchi GR, Theodoro TR, Castelo A, Pinhal MA, Nicolau SM (2012) The immunoexpression of heparanase 2 in normal epithelium, intraepithelial, and invasive squamous neoplasia of the cervix. *J Low Genit Tract Dis* 16(3):256–262. <https://doi.org/10.1097/LGT.0b013e3182422c69>
528. Cao CH, Liu R, Lin XR, Luo JQ, Cao LJ, Zhang QJ, Lin SR, Geng L, Sun ZY, Ye SK et al (2021) LRP1B mutation is associated with tumor HPV status and promotes poor disease outcomes with a higher mutation count in HPV-related cervical carcinoma and head & neck squamous cell carcinoma. *Int J Biol Sci* 17(7):1744–1756. <https://doi.org/10.7150/ijbs.56970>
529. Yu J, Zheng Q, Ding X, Zheng B, Chen X, Chen B, Shen C, Zhang Y, Luan X, Yan Y et al (2019) Systematic re-analysis strategy of serum indices identifies alkaline phosphatase as a potential predictive factor for cervical cancer. *Oncol Lett* 18(3):2356–2365. <https://doi.org/10.3892/ol.2019.10527>
530. Guo WW, Feng MM, Li SF, Wei LH (2022) Circular RNA circ_0023404 serves as a miR-636 sponge to promote malignant behaviors in cervical cancer cells through upregulation of CYP251. *Kaohsiung J Med Sci* 38(3):218–229. <https://doi.org/10.1002/kjm2.12478>
531. Adiga D, Bhat S, Chakraborty S, Kabekkodu SP (2022) DOC2B is a negative regulator of Wnt/ β -catenin signaling pathway in cervical cancer. *Pharmacol Res* 180:106239. <https://doi.org/10.1016/j.phrs.2022.106239>
532. Zheng G, Wang Z, Fan Y, Wang T, Zhang L, Wang M, Chen S, Jiang L (2021) The clinical significance and immunization of MSMO1 in cervical squamous cell carcinoma based on bioinformatics analysis. *Front Genet* 12:705851. <https://doi.org/10.3389/fgene.2021.705851>
533. Xu J, Lu W (2021) CircSPIDR acts as a tumour suppressor in cervical adenocarcinoma by sponging miR-431-5p and regulating SORCS1 and CUBN expression. *Aging (Albany NY)* 13(14):18340–18359. <https://doi.org/10.18632/aging.203283>
534. Madeleine MM, Johnson LG, Smith AG, Hansen JA, Nisperos BB, Li S, Zhao LP, Daling JR, Schwartz SM, Galloway DA (2008) Comprehensive analysis of HLA-A, HLA-B, HLA-C, HLA-DRB1, and HLA-DQB1 loci and squamous cell cervical cancer risk. *Cancer Res* 68(9):3532–3539. <https://doi.org/10.1158/0008-5472.CAN-07-6471>
535. Lee CY, Hsin MC, Chen PN, Lin CW, Wang PH, Yang SF, Hsiao YH (2022) Arctin inhibits cervical cancer cell migration and invasion through suppression of S100A4 expression via PI3K/Akt pathway. *Pharmaceutics* 14(2):365. <https://doi.org/10.3390/pharmaceutics14020365>
536. Mei J, Zhu C, Pan L, Li M (2022) MACC1 regulates the AKT/STAT3 signaling pathway to induce migration, invasion, cancer stemness, and suppress apoptosis in cervical cancer cells. *Bioengineered* 13(1):61–70. <https://doi.org/10.1080/21655979.2021.2006567>
537. Xie B, Zhang Z, Wang H, Chen Z, Wang Y, Liang H, Yang G, Yang X, Zhang H (2016) Genetic polymorphisms in MMP 2, 3, 7, and 9 genes and the susceptibility and clinical outcome of cervical cancer in a Chinese Han population. *Tumour Biol* 37(4):4883–4888. <https://doi.org/10.1007/s13277-015-4204-6>
538. Gill CM, Orfanelli T, Yoxtheimer L, Roy-McMahon C, Suhner J, Tomita S, Kalir T, Liu Y, Houldsworth J, Kolev V (2020) Histology-specific FGFR2 alterations and FGFR2-TACC2 fusion in mixed adenoid cystic and neuroendocrine small cell carcinoma of the uterine cervix. *Gynecol Oncol Rep* 34:100668. <https://doi.org/10.1016/j.gore.2020.100668>
539. Zhang Y, Li J, Yang F, Zhang X, Ren X, Wei F (2022) Relationship and prognostic significance of IL-33, PD-1/PD-L1, and tertiary lymphoid structures in cervical cancer. *J Leukoc Biol* 112(6):1591–1603. <https://doi.org/10.1002/JLB.5MA0322-746R>
540. Faulkner S, Griffin N, Rowe CW, Jobling P, Lombard JM, Oliveira SM, Walker MM, Hondermarck H (2020) Nerve growth factor and its receptor tyrosine kinase TrkA are overexpressed in cervical squamous cell carcinoma. *FASEB Bioadv* 2(7):398–408. <https://doi.org/10.1096/fba.2020-00016>
541. Wang J, Yue X (2017) Role and importance of the expression of transcription factor FOXC2 in cervical cancer. *Oncol Lett* 14(6):6627–6631. <https://doi.org/10.3892/ol.2017.7004>
542. Backsch C, Rudolph B, Steinbach D, Scheungraber C, Liesenfeld M, Häfner N, Hildner M, Habenicht A, Runnebaum IB, Dürst M (2011) An integrative functional genomic and gene expression approach revealed SORBS2 as a putative tumour suppressor gene involved in cervical carcinogenesis. *Carcinogenesis* 32(7):1100–1106. <https://doi.org/10.1093/carcin/bgr093>
543. Yuan N, Wang L, Xi Q, Zou N, Zhang X, Lu X, Zhang Z (2022) ITGA7, CD133, ALDH1 are inter-correlated, and linked with poor differentiation, lymph node metastasis as well as worse survival in surgical cervical cancer. *J Obstet Gynaecol Res* 48(4):1011–1018. <https://doi.org/10.1111/jog.151>
544. Song EL, Hou YP, Yu SP, Chen SG, Huang JT, Luo T, Kong LP, Xu J, Wang HQ (2011) EFEMP1 expression promotes angiogenesis and accelerates the growth of cervical cancer in vivo. *Gynecol Oncol* 121(1):174–180. <https://doi.org/10.1016/j.ygyno.2010.11.004>
545. Zhao X, Zheng H, Chen J (2020) LncRNA GATA6-AS inhibits cancer cell proliferation and promotes cancer cell apoptosis in cervical cancer by down-regulating miR-205. *BMC Womens Health* 20(1):247. <https://doi.org/10.1186/s12905-020-01082-7>
546. Zhao Y, Dong X, Hou R (2020) LncRNA PICART1 alleviates progression of cervical cancer by upregulating TCF21. *Oncol Lett* 19(6):3719–3724. <https://doi.org/10.3892/ol.2020.11486>
547. Zou C, Xu F, Shen J, Xu S (2022) Identification of a ferroptosis-related prognostic gene PTGS2 based on risk modeling and immune microenvironment of early-stage cervical cancer. *J Oncol* 2022:3997562. <https://doi.org/10.1155/2022/3997562>

548. Zhang J, Tong Y, Ren L, Li CD (2014) Expression of metastasis suppressor 1 in cervical carcinoma and the clinical significance. *Oncol Lett* 8(5):2145–2149. <https://doi.org/10.3892/ol.2014.2508>
549. Zeng L, Chen C, Yao C (2021) Histone deacetylation regulated by KDM1A to suppress DACT1 in proliferation and migration of cervical cancer. *Anal Cell Pathol (Amst)* 2021:5555452. <https://doi.org/10.1155/2021/5555452>
550. Wang N, Che Y, Yin F, Yu F, Bi X, Wang Y (2018) Study on the methylation status of SPINT2 gene and its expression in cervical carcinoma. *Cancer Biomark* 22(3):435–442. <https://doi.org/10.3233/CBM-171050>
551. Stewart CJR, Moses J (2021) NKX3.1 expression in cervical “adenoid basal cell carcinoma”: another gynaecological lesion with prostatic differentiation? *Pathology* 53(2):193–198. <https://doi.org/10.1016/j.pathol.2020.07.011>
552. Huang Y, Chen L, Guo A (2018) Upregulated expression of HOXC8 is associated with poor prognosis of cervical cancer. *Oncol Lett* 15(5):7291–7296. <https://doi.org/10.3892/ol.2018.8200>
553. Wu CH, Ko JL, Chen SC, Lin YW, Han CP, Yang TY, Chien MH, Wang PH (2014) Clinical implications of aldo-keto reductase family 1 member C3 and its relationship with lipocalin 2 in cancer of the uterine cervix. *Gynecol Oncol* 132(2):474–482. <https://doi.org/10.1016/j.ygyno.2013.11.032>
554. Liu J, Yang L, Zhang J, Zhang J, Chen Y, Li K, Li Y, Li Y, Yao L, Guo G (2012) Knock-down of NDRG2 sensitizes cervical cancer HeLa cells to cisplatin through suppressing Bcl-2 expression. *BMC Cancer* 12:370. <https://doi.org/10.1186/1471-2407-12-370>
555. Bierkens M, Krijgsman O, Wiltling SM, Bosch L, Jaspers A, Meijer GA, Meijer CJ, Snijders PJ, Ylstra B, Steenbergen RD (2013) Focal aberrations indicate EYA2 and hsa-miR-375 as oncogene and tumor suppressor in cervical carcinogenesis. *Genes Chromosomes Cancer* 52(1):56–68. <https://doi.org/10.1002/gcc.22006>
556. Wu Z, Chen J, Yang L, Sun K, Jiang Q, Dong F, Lu W, Chen R, Chen Y (2024) Elevated INHBA promotes tumor progression of cervical cancer. *Technol Cancer Res Treat* 23:15330338241234798. <https://doi.org/10.1177/15330338241234798>
557. Yu J, Zheng Q, Ding X, Zheng B, Chen X, Chen B, Shen C, Zhang Y, Luan X, Yan Y et al (2019) Systematic re-analysis strategy of serum indices identifies alkaline phosphatase as a potential predictive factor for cervical cancer. *Oncol Lett* 18(3):2356–2365. <https://doi.org/10.3892/ol.2019.10527>
558. Wang M, Xue Y, Shen L, Qin P, Sang X, Tao Z, Yi J, Wang J, Liu P, Cheng H (2019) Inhibition of SGK1 confers vulnerability to redox dysregulation in cervical cancer. *Redox Biol* 24:101225. <https://doi.org/10.1016/j.redox.2019.101225>
559. Wang Y, Xie Y, Sun B, Guo Y, Song L, Mohammednur DE, Zhao C (2021) The degradation of Rap1GAP via E6AP-mediated ubiquitin-proteasome pathway is associated with HPV16/18-infection in cervical cancer cells. *Infect Agent Cancer* 16(1):71. <https://doi.org/10.1186/s13027-021-00409-9>
560. Gu YY, Zhou GN, Li Y, He HY, Ding JX, Hua KQ (2022) HDAC10 inhibits cervical cancer progression through downregulating the HDAC10-microRNA-223-EPB41L3 axis. *J Oncol* 2022:8092751. <https://doi.org/10.1155/2022/8092751>
561. Zidi S, Sghaier I, Gazouani E, Mezlini A, Yacoubi-Loueslati B (2016) Evaluation of toll-like receptors 2/3/4/9 gene polymorphisms in cervical cancer evolution. *Pathol Oncol Res* 22(2):323–330. <https://doi.org/10.1007/s12253-015-0009-6>
562. Rahmani F, Hasanazadeh M, Hassanian SM, Khazaei M, Esmaily H, Asef-Agah SA, Naghipour A, Ferns A (2020) Association of a genetic variant in the angiotensin-like protein 4 gene with cervical cancer. *Pathol Res Pract* 216(7):153011. <https://doi.org/10.1016/j.prp.2020.153011>
563. Duan S, Wu A, Chen Z, Yang Y, Liu L, Shu Q (2018) miR-204 regulates cell proliferation and invasion by targeting EphB2 in human cervical cancer. *Oncol Res* 26(5):713–723. <https://doi.org/10.3727/096504017X15016337254641>
564. Salmerón-Bárceñas EG, Mendoza-Catalan MA, Ramírez-Bautista ÁU, Lozano-Santos RA, Torres-Rojas FI, Ávila-López PA, Zacapala-Gómez AE (2023) Identification of Mir-182-3p/Fli-1 axis as a key signaling in immune response in cervical cancer: a comprehensive bioinformatic analysis. *Int J Mol Sci* 24(7):6032. <https://doi.org/10.3390/ijms24076032>
565. Tian R, Li H, Ren S, Li S, Fang R, Liu Y (2023) circRNA THBS1 silencing inhibits the malignant biological behavior of cervical cancer cells via the regulation of miR-543/HMGB2 axis. *Open Med (Wars)* 18(1):20230709. <https://doi.org/10.1515/med-2023-0709>
566. Wang Q, Lin B, Wei H, Wang X, Nie X, Shi Y (2024) AQP3 promotes the invasion and metastasis in cervical cancer by regulating NOX4-derived H₂O₂ activation of Syk/PI3K/Akt signaling axis. *J Cancer* 15(4):1124–1137. <https://doi.org/10.7150/jca.91360>
567. Tian P, Feng Y, Tao L (2024) LINC00460 knockdown sensitizes cervical cancer to cisplatin by downregulating TGFBI. *Chem Biol Drug Des* 103(1):e14424. <https://doi.org/10.1111/cbdd.14424>
568. Sui H, Shi C, Yan Z, Chen J, Man L, Wang F (2024) LRRC75A-AS1 drives the epithelial-mesenchymal transition in cervical cancer by binding IGF2BP1 and inhibiting SYVN1-mediated NLRP3 ubiquitination. *Mol Cancer Res*. <https://doi.org/10.1158/1541-7786.MCR-23-0478>
569. Chen M, Li L, Zheng PS (2019) SALL4 promotes the tumorigenicity of cervical cancer cells through activation of the Wnt/β-catenin pathway via CTNNB1. *Cancer Sci* 110(9):2794–2805. <https://doi.org/10.1111/cas.14140>
570. Dong H, Chen S, Liang X, Cai Q, Zhang X, Xie J, Sun Z (2024) Vitamin D and its receptors in cervical cancer. *J Cancer* 15(4):926–938. <https://doi.org/10.7150/jca.87499>
571. Wongwarangkana C, Wanlapakorn N, Chansaenroj J, Poovorawan Y (2018) Retinoic acid receptor beta promoter methylation and risk of cervical cancer. *World J Virol* 7(1):1–9. <https://doi.org/10.5501/wjv.v7.i1.1>
572. Kina S, Kinjo T, Liang F, Nakasone T, Yamamoto H, Arasaki A (2018) Targeting EphA4 abrogates intrinsic resistance to chemotherapy in well-differentiated cervical cancer cell line. *Eur J Pharmacol* 840:70–78. <https://doi.org/10.1016/j.ejphar.2018.09.031>
573. Zhang P, Zhao F, Jia K, Liu X (2022) The LOXL1 antisense RNA 1 (LOXL1-AS1)/microRNA-423-5p (miR-423-5p)/ectodermal-neural cortex 1 (ENC1) axis promotes cervical cancer through the mitogen-activated protein kinase (MEK)/extracellular signal-regulated kinase (ERK) pathway. *Bioengineered* 13(2):2567–2584. <https://doi.org/10.1080/21655979.2021.2018975>
574. Chen H, Chen X, Zeng F, Fu A, Huang M (2022) Prognostic value of SOX9 in cervical cancer: bioinformatics and experimental approaches. *Front Genet* 13:939328. <https://doi.org/10.3389/fgene.2022.939328>
575. Li J, Wang X, Li Z, Li M, Zheng X, Zheng D, Wang Y, Xi M (2023) SULF1 activates the VEGFR2/PI3K/AKT pathway to promote the development of cervical cancer. *Curr Cancer Drug Targets*. <https://doi.org/10.2174/1568009623666230804161607>
576. Yang S, Liu L, Xu D, Li X (2020) The relationship of the TLR9 and TLR2 genetic polymorphisms with cervical cancer risk: a meta-analysis of case-control studies. *Pathol Oncol Res* 26(1):307–315. <https://doi.org/10.1007/s12253-018-0465-x>
577. Hao Y, Yan Z, Zhang A, Hu S, Wang N, Luo XG, Ma W, Zhang TC, He H (2020) IL-6/STAT3 mediates the HPV18 E6/E7 stimulated upregulation of MALAT1 gene in cervical cancer HeLa cells. *Virus Res* 281:197907. <https://doi.org/10.1016/j.virusres.2020.197907>
578. Hu R, Zhu Z (2019) ELK1-activated GPC3-AS1/GPC3 axis promotes the proliferation and migration of cervical cancer cells. *J Gene Med* 21(8):e3099. <https://doi.org/10.1002/jgm.3099>
579. Ji H, Li K, Jiang W, Li J, Zhang JA, Zhu X (2022) MRV11 and NTRK3 are potential tumor suppressor genes commonly inactivated by DNA methylation in cervical cancer. *Front Oncol* 11:802068. <https://doi.org/10.3389/fonc.2021.802068>
580. Cao J, Dong J, Wang Y, Chen Y (2019) The expressions of DNA methyltransferase 1 (DNMT1) and cyclin A1 (CCNA1) in cervical carcinogenesis. *Int J Clin Exp Pathol* 12(1):40–49
581. Iida Y, Osaki M, Sato S, Izutsu R, Seong H, Okawa M, Osaku D, Komatsu H, Taniguchi F, Okada F (2023) AMIGO2 expression as a predictor of recurrence in cervical cancer with intermediate risk. *Mol Clin Oncol* 19(1):56. <https://doi.org/10.3892/mco.2023.2652>
582. Chang S, Sun L, Feng G (2019) SP1-mediated long noncoding RNA POU3F3 accelerates the cervical cancer through miR-127-5p/FOXO1. *Biomed Pharmacother* 117:109133. <https://doi.org/10.1016/j.biopha.2019.109133>
583. Wang J, Chen S (2020) RACK1 promotes miR-302b/c/d-3p expression and inhibits CCNO expression to induce cell apoptosis in cervical

- squamous cell carcinoma. *Cancer Cell Int* 20:385. <https://doi.org/10.1186/s12935-020-01435-0>
584. Ma X, Wu Z, Zhang J, Shao X, Shen H (2021) Increased ADAM12 expression predicts poor prognosis in cervical cancer patients before general anesthesia. *Clin Lab*. <https://doi.org/10.7754/Clin.Lab.2020.200611>
 585. Guerrero-Setas D, Pérez-Janices N, Blanco-Fernandez L, Ojer A, Cambra K, Berdasco M, Esteller M, Maria-Ruiz S, Torrea N, Gaurch R (2013) RASSF2 hypermethylation is present and related to shorter survival in squamous cervical cancer. *Mod Pathol* 26(8):1111–1122. <https://doi.org/10.1038/modpathol.2013.32>
 586. Zhao X, Dong W, Luo G, Xie J, Liu J, Yu F (2021) Silencing of hsa_circ_0009035 suppresses cervical cancer progression and enhances radiosensitivity through MicroRNA 889–3p-dependent regulation of HOXB7. *Mol Cell Biol* 41(6):e0063120. <https://doi.org/10.1128/MCB.00631-20>
 587. Wu L, Gong Y, Yan T, Zhang H (2020) LINP1 promotes the progression of cervical cancer by scaffolding EZH2, LSD1, and DNMT1 to inhibit the expression of KLF2 and PRSS8. *Biochem Cell Biol* 98(5):591–599. <https://doi.org/10.1139/bcb-2019-0446>
 588. Peng J, Hou F, Zhu W, Li J, Teng Z (2020) lncRNA NR2F1-AS1 regulates miR-17/SIK1 axis to suppress the invasion and migration of cervical squamous cell carcinoma cells. *Reprod Sci* 27(7):1534–1539. <https://doi.org/10.1007/s43032-020-00149-y>
 589. Liu X, Xie S, Zhang J, Kang Y (2020) Long noncoding RNA XIST contributes to cervical cancer development through targeting miR-889-3p/SIX1 Axis. *Cancer Biother Radiopharm* 35(9):640–649. <https://doi.org/10.1089/cbr.2019.3318>
 590. Shang A, Zhou C, Bian G, Chen W, Lu W, Wang W, Li D (2019) miR-381-3p restrains cervical cancer progression by downregulating FGF7. *J Cell Biochem* 120(1):778–789. <https://doi.org/10.1002/jcb.27438>
 591. Lee SW, Lee SY, Lee SR, Ju W, Kim SC (2010) Plasma levels of insulin-like growth factor-1 and insulin-like growth factor binding protein-3 in women with cervical neoplasia. *J Gynecol Oncol* 21(3):174–180. <https://doi.org/10.3802/jgo.2010.21.3.174>
 592. Chu Y, Ouyang Y, Wang F, Zheng A, Bai L, Han L, Chen Y, Wang H (2014) MicroRNA-590 promotes cervical cancer cell growth and invasion by targeting CHL1. *J Cell Biochem* 115(5):847–853. <https://doi.org/10.1002/jcb.24726>
 593. Ma D, Pan Z, Chang Q, Zhang JJ, Liu X, Hua N, Li GH (2021) KLF5-mediated Eppk1 expression promotes cell proliferation in cervical cancer via the p38 signaling pathway. *BMC Cancer* 21(1):377. <https://doi.org/10.1186/s12885-021-08040-y>
 594. Nie J, Shao J, Guo SW, Liu X (2021) The relevance of plasma R-spondin 1 and Slit2 as predictive biomarkers in cervical cancer chemotherapy and radiotherapy. *Ann Transl Med* 9(10):837. <https://doi.org/10.21037/atm-21-87>
 595. Bager P, Wohlfahrt J, Sørensen E, Ullum H, Høgdall CK, Palle C, Husemoen LL, Linneberg A, Kjaer SK, Melbye M et al (2015) Common flaggrin gene mutations and risk of cervical cancer. *Acta Oncol* 54(2):217–223. <https://doi.org/10.3109/0284186X.2014.973613>
 596. Zhang M, Song Y, Zhai F (2018) ARFHPV E7 oncogene, lncRNA HOTAIR, miR-331-3p and its target, NRP2, form a negative feedback loop to regulate the apoptosis in the tumorigenesis in HPV positive cervical cancer. *J Cell Biochem* 119(6):4397–4407. <https://doi.org/10.1002/jcb.26503>
 597. Akar S, Harmankaya İ, Uğraş S, Çelik Ç (2020) Expression and clinical significance of nicotinamide N-methyltransferase in cervical squamous cell carcinoma. *Int J Gynecol Pathol* 39(3):289–295. <https://doi.org/10.1097/PGP.0000000000000605>
 598. Qin Y, Zhao W (2022) The modeling analysis and effect of CHI3L1 and CD31-marked microvessel density in the occurrence and development of cervical squamous cell carcinoma. *Comput Math Methods Med* 2022:3516335. <https://doi.org/10.1155/2022/3516335>
 599. Zheng LL, Cai L, Zhang XQ, Lei Z, Yi CS, Liu XD, Yang JG (2022) Dysregulated RUNX1 predicts poor prognosis by mediating epithelial-mesenchymal transition in cervical cancer. *Curr Med Sci* 42(6):1285–1296. <https://doi.org/10.1007/s11596-022-2661-x>
 600. Wang Q, Wang B, Zhang W, Zhang T, Liu Q, Jiao X, Ye J, Hao Y, Gao Q, Ma G et al (2024) APLN promotes the proliferation, migration, and glycolysis of cervical cancer through the PI3K/AKT/mTOR pathway. *Arch Biochem Biophys* 755:109983. <https://doi.org/10.1016/j.abb.2024.109983>
 601. Liu R, Shuai Y, Luo J, Zhang Z (2019) SEMA3C promotes cervical cancer growth and is associated with poor prognosis. *Front Oncol* 9:1035. <https://doi.org/10.3389/fonc.2019.01035>
 602. Ou R, Lv M, Liu X, Lv J, Zhao J, Zhao Y, Li X, Li W, Zhao L, Li J et al (2020) HPV16 E6 oncoprotein-induced upregulation of lncRNA GABPB1-AS1 facilitates cervical cancer progression by regulating miR-519e-5p/Notch2 axis. *FASEB J* 34(10):13211–13223. <https://doi.org/10.1096/fj.202000762R>
 603. Wang X, Xu J, Hua F, Wang Y, Fang G, Zhang H, Wu X, Wang X, Xu J, Hua F, Wang Y, Fang G, Zhang H, Wu X (2023) MiR-214-3p suppresses cervical cancer cell metastasis by downregulating THBS2. *Cell Mol Biol (Noisy-le-grand)* 69(9):195–200. <https://doi.org/10.14715/cmb/2023.69.9.30>
 604. Chang PA, Sun YJ, Huang FF, Qin WZ, Chen YY, Zeng X, Wu YJ (2013) Identification of human patatin-like phospholipase domain-containing protein 1 and a mutant in human cervical cancer HeLa cells. *Mol Biol Rep* 40(10):5597–5605. <https://doi.org/10.1007/s11033-013-2661-9>
 605. Liu G, Du X, Xiao L, Zeng Q, Liu Q (2021) Activation of FGD5-AS1 promotes progression of cervical cancer through regulating BST2 to inhibit macrophage M1 polarization. *J Immunol Res* 2021:5857214. <https://doi.org/10.1155/2021/5857214>
 606. Wen D, Wang L, Tan S, Tang R, Xie W, Liu S, Tang C, He Y (2022) HOXD9 aggravates the development of cervical cancer by transcriptionally activating HMCN1. *Panminerva Med* 64(4):532–536. <https://doi.org/10.23736/S0031-0808.20.03911-7>
 607. Cho H, Chung JY, Kim S, Braunschweig T, Kang TH, Kim J, Chung EJ, Hewitt SM, Kim JH (2014) MICA/B and ULBP1 NKG2D ligands are independent predictors of good prognosis in cervical cancer. *BMC Cancer* 14:957. <https://doi.org/10.1186/1471-2407-14-957>
 608. Fullár A, Karázi K, Hollósi P, Lendvai G, Oláh L, Reszegi A, Papp Z, Sobel G, Dudás J, Kovalszky I (2020) Two ways of epigenetic silencing of TFPI2 in cervical cancer. *PLoS ONE* 15(6):e0234873. <https://doi.org/10.1371/journal.pone.0234873>
 609. Gasimli K, Raab M, Becker S, Sanhaji M, Strebhardt K (2022) The role of DAPK1 in the cell cycle regulation of cervical cancer cells and in response to topotecan. *J Cancer* 13(3):728–743. <https://doi.org/10.7150/jca.66492>
 610. Taniguchi-Ponciano K, Ribas-Aparicio RM, Marrero-Rodríguez D, Arreola-De la Cruz H, Huerta-Padilla V, Muñoz N, Gómez-Ortiz L, Ponce-Navarrete G, Rodríguez-Esquivel M, Mendoza-Rodríguez M, Gómez-Virgilio L et al (2018) The KISS1 gene overexpression as a potential molecular marker for cervical cancer cells. *Cancer Biomark* 22(4):709–719. <https://doi.org/10.3233/CBM-181215>
 611. Cao G, Zhang Z (2018) FPR1 mediates the tumorigenicity of human cervical cancer cells. *Cancer Manag Res* 10:5855–5865. <https://doi.org/10.2147/CMAR.S182795>
 612. Tong W, Zhang H (2021) Overexpression of long non-coding RNA WT1-AS or silencing of PIK3AP1 are inhibitory to cervical cancer progression. *Cell Cycle* 20(24):2583–2596. <https://doi.org/10.1080/15384101.2021.1991106>
 613. Wang EL, Zhang JJ, Luo FM, Fu MY, Li D, Peng J, Liu B (2023) Cerebellin-2 promotes endothelial-mesenchymal transition in hypoxic pulmonary hypertension rats by activating NF-κB/HIF-1α/Twist1 pathway. *Life Sci* 328:121879. <https://doi.org/10.1016/j.lfs.2023.121879>
 614. Oguri M, Kato K, Yokoi K, Yoshida T, Watanabe S, Metoki N, Yoshida H, Satoh K, Aoyagi Y, Nozawa Y et al (2010) Assessment of a polymorphism of SDK1 with hypertension in Japanese individuals. *Am J Hypertens* 23(1):70–77. <https://doi.org/10.1038/ajh.2009.190>
 615. Li L, Chu Y, Fink GD, Engelhardt JF, Heistad DD, Chen AF (2003) Endothelin-1 stimulates arterial VCAM-1 expression via NADPH oxidase-derived superoxide in mineralocorticoid hypertension. *Hypertension* 42(5):997–1003. <https://doi.org/10.1161/01.HYP.0000095980.43859.59>
 616. Fogelgren B, Yang S, Sharp IC, Huckstep OJ, Ma W, Somponpun SJ, Carlson EC, Uyehara CF, Lozanoff S (2009) Deficiency in Six2 during prenatal development is associated with reduced nephron number, chronic renal failure, and hypertension in Br/+ adult mice. *Am J Physiol Renal Physiol* 296(5):F1166–F1178. <https://doi.org/10.1152/ajprenal.90550.2008>

617. Zhang L, Liu J, Cheng P, Lv F (2019) Correlation between miRNA target site polymorphisms in the 3' UTR of AVPR1A and the risk of hypertension in the Chinese Han population. *Biosci Rep* 39(5):BSR20182232. <https://doi.org/10.1042/BSR20182232>
618. Kim M, Yoo HJ, Kim M, Kim J, Baek SH, Song M, Lee JH (2017) EPHA6 rs4857055 C > T polymorphism associates with hypertension through triglyceride and LDL particle size in the Korean population. *Lipids Health Dis* 16(1):230. <https://doi.org/10.1186/s12944-017-0620-5>
619. Li B, Yang H, Zhang W, Shi Y, Qin S, Wei Y, He Y, Yang W, Jiang S, Jin H (2018) Fatty acid-binding protein 4 predicts gestational hypertension and preeclampsia in women with gestational diabetes mellitus. *PLoS ONE* 13(2):e0192347. <https://doi.org/10.1371/journal.pone.0192347>
620. Toprak K, Yıldız Z, Akdemir S, Esen K, Kada R, Can Güleç N, Omar B, Biçer A, Demirbağ R (2023) Low pregnancy-specific beta-1-glycoprotein is associated with nondipper hypertension and increased risk of preeclampsia in pregnant women with newly diagnosed chronic hypertension. *Scand J Clin Lab Invest* 83(7):479–488. <https://doi.org/10.1080/00365513.2023.2275083>
621. Jimenez C, Hawn MB, Akin E, Leblanc N (2022) Translational potential of targeting anoctamin-1-encoded calcium-activated chloride channels in hypertension. *Biochem Pharmacol* 206:115320. <https://doi.org/10.1016/j.bcp.2022.115320>
622. Saleem M, Saavedra-Sánchez L, Barturen-Larrea P, Gomez JA (2021) The transcription factor Sox6 controls renin expression during renal artery stenosis. *Kidney360* 2(5):842–856. <https://doi.org/10.34067/KID.0002792020>
623. Shi L, Tian C, Sun L, Cao F, Meng Z (2018) The lncRNA TUG1/miR-145-5p/FGF10 regulates proliferation and migration in VSMCs of hypertension. *Biochem Biophys Res Commun* 501(3):688–695. <https://doi.org/10.1016/j.bbrc.2018.05.049>
624. Miwa Y, Kamide K, Takiuchi S, Yoshii M, Horio T, Tanaka C, Banno M, Miyata T, Kawano Y (2009) Association of PLA2G7 polymorphisms with carotid atherosclerosis in hypertensive Japanese. *Hypertens Res* 32(12):1112–1118. <https://doi.org/10.1038/hr.2009.151>
625. Soler A, Hunter I, Joseph G, Hutcheson R, Hutcheson B, Yang J, Zhang FF, Joshi SR, Bradford C, Gotlinger KH et al (2018) Elevated 20-HETE in metabolic syndrome regulates arterial stiffness and systolic hypertension via MMP12 activation. *J Mol Cell Cardiol* 117:88–99. <https://doi.org/10.1016/j.yjmcc.2018.02.005>
626. Du H, Xiao G, Xue Z, Li Z, He S, Du X, Zhou Z, Cao L, Wang Y, Yang J et al (2021) QiShenYiQi ameliorates salt-induced hypertensive nephropathy by balancing ADRA1D and SIK1 expression in Dahl salt-sensitive rats. *Biomed Pharmacother* 141:111941. <https://doi.org/10.1016/j.biopha.2021.111941>
627. Liang Y, Liu Y, Wang S, Gu Y, Wang P, Meng J (2022) Abnormal expression of the LAG-3/FGL-1 signaling pathway in patients with early-onset preeclampsia. *Med Sci Monit* 28:e937498. <https://doi.org/10.12659/MSM.937498>
628. Morris BJ, Chen R, Donlon TA, Kallianpur KJ, Masaki KH, Willcox BJ (2023) Vascular endothelial growth factor receptor 1 gene (FLT1) longevity variant increases lifespan by reducing mortality risk posed by hypertension. *Aging (Albany NY)* 15(10):3967–3983. <https://doi.org/10.18632/aging.204722>
629. Lei Q, Yu Z, Li H, Cheng J, Wang Y (2022) Fatty acid-binding protein 5 aggravates pulmonary artery fibrosis in pulmonary hypertension secondary to left heart disease via activating wnt/ β -catenin pathway. *J Adv Res* 40:197–206. <https://doi.org/10.1016/j.jare.2021.11.011>
630. Wu X, Zhang N, Yu J, Liang M, Xu H, Hu J, Lin S, Qiu J, Lin C et al (2023) The underlying mechanism of transcription factor IRF1, PRDM1, and ZNF263 involved in the regulation of NPPB rs3753581 on pulse pressure hypertension. *Gene* 878:147580. <https://doi.org/10.1016/j.gene.2023.147580>
631. Hu Y, Xia W, Li Y, Wang Q, Lin S, Wang B, Zhou C, Cui Y, Jiang Y, Pu X et al (2020) High-salt intake increases TRPC3 expression and enhances TRPC3-mediated calcium influx and systolic blood pressure in hypertensive patients. *Hypertens Res* 43(7):679–687. <https://doi.org/10.1038/s41440-020-0409-1>
632. Peng W, Xie Y, Xia J, Li B, Zhang F, Wen F, Liu K, Cao H, Qi H, Zhang L (2022) Association of circulating biomarkers of lnc-IGSF3-1:1, SCOC-AS1, and SLC8A1-AS1 with salt sensitivity of blood pressure in Chinese population. *J Cardiovasc Transl Res* 15(4):906–917. <https://doi.org/10.1007/s12265-021-10190-0>
633. Cai B, Du J (2021) Role of bone morphogenic protein-4 in gestational diabetes mellitus-related hypertension. *Exp Ther Med* 22(1):762. <https://doi.org/10.3892/etm.2021.10194>
634. Chen P, Zhang K, Zhou B, Zhang Z, Song Y, Pu Y, Yang Y, Zhang Y, Zhou R, Wang T et al (2014) The variations in the IL1RL1 gene and susceptibility to preeclampsia. *Immunol Invest* 43(5):424–435. <https://doi.org/10.3109/08820139.2013.879173>
635. Stern N, Eshkol A, Lunenfeld B, Rosenthal T (1983) Prolactin secretion in essential hypertension. *Clin Exp Hypertens A* 5(4):543–558. <https://doi.org/10.3109/10641968309081791>
636. Pajewski NM, Elahi FM, Tamura MK, Hinman JD, Nasrallah IM, Ix JH, Miller LM, Launer LJ, Wright CB, Supiano MA et al (2022) Plasma amyloid beta, neurofilament light chain, and total tau in the systolic blood pressure intervention trial (SPRINT). *Alzheimers Dement* 18(8):1472–1483. <https://doi.org/10.1002/alz.12496>
637. Kawarazaki W, Mizuno R, Nishimoto M, Ayuzawa N, Hirohama D, Ueda K, Kawakami-Mori F, Oba S, Marumo T, Fujita T (2020) Salt causes aging-associated hypertension via vascular Wnt5a under Klotho deficiency. *J Clin Invest* 130(8):4152–4166. <https://doi.org/10.1172/JCI134431>
638. Wang C, Xu H, Liao X, Wang W, Wu W, Li W, Niu L, Li Z, Li A, Sun Y et al (2023) Hypertension promotes the proliferation and migration of ccRCC cells by downregulation of TIMP3 in tumor endothelial cells through the miR-21-5p/TGFBR2/P38/EGR1 axis. *Mol Cancer Res* 21(1):62–75. <https://doi.org/10.1158/1541-7786.MCR-22-0089>
639. Kadoya M, Koyama H, Kanzaki A, Kurajoh M, Hatayama M, Shiraishi J, Okazaki H, Shoji T, Moriawaki Y, Yamamoto T et al (2014) Plasma brain-derived neurotrophic factor and reverse dipping pattern of nocturnal blood pressure in patients with cardiovascular risk factors. *PLoS ONE* 9(8):e105977. <https://doi.org/10.1371/journal.pone.0105977>
640. Li Q, Wong JH, Lu G, Antonio GE, Yeung DK, Ng TB, Forster LE, Yew DT (2009) Gene expression of synaptosomal-associated protein 25 (SNAP-25) in the prefrontal cortex of the spontaneously hypertensive rat (SHR). *Biochim Biophys Acta* 1792(8):766–776. <https://doi.org/10.1016/j.bbadis.2009.05.006>
641. Sammar M, Apicella C, Altevogt P, Meiri H, Vaiman D (2022) Modeling preeclampsia in vitro: polymorphic variants of STOX1-A/B genes can downregulate CD24 in trophoblast cell lines. *Int J Mol Sci* 23(24):15927. <https://doi.org/10.3390/ijms232415927>
642. Rossi E, Casali B, Regolisti G, Davoli S, Perazzoli F, Negro A, Sani C, Tumiatì B, Nicoli D (1998) Increased plasma levels of platelet-derived growth factor (PDGF-BB + PDGF-AB) in patients with never-treated mild essential hypertension. *Am J Hypertens* 11(10):1239–1243. [https://doi.org/10.1016/s0895-7061\(98\)00124-1](https://doi.org/10.1016/s0895-7061(98)00124-1)
643. Calvier L, Herz J, Hansmann G (2022) Interplay of low-density lipoprotein receptors, LRP6, and lipoproteins in pulmonary hypertension. *JACC Basic Transl Sci* 7(2):164–180. <https://doi.org/10.1016/j.jaccbts.2021.09.011>
644. Manner IW, Trøseid M, Oektedalen O, Baekken M, Os I (2013) Low nadir CD4 cell count predicts sustained hypertension in HIV-infected individuals. *J Clin Hypertens (Greenwich)* 15(2):101–106. <https://doi.org/10.1111/jch.12029>
645. Sullivan MN, Thakore P, Krishnan V, Alphonsa S, Li W, Feng Earley Y, Earley S (2023) Endothelial cell TRPA1 activity exacerbates cerebral hemorrhage during severe hypertension. *Front Mol Biosci* 10:1129435. <https://doi.org/10.3389/fmolb.2023.1129435>
646. Namli Kalem M, Kalem Z, Yüce T, Soylemez F (2018) ADAMTS 1, 4, 12, and 13 levels in maternal blood, cord blood, and placenta in preeclampsia. *Hypertens Pregnancy* 37(1):9–17. <https://doi.org/10.1080/10641955.2017.1397690>
647. Pan Z, Wu X, Zhang X, Hu K (2023) Phosphodiesterase 4B activation exacerbates pulmonary hypertension induced by intermittent hypoxia by regulating mitochondrial injury and cAMP/PKA/p-CREB/PGC-1 α signaling. *Biomed Pharmacother* 158:114095. <https://doi.org/10.1016/j.biopha.2022.114095>
648. Zhou JJ, Li H, Qian YL, Quan RL, Chen XX, Li L, Li Y, Wang PH, Meng XM, Jing XL et al (2020) Nestin represents a potential marker of pulmonary vascular remodeling in pulmonary arterial hypertension associated with congenital heart disease. *J Mol Cell Cardiol* 149:41–53. <https://doi.org/10.1016/j.yjmcc.2020.09.00>

649. Lee YH, Kim YG, Moon JY, Kim JS, Jeong KH, Lee TW, Ihm CG, Lee SH (2016) Genetic variations of tyrosine hydroxylase in the pathogenesis of hypertension. *Electrolyte Blood Press* 14(2):21–26. <https://doi.org/10.5049/EBP.2016.14.2.21>
650. Kandel M, MacDonald TM, Walker SP, Cluver C, Bergman L, Myers J, Hastie R, Keenan E, Hannan NJ, Cannon P et al (2022) PSG7 and 9 (pregnancy-specific β -1 glycoproteins 7 and 9): Novel biomarkers for preeclampsia. *J Am Heart Assoc* 11(7):e024536. <https://doi.org/10.1161/JAHA.121.024536>
651. Ortnr NJ (2023) CACNA1D-related channelopathies: from hypertension to autism. *Handb Exp Pharmacol* 279:183–225. https://doi.org/10.1007/164_2022_626
652. Maghajothe S, Subramanian L, Mani P, Singh M, Iyer DR, Sharma S, Khullar M, Victor SM, Asthana S, Mulasari AS et al (2022) A common Matrix metalloproteinase 8 promoter haplotype enhances the risk for hypertension via diminished interactions with nuclear factor kappa B. *J Hypertens* 40(11):2147–2160. <https://doi.org/10.1097/HJH.00000000000003234>
653. Gjesing AP, Andersen G, Burgdorf KS, Borch-Johnsen K, Jørgensen T, Hansen T, Pedersen O (2007) Studies of the associations between functional beta2-adrenergic receptor variants and obesity, hypertension and type 2 diabetes in 7,808 white subjects. *Diabetologia* 50(3):563–568. <https://doi.org/10.1007/s00125-006-0578-8>
654. Kanbay M, Demiray A, Afsar B, Covic A, Tapoi L, Ureche C, Ortiz A (2021) Role of klotho in the development of essential hypertension. *Hypertension* 77(3):740–750. <https://doi.org/10.1161/HYPERTENSIONAHA.120.16635>
655. Levi M, Moons L, Bouché A, Shapiro SD, Collen D, Carmeliet P (2001) Deficiency of urokinase-type plasminogen activator-mediated plasmin generation impairs vascular remodeling during hypoxia-induced pulmonary hypertension in mice. *Circulation* 103(15):2014–2020. <https://doi.org/10.1161/01.cir.103.15.2014>
656. Jin X, Narisawa M, Piao L, Cheng XW (2022) Protein tyrosine phosphatase receptor type D as a potential therapeutic target in pulmonary artery hypertension. *J Hypertens* 40(9):1650–1654. <https://doi.org/10.1097/HJH.00000000000003232>
657. Samara TD, Liem IK, Prijanti AR (2019) SEMA3B but Not CUL1 as marker for pre-eclampsia progression. *Malays J Med Sci* 26(1):66–72. <https://doi.org/10.21315/mjms2019.26.1.6>
658. Dzieza-Grudnik A, Siga O, Walczewska J, Wizner B, Wolkow PP, Messerli FH, Grodzicki T (2023) Urocortin 2 - a protective effect in hypertension? *J Physiol Pharmacol*. <https://doi.org/10.26402/jpp.2023.1.01>
659. Ongun MC, Tonyali NV, Kaplan O, Deger I, Celebier M, Basci Akduman NE, Sahin D, Yucel A, Babaoglu MO (2023) Effects of genetic polymorphisms of CYP2J2, CYP2C9, CYP2C19, CYP4F2, CYP4F3 and CYP4A11 enzymes in preeclampsia and gestational hypertension. *Placenta* 137:88–95. <https://doi.org/10.1016/j.placenta.2023.04.019>
660. Park HY, Kim JH, Bae S, Choi YY, Park JY, Hong YC (2015) Interaction effect of serum 25-hydroxyvitamin D levels and CYP1A1, CYP1B1 polymorphisms on blood pressure in an elderly population. *J Hypertens* 33(1):69–76. <https://doi.org/10.1097/HJH.0000000000000381>
661. Montani D, Girerd B, Günther S, Riant F, Tournier-Lasserre E, Magy L, Maazi N, Guignabert C, Savale L, Sitbon O et al (2014) Pulmonary arterial hypertension in familial hemiplegic migraine with ATP1A2 channelopathy. *Eur Respir J* 43(2):641–643. <https://doi.org/10.1183/09031936.001470>
662. Zhao A, Qi Y, Liu K (2020) CLDN3 expression and function in pregnancy-induced hypertension. *Exp Ther Med* 20(4):3798–3806. <https://doi.org/10.3892/etm.2020.9084>
663. Fernández-Solà J, Borrissier-Pairó F, Antúnez E, Tobías E (2015) Myostatin and insulin-like growth factor-1 in hypertensive heart disease: a prospective study in human heart donors. *J Hypertens* 33(4):851–859. <https://doi.org/10.1097/HJH.0000000000000493>
664. Nowzari Z, Masoumi M, Nazari-Robati M, Akbari H, Shahrokhii N, Asadikaram G (2018) Association of polymorphisms of leptin, leptin receptor and apelin receptor genes with susceptibility to coronary artery disease and hypertension. *Life Sci* 207:166–171. <https://doi.org/10.1016/j.lfs.2018.06.007>
665. Hilton LR, Rätsep MT, VandenBroek MM, Jafri S, Lavery KJ, Mitchell M, Theilmann AL, Smart JA, Hawke LG, Moore SD et al (2022) Impaired interleukin-15 signaling via BMP2R loss drives natural killer cell deficiency and pulmonary hypertension. *Hypertension* 79(11):2493–2504. <https://doi.org/10.1161/HYPERTENSIONAHA.122.19178>
666. Scholl UI, Stölting G, Nelson-Williams C, Vichot AA, Choi M, Loring E, Prasad ML, Goh G, Carling T, Juhlin CC et al (2015) Recurrent gain of function mutation in calcium channel CACNA1H causes early-onset hypertension with primary aldosteronism. *Elife* 4:e06315. <https://doi.org/10.7554/eLife.06315>
667. Anderson L, Lowery JW, Frank DB, Novitskaya T, Jones M, Mortlock DP, Chandler RL, de Caestecker MP (2010) Bmp2 and Bmp4 exert opposing effects in hypoxic pulmonary hypertension. *Am J Physiol Regul Integr Comp Physiol* 298(3):R833–R842. <https://doi.org/10.1152/ajpregu.00534.2009>
668. Zhang X, Li Q, Jiang W, Xiong X, Li H, Zhao J, Qi H (2020) LAMA5 promotes human umbilical vein endothelial cells migration, proliferation, and angiogenesis and is decreased in preeclampsia. *J Matern Fetal Neonatal Med* 33(7):1114–1124. <https://doi.org/10.1080/14767058.2018.1514597>
669. Yamada Y, Kato K, Yoshida T, Yokoi K, Matsuo H, Watanabe S, Ichihara S, Metoki N, Yoshida H, Satoh K et al (2008) Association of polymorphisms of ABCA1 and ROS1 with hypertension in Japanese individuals. *Int J Mol Med* 21(1):83–89
670. Shi J, Liu Y, Liu Y, Li Y, Qiu S, Bai Y, Gu Y, Luo J, Cui H, Li Y et al (2018) Association between ApoE polymorphism and hypertension: a meta-analysis of 28 studies including 5898 cases and 7518 controls. *Gene* 675:197–207. <https://doi.org/10.1016/j.gene.2018.06.097>
671. Udjus C, Cero FT, Halvorsen B, Behmen D, Carlson CR, Bendiksen BA, Espe EKS, Sjaastad I, Løberg EM, Yndestad A et al (2019) Caspase-1 induces smooth muscle cell growth in hypoxia-induced pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol* 316(6):L999–L1012. <https://doi.org/10.1152/ajplung.00322.2018>
672. Kolb TM, Johnston L, Damarla M, Kass DA, Hassoun PM (2021) PDE9A deficiency does not prevent chronic-hypoxic pulmonary hypertension in mice. *Physiol Rep* 9(18):e15057. <https://doi.org/10.14814/phy2.15057>
673. Crnkovic S, Rittchen S, Jandl K, Gindlhuber J, Zabini D, Mutgan AC, Valzano F, Boehm PM, Hoetzenecker K, Toller W et al (2023) Divergent roles of Ephrin-B2/EphB4 guidance system in pulmonary hypertension. *Hypertension* 80(2):e17–e28. <https://doi.org/10.1161/HYPERTENSIONAHA.122.19479>
674. Malfará BN, Benzi JRL, de Oliveira Filgueira GC, Zanelli CF, Duarte G, de Carvalho CR, de Moraes NV (2019) ABCG2 c.421C>A polymorphism alters nifedipine transport to breast milk in hypertensive breastfeeding women. *Reprod Toxicol* 85:1–5. <https://doi.org/10.1016/j.reprotox.2019.01.007>
675. Ishigami T, Iwamoto T, Tamura K, Yamaguchi S, Iwasawa K, Uchino K, Umemura S, Ishii M (1995) Angiotensin I converting enzyme (ACE) gene polymorphism and essential hypertension in Japan. Ethnic difference of ACE genotype. *Am J Hypertens* 8(1):95–97. [https://doi.org/10.1016/0895-7061\(94\)00184-D](https://doi.org/10.1016/0895-7061(94)00184-D)
676. Barberis MC, Veronese S, Bauer D, De Juli E, Harari S (1995) Immunocytochemical detection of progesterone receptors. A study in a patient with primary pulmonary hypertension. *Chest* 107(3):869–872. <https://doi.org/10.1378/chest.107.3.869>
677. Seo JY, Choi JH (2021) Genetic variations in thiamin transferase SLC35F3 and the risk of hypertension in Koreans. *Clin Nutr Res* 10(2):140–149. <https://doi.org/10.7762/cnr.2021.10.2.140>
678. Ariff A, Melton PE, Brennecke SP, Moses EK (2019) Analysis of the epigenome in multiplex pre-eclampsia families identifies SORD, DGKI, and ICA1 as novel candidate risk genes. *Front Genet* 10:227. <https://doi.org/10.3389/fgene.2019.00227>
679. Rajagambeeram R, Abu Raghavan S, Ghosh S, Basu S, Ramasamy R, Murugaiyan SB (2014) Diagnostic utility of heat stable alkaline phosphatase in hypertensive disorders of pregnancy. *J Clin Diagn Res* 8(11):CC10–CC13. <https://doi.org/10.7860/JCDR/2014/10895.5084>
680. Jain PP, Lai N, Xiong M, Chen J, Babicheva A, Zhao T, Parmisano S, Zhao M, Paquin C, Matti M et al (2021) TRPC6, a therapeutic target for pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol* 321(6):L1161–L1182. <https://doi.org/10.1152/ajplung.00159.2021>
681. Fiorucci S, Distrutti E (2016) Targeting the transsulfuration-H2S pathway by FXR and GPBAR1 ligands in the treatment of portal hypertension. *Pharmacol Res* 111:749–756. <https://doi.org/10.1016/j.phrs.2016.07.040>

682. Scheiner B, Mandorfer M, Schwabl P, Payer BA, Bucsecs T, Bota S, Aichelburg MC, Grabmeier-Pfistershammer K, Stättermayer A, Ferenci P et al (2015) The Impact of PNPLA3 rs738409 SNP on liver fibrosis progression, portal hypertension and hepatic steatosis in HIV/HCV coinfection. *PLoS ONE* 10(11):e0143429. <https://doi.org/10.1371/journal.pone.0143429>
683. Dunk CE, Bucher M, Zhang J, Hayder H, Geraghty DE, Lye SJ, Myatt L, Hackmon R (2022) Human leukocyte antigen HLA-C, HLA-G, HLA-F, and HLA-E placental profiles are altered in early severe preeclampsia and preterm birth with chorioamnionitis. *Am J Obstet Gynecol* 227(4):641.e1–641.e13. <https://doi.org/10.1016/j.ajog.2022.07.021>
684. Laggner M, Hacker P, Oberndorfer F, Bauer J, Raunegger T, Gerges C, Szerafin T, Thanner J, Lang I, Skoro-Sajer N et al (2022) The roles of S100A4 and the EGF/EGFR signaling axis in pulmonary hypertension with right ventricular hypertrophy. *Biology (Basel)* 11(1):118. <https://doi.org/10.3390/biology11010118>
685. Zhou X, Liang B, Lin W, Zha L (2024) Identification of MACC1 as a potential biomarker for pulmonary arterial hypertension based on bioinformatics and machine learning. *Comput Biol Med.* <https://doi.org/10.1016/j.compbmed.2024.108372>
686. Zou X, Wu Z, Huang J, Liu P, Qin X, Chen L, Zhu W, Zhao Y, Li P, Song J et al (2018) The role of matrix metalloproteinase-3 in the doxycycline attenuation of intracranial venous hypertension-induced angiogenesis. *Neurosurgery* 83(6):1317–1327. <https://doi.org/10.1093/neuros/nyx633>
687. Yamagata K, Tagami M, Ikeda K, Tsumagari S, Yamori Y, Nara Y (2002) Differential regulation of glial cell line-derived neurotrophic factor (GDNF) mRNA expression during hypoxia and reoxygenation in astrocytes isolated from stroke-prone spontaneously hypertensive rats. *Glia* 37(1):1–7. <https://doi.org/10.1002/glia.10003>
688. Jiao K, Su P, Li Y (2023) FGFR2 modulates the Akt/Nrf2/ARE signaling pathway to improve angiotensin II-induced hypertension-related endothelial dysfunction. *Clin Exp Hypertens* 45(1):2208777. <https://doi.org/10.1080/10641963.2023.2208777>
689. Wang X, Shields C, Tardo G, Peacock G, Hester E, Anderson M, Williams JM, Cornelius DC (2024) IL-33 supplementation improves uterine artery resistance and maternal hypertension in response to placental ischemia. *Am J Physiol Heart Circ Physiol* 326(4):H1006–H1016. <https://doi.org/10.1152/ajpheart.00045.2024>
690. Tomoda F, Nitta A, Sugimori H, Koike T, Kinugawa K (2022) Plasma and urinary levels of nerve growth factor are elevated in primary hypertension. *Int J Hypertens* 2022:3003269. <https://doi.org/10.1155/2022/3003269>
691. Wang Y, Jia H, Gao WH, Zou T, Yao S, Du MF, Zhang XY, Chu C, Liao YY, Chen C et al (2021) Associations of plasma PAPP-A2 and genetic variations with salt sensitivity, blood pressure changes and hypertension incidence in Chinese adults. *J Hypertens* 39(9):1817–1825. <https://doi.org/10.1097/HJH.0000000000002846>
692. Wang H, Yuan Z, Wang B, Li B, Lv H, He J, Huang Y, Cui Z, Ma Q, Li T et al (2022) COMP (cartilage oligomeric matrix protein), a novel PIEZO1 regulator that controls blood pressure. *Hypertension* 79(3):549–561. <https://doi.org/10.1161/HYPERTENSIONAHA.121.179>
693. Toyama T, Kudryashova TV, Ichihara A, Lenna S, Looney A, Shen Y, Jiang L, Teos L, Avolio T, Lin D et al (2023) GATA6 coordinates cross-talk between BMP10 and oxidative stress axis in pulmonary arterial hypertension. *Sci Rep* 13(1):6593. <https://doi.org/10.1038/s41598-023-33779-8>
694. van der Have O, Mead TJ, Westöö C, Peruzzi N, Mutgan AC, Norvik C, Bech M, Struglics A, Hoetzenecker K, Brunström H et al (2023) Aggrecan accumulates at sites of increased pulmonary arterial pressure in idiopathic pulmonary arterial hypertension. *Pulm Circ* 13(1):e12200. <https://doi.org/10.1002/pul2.12200>
695. Fujimaki T, Oguri M, Horibe H, Kato K, Matsuoka R, Abe S, Tokoro F, Arai M, Noda T, Watanabe S et al (2015) Association of a transcription factor 21 gene polymorphism with hypertension. *Biomed Rep* 3(1):118–122. <https://doi.org/10.3892/br.2014.371>
696. Li R, Xie J, Xu W, Zhang L, Lin H, Huang W (2022) LPS-induced PTGS2 manipulates the inflammatory response through trophoblast invasion in preeclampsia via NF- κ B pathway. *Reprod Biol* 22(4):100696. <https://doi.org/10.1016/j.repbio.2022.100696>
697. Lozano-Velasco E, Wangenstein R, Quesada A, Garcia-Padilla C, Osorio JA, Ruiz-Torres MD, Aranega A, Franco D (2017) Hyperthyroidism, but not hypertension, impairs PITX2 expression leading to Wnt-microRNA-ion channel remodeling. *PLoS ONE* 12(12):e0188473. <https://doi.org/10.1371/journal.pone.0188473>
698. Sun CJ, Li L, Li X, Zhang WY, Liu XW (2018) Novel SNPs of WNK1 and AKR1C3 are associated with preeclampsia. *Gene* 668:27–32. <https://doi.org/10.1016/j.gene.2018.05.055>
699. Schäfer K, Tello K, Pak O, Richter M, Gierhardt M, Kwapiszewska G, Veith C, Fink L, Gall H, Hecker M et al (2023) Decreased plasma levels of the brain-derived neurotrophic factor correlate with right heart congestion in pulmonary arterial hypertension. *ERJ Open Res* 9(2):00230–02022. <https://doi.org/10.1183/23120541.00230-2022>
700. Sierra-Ramos C, Velazquez-Garcia S, Vastola-Mascolo A, Hernández G, Faresse N, de la Rosa DA (2020) SGK1 activation exacerbates diet-induced obesity, metabolic syndrome and hypertension. *J Endocrinol* 244(1):149–162
701. Bhagwani AR, Ali M, Piper B, Liu M, Hudson J, Kelly N, Bogamuwa S, Yang H, Londino JD, Bednash JS et al (2023) A p53-TLR3 axis ameliorates pulmonary hypertension by inducing BMPR2 via IRF3. *iScience* 26(2):105935. <https://doi.org/10.1016/j.isci.2023.105935>
702. Abu-Farha M, Cherian P, Qaddoumi MG, AlKhairi I, Sriraman D, Alanbaei M, Abubaker J (2018) Increased plasma and adipose tissue levels of ANGPTL8/Betatrophin and ANGPTL4 in people with hypertension. *Lipids Health Dis* 17(1):35. <https://doi.org/10.1186/s12944-018-0681-0>
703. Agalakova NI, Reznik VA, Ershov IA, Lupanova EA, Nadei OV, Ivanov DO, David Adair C, Bagrov AY (2022) Silencing of Fli1 gene mimics effects of preeclampsia and induces collagen synthesis in human umbilical arteries. *Am J Hypertens* 35(9):828–832. <https://doi.org/10.1093/ajh/hpac065>
704. Wei ZX, Cai XX, Fei YD, Wang Q, Hu XL, Li C, Hou JW, Yang YL, Wang YP, Li YG (2024) Ntsr1 contributes to pulmonary hypertension by enhancing endoplasmic reticulum stress via JAK2-STAT3-Thbs1 signaling. *Transl Res.* <https://doi.org/10.1016/j.trsl.2024.02.002>
705. Avecilla V (2019) Effect of transcriptional regulator ID3 on pulmonary arterial hypertension and hereditary hemorrhagic telangiectasia. *Int J Vasc Med* 2019:2123906. <https://doi.org/10.1155/2019/2123906>
706. Pavlov TS, Palygin O, Isaeva E, Levchenko V, Khedr S, Blass G, Ilatovskaya DV, Cowley AW Jr, Staruschenko A (2020) NOX4-dependent regulation of ENaC in hypertension and diabetic kidney disease. *FASEB J* 34(10):13396–13408. <https://doi.org/10.1096/fj.202000966RR>
707. Gu Q, Yazdanpanah M, van Hoek M, Hofman A, Gao X, de Rooij FW, Sijbrands EJ (2015) Common variants in PCSK1 influence blood pressure and body mass index. *J Hum Hypertens* 29(2):82–86. <https://doi.org/10.1038/jhh.2014.59>
708. Palumbo V, Segat L, Padovan L, Amoroso A, Trimarco B, Izzo R, Lembo G, Regitz-Zagrosek V, Knoll R, Brancaccio M et al (2009) Melusin gene (ITGB1BP2) nucleotide variations study in hypertensive and cardiopathic patients. *BMC Med Genet* 10:140. <https://doi.org/10.1186/1471-2350-10-140>
709. Guo XG, Ding J, Xu H, Xuan TM, Jin WQ, Yin X, Shang YP, Zhang FR, Zhu JH, Zheng LR (2014) Comprehensive assessment of the association of WNK4 polymorphisms with hypertension: evidence from a meta-analysis. *Sci Rep* 4:6507. <https://doi.org/10.1038/srep06507>
710. Zadora J, Singh M, Herse F, Przybyl L, Haase N, Golic M, Yung HW, Huppertz B, Cartwright JE, Whitley G et al (2017) Disturbed placental imprinting in preeclampsia leads to altered expression of DLX5, a human-specific early trophoblast marker. *Circulation* 136(19):1824–1839. <https://doi.org/10.1161/CIRCULATIONAHA.117.028110>
711. Caccamo D, Cannata A, Ricca S, Catalano LM, Montalto AF, Alibrandi A, Ercoli A, Granese R (2020) Role of vitamin-D receptor (VDR) single nucleotide polymorphisms in gestational hypertension development: a case-control study. *PLoS ONE* 15(11):e0239407. <https://doi.org/10.1371/journal.pone.0239407>
712. Zhang Z, Tremblay J, Raelson J, Sofer T, Du L, Fang Q, Argos M, Marois-Blanchet FC, Wang Y, Yan L et al (2019) EPHA4 regulates vascular smooth muscle cell contractility and is a sex-specific hypertension risk gene in individuals with type 2 diabetes. *J Hypertens* 37(4):775–789. <https://doi.org/10.1097/HJH.0000000000001948>
713. Chirinos JA, Sardana M, Syed AA, Koppula MR, Varakantam S, Vasim I, Oldland HG, Phan TS, Drummen NEA, Vermeer C et al (2018) Aldosterone, inactive matrix gla-protein, and large artery stiffness in

- hypertension. *J Am Soc Hypertens* 12(9):681–689. <https://doi.org/10.1016/j.jash.2018.06.018>
714. Bao Q, Wang D, Zhang Y, Bao L, Jia H (2020) The impact of CYP24A1 polymorphisms on hypertension susceptibility. *Kidney Blood Press Res* 45(1):28–37. <https://doi.org/10.1159/000503925>
715. Grätze P, Dechend R, Stocker C, Park JK, Feldt S, Shagdarsuren E, Wellner M, Gueler F, Rong S, Gross V et al (2008) Novel role for inhibitor of differentiation 2 in the genesis of angiotensin II-induced hypertension. *Circulation* 117(20):2645–2656. <https://doi.org/10.1161/CIRCULATIONAHA.107.760116>
716. Holmes L Jr, Lim A, Comeaux CR, Dabney KW, Okundaye O (2019) DNA methylation of candidate genes (ACE II, IFN- γ , AGTR 1, CKG, ADD1, SCNN1B and TLR2) in essential hypertension: a systematic review and quantitative evidence synthesis. *Int J Environ Res Public Health* 16(23):4829. <https://doi.org/10.3390/ijerph16234829>
717. Manhiani MM, Seth DM, Banes-Berceli AK, Satou R, Navar LG, Brands MW (2015) The role of IL-6 in the physiologic versus hypertensive blood pressure actions of angiotensin II. *Physiol Rep* 3(10):e12595. <https://doi.org/10.14814/phy2.12595>
718. Seidelmann SB, Vardeny O, Claggett B, Yu B, Shah AM, Ballantyne CM, Selvin E, MacRae CA, Boerwinkle E, Solomon SD (2017) An NPPB promoter polymorphism associated with elevated N-terminal pro-B-type natriuretic peptide and lower blood pressure, hypertension, and mortality. *J Am Heart Assoc* 6(4):e005257. <https://doi.org/10.1161/JAHA.116.005257>
719. Van Beusecum JP, Barbaro NR, Smart CD, Patrick DM, Loperena R, Zhao S, de la Visitacion N, Ao M, Xiao L, Shibao CA et al (2021) Growth arrest specific-6 and Axl coordinate inflammation and hypertension. *Circ Res* 129(11):975–991. <https://doi.org/10.1161/CIRCRESAHA.121.319643>
720. Ong KL, Leung RY, Babinska A, Salifu MO, Ehrlich YH, Kornecki E, Wong LY, Tso AW, Cherny SS, Sham PC et al (2009) Elevated plasma level of soluble F11 receptor/junctional adhesion molecule-A (F11R/JAM-A) in hypertension. *Am J Hypertens* 22(5):500–505. <https://doi.org/10.1038/ajh.2009.2>
721. Quintero-Ronderos P, Jiménez KM, Esteban-Pérez C, Ojeda DA, Bello S, Fonseca DJ, Coronel MA, Moreno-Ortiz H, Sierra-Díaz DC, Lucena E et al (2019) FOXD1 mutations are related to repeated implantation failure, intra-uterine growth restriction and preeclampsia. *Mol Med* 25(1):37. <https://doi.org/10.1186/s10020-019-0104-3>
722. Wang X, Chow FL, Oka T, Hao L, Lopez-Campistrous A, Kelly S, Cooper S, Odenbach J, Finegan BA, Schulz R et al (2009) Matrix metalloproteinase-7 and ADAM-12 (a disintegrin and metalloproteinase-12) define a signaling axis in agonist-induced hypertension and cardiac hypertrophy. *Circulation* 119(18):2480–2489. <https://doi.org/10.1161/CIRCULATIONAHA.108.835488>
723. Arnett DK, Meyers KJ, Devereux RB, Tiwari HK, Gu CC, Vaughan LK, Perry RT, Patki A, Claas SA, Sun YV et al (2011) Genetic variation in NCAM1 contributes to left ventricular wall thickness in hypertensive families. *Circ Res* 108(3):279–283. <https://doi.org/10.1161/CIRCRESAHA.110.239210>
724. Jin HS, Hong KW, Lim JE, Hwang SY, Lee SH, Shin C, Park HK, Oh B (2010) Genetic variations in the sodium balance-regulating genes ENaC, NEDD4L, NDFIP2 and USP2 influence blood pressure and hypertension. *Kidney Blood Press Res* 33(1):15–23. <https://doi.org/10.1159/000275706>
725. Sindi HA, Russomanno G, Satta S, Abdul-Salam VB, Jo KB, Qazi-Chaudhry B, Ainscough AJ, Szulcek R, Jan Bogaard H, Morgan CC et al (2020) Therapeutic potential of KLF2-induced exosomal microRNAs in pulmonary hypertension. *Nat Commun* 11(1):1185. <https://doi.org/10.1038/s41467-020-14966-x>
726. Pu J, Wang F, Ye P, Jiang X, Zhou W, Gu Y, Chen S (2022) Salt-inducible kinase 1 deficiency promotes vascular remodeling in pulmonary arterial hypertension via enhancement of yes-associated protein-mediated proliferation. *Heliyon* 8(10):e11016. <https://doi.org/10.1016/j.heliyon.2022.e11016>
727. Zhou C, Chen Y, Kang W, Lv H, Fang Z, Yan F, Li L, Zhang W, Shi J (2019) Mir-455-3p-1 represses FGF7 expression to inhibit pulmonary arterial hypertension through inhibiting the RAS/ERK signaling pathway. *J Mol Cell Cardiol* 130:23–35. <https://doi.org/10.1016/j.yjmcc.2019.03.002>
728. Neto ABL, Vasconcelos NBR, Dos Santos TR, Duarte LEC, Assunção ML, de Sales-Marques C, Ferreira HDS (2021) Prevalence of IGFBP3, NOS3 and TCF7L2 polymorphisms and their association with hypertension: a population-based study with Brazilian women of African descent. *BMC Res Notes* 14(1):186. <https://doi.org/10.1186/s13104-021-05598-5>
729. Sardo MA, Mandraffino G, Riggio S, D'Ascola A, Alibrandi A, Saitta C, Imbalzano E, Castaldo M, Cinquegrani M, Saitta A (2010) Effects of the angiotensin II receptor blocker losartan on the monocyte expression of biglycan in hypertensive patients. *Clin Exp Pharmacol Physiol* 37(9):933–938. <https://doi.org/10.1111/j.1440-1681.2010.05407.x>
730. Su L, Li X, Mao X, Xu T, Zhang Y, Li S, Zhu X, Wang L, Yao D, Wang J et al (2023) Circ-Ntrk2 acts as a miR-296-5p sponge to activate the TGF- β 1/p38 MAPK pathway and promote pulmonary hypertension and vascular remodelling. *Respir Res* 24(1):78. <https://doi.org/10.1186/s12931-023-02385-7>
731. Fedorowicz A, Mateuszuk Ł, Kopec G, Skórka T, Kutryb-Zając B, Zakrzewska A, Walczak M, Jakubowski A, Lomnicka M, Słomińska E et al (2016) Activation of the nicotinamide N-methyltransferase (NNMT)-1-methyl-nicotinamide (MNA) pathway in pulmonary hypertension. *Respir Res* 17(1):108. <https://doi.org/10.1186/s12931-016-0423-7>
732. Sun X, Nakajima E, Norbrun C, Sorkhdini P, Yang AX, Yang D, Ventetuolo CE, Braza J, Vang A, Aliotta J et al (2022) Chitinase 3 like 1 contributes to the development of pulmonary vascular remodeling in pulmonary hypertension. *JCI Insight* 7(18):e159578. <https://doi.org/10.1172/jci.insight.159578>
733. Simmons Beck R, Liang OD, Klinger JR (2023) Light at the Endothelium: role of Sox17 and Runx1 in endothelial dysfunction and pulmonary arterial hypertension. *Front Cardiovasc Med* 10:1274033. <https://doi.org/10.3389/fcvm.2023.1274033>
734. Gandham R, Dayanand CD, Sheela SR, Kiranmayee P (2022) Maternal serum Apelin 13 and APLN gene promoter variant -1860T > C in preeclampsia. *J Matern Fetal Neonatal Med* 35(25):5008–5016. <https://doi.org/10.1080/14767058.2021.1874341>
735. Oudejans CB, Poutsma A, Michel OJ, Thulluru HK, Mulders J, van de Vrugt HJ, Sistermans EA, van Dijk M (2016) Noncoding RNA-regulated gain-of-function of STOX2 in Finnish pre-eclamptic families. *Sci Rep* 6:32129. <https://doi.org/10.1038/srep32129>
736. Carr G, Barrese V, Stott JB, Povstyan OV, Jepps TA, Figueiredo HB, Zheng D, Jamshidi Y, Greenwood IA (2016) MicroRNA-153 targeting of KCNQ4 contributes to vascular dysfunction in hypertension. *Cardiovasc Res* 112(2):581–589. <https://doi.org/10.1093/cvr/cvw177>
737. Sahoo S, Li Y, de Jesus D, Sembrat J, Rojas MM, Goncharova E, Cifuentes-Pagano E, Straub AC, Pagano PJ (2021) Notch2 suppression mimicking changes in human pulmonary hypertension modulates Notch1 and promotes endothelial cell proliferation. *Am J Physiol Heart Circ Physiol* 321(3):H542–H557. <https://doi.org/10.1152/ajpheart.00125.2021>
738. Spradley FT, Tan AY, Joo WS, Daniels G, Kussie P, Karumanchi SA, Granger JP (2016) Placental growth factor administration abolishes placental ischemia-induced hypertension. *Hypertension* 67(4):740–747. <https://doi.org/10.1161/HYPERTENSIONAHA.115.06783>
739. Ye Y, Li M, Chen L, Li S, Quan Z (2021) Circ-AK2 is associated with preeclampsia and regulates biological behaviors of trophoblast cells through miR-454-3p/THBS2. *Placenta* 103:156–163. <https://doi.org/10.1016/j.placenta.2020.10.023>
740. Owusu D, Pan Y, Xie C, Harirforoosh S, Wang KS (2017) Polymorphisms in PDLIM5 gene are associated with alcohol dependence, type 2 diabetes, and hypertension. *J Psychiatr Res* 84:27–34. <https://doi.org/10.1016/j.jpsychires.2016.09.015>
741. Gunawardhana KL, Hong L, Rugira T, Uebbing S, Kucharczak J, Mehta S, Karunamuni DR, Cabera-Mendoza B, Gandotra N, Scharfe C et al (2023) A systems biology approach identifies the role of dysregulated PRDM6 in the development of hypertension. *J Clin Invest* 133(4):e160036. <https://doi.org/10.1172/JCI160036>
742. Chechekhin VI, Ivanova AM, Kulebyakin KY, Antropova YG, Karagayur MN, Skryabina MN, Chechekhina ES, Basalova NA, Grigorova OA, Syssoeva VY et al (2024) Peripheral 5-HT/HTR6 axis is responsible for obesity-associated hypertension. *Biochim Biophys Acta Mol Cell Res* 1871(2):119651. <https://doi.org/10.1016/j.bbamcr.2023.119651>
743. Adão R, Mendes-Ferreira P, Maia-Rocha C, Santos-Ribeiro D, Rodrigues PG, Vidal-Meireles A, Monteiro-Pinto C, Pimentel LD, Falcão-Pires I, De Keulenaer GW et al (2019) Neuregulin-1 attenuates right ventricular diastolic stiffness in experimental pulmonary hypertension. *Clin Exp Pharmacol Physiol* 46(3):255–265. <https://doi.org/10.1111/1440-1681.13043>

744. Tomaszewski M, Grywalska E, Topyła-Putowska W, Błaszczak P, Kurzyńska M, Roliński J, Kopeć G (2021) High CD200 expression on T CD4+ and T CD8+ lymphocytes as a non-invasive marker of idiopathic pulmonary hypertension-preliminary study. *J Clin Med* 10(5):950. <https://doi.org/10.3390/jcm10050950>
745. Jinyu L, Shuying W, Panchan Z, Dan C, Chao C, Xingyu Y, Weiwei C (2022) Bone marrow stromal cell antigen 2(BST2) suppresses the migration and invasion of trophoblasts in preeclampsia by downregulating matrix metalloproteinase 2(MMP2). *Bioengineered* 13(5):13174–13187. <https://doi.org/10.1080/21655979.2022.2074712>
746. Koot BG, Alders M, Verheij J, Beuers U, Cobben JM (2016) A de novo mutation in KCNN3 associated with autosomal dominant idiopathic non-cirrhotic portal hypertension. *J Hepatol* 64(4):974–977. <https://doi.org/10.1016/j.jhep.2015.11.027>
747. Le MT, Lobmeyer MT, Campbell M, Cheng J, Wang Z, Turner ST, Chapman AB, Boerwinkle E, Gums JG, Gong Y et al (2013) Impact of genetic polymorphisms of SLC2A2, SLC2A5, and KHK on metabolic phenotypes in hypertensive individuals. *PLoS ONE* 8(1):e52062. <https://doi.org/10.1371/journal.pone.0052062>
748. Hu Z, Dong C, Dong Q (2021) Circ_0015382 is associated with preeclampsia and regulates biological behaviors of trophoblast cells through miR-149-5p/TFPI2 axis. *Placenta* 108:73–80. <https://doi.org/10.1016/j.placenta.2021.03.005>
749. Lang CT, Markham KB, Behrendt NJ, Suarez AA, Samuels P, Vandre DD, Robinson JM, Ackerman WE 4th (2009) Placental dysferlin expression is reduced in severe preeclampsia. *Placenta* 30(8):711–718. <https://doi.org/10.1016/j.placenta.2009.05.008>
750. Sasagawa S, Nishimura Y, Sawada H, Zhang E, Okabe S, Murakami S, Ashikawa Y, Yuge M, Kawaguchi K, Kawase R et al (2016) Comparative transcriptome analysis identifies CCDC80 as a novel gene associated with pulmonary arterial hypertension. *Front Pharmacol* 7:142. <https://doi.org/10.3389/fphar.2016.00142>
751. Yung C, MacDonald TM, Walker SP, Cannon P, Harper A, Pritchard N, Hannan NJ, Kaitu'u-Lino TJ, Tong S (2019) Death associated protein kinase 1 (DAPK-1) is increased in preeclampsia. *Placenta* 88:1–7. <https://doi.org/10.1016/j.placenta.2019.09.010>
752. Pinilla L, Castellano JM, Romero M, Tena-Sempere M, Gaytán F, Aguilar E (2009) Delayed puberty in spontaneously hypertensive rats involves a primary ovarian failure independent of the hypothalamic KiSS-1/GPR54/GnRH system. *Endocrinology* 150(6):2889–2897. <https://doi.org/10.1210/en.2008-1381>
753. Yang HC, Liang YJ, Chen JW, Chiang KM, Chung CM, Ho HY, Ting CT, Lin TH, Sheu SH, Tsai WC et al (2012) Identification of IGF1, SLC4A4, WWOX, and SFMBT1 as hypertension susceptibility genes in Han Chinese with a genome-wide gene-based association study. *PLoS ONE* 7(3):e32907. <https://doi.org/10.1371/journal.pone.0032907>
754. Batool M, Berghausen EM, Zierden M, Vantler M, Schermuly RT, Baldus S, Rosenkranz S, Ten Freyhaus H (2020) The six-transmembrane protein Stamp2 ameliorates pulmonary vascular remodeling and pulmonary hypertension in mice. *Basic Res Cardiol* 115(6):68. <https://doi.org/10.1007/s00395-020-00826-8>
755. Chang TJ, Wang WC, Hsiung CA, He CT, Lin MW, Sheu WH, Chang YC, Quertermous T, Chen I, Rotter J et al (2016) Genetic variation in the human SORBS1 gene is associated with blood pressure regulation and age at onset of hypertension: a SAPHIRE cohort study. *Medicine (Baltimore)* 95(10):e2970. <https://doi.org/10.1097/MD.0000000000002970>
756. Yan S, Cui S, Zhang L, Yang B, Yuan Y, Lv X, Fu H, Li Y, Huang C, Wang P (2020) Expression of ACKR2 in placentas from different types of preeclampsia. *Placenta* 90:121–127. <https://doi.org/10.1016/j.placenta.2019.12.01>
757. Edwards JM, Roy S, Galla SL, Tomcho JC, Bearss NR, Waigi EW, Mell B, Cheng X, Saha P, Vijay-Kumar M et al (2021) FPR-1 (Formyl peptide receptor-1) activation promotes spontaneous, premature hypertension in Dahl salt-sensitive rats. *Hypertension* 77(4):1191–1202. <https://doi.org/10.1161/HYPERTENSIONAHA.120.16237>
758. Nakano M, Koga M, Hashimoto T, Matsushita N, Masukawa D, Mizuno Y, Uchimura H, Niikura R, Miyazaki T, Nakamura F et al (2022) Right ventricular overloading is attenuated in monocrotaline-induced pulmonary hypertension model rats with a disrupted Gpr143 gene, the gene that encodes the 3,4-l-dihydroxyphenylalanine (l-DOPA) receptor. *J Pharmacol Sci* 148(2):214–220. <https://doi.org/10.1016/j.jphs.2021.11.008>
759. García SI, Porto PI, Dieuzeide G, Landa MS, Kirsznér T, Plotquin Y, Gonzalez C, Pirola CJ (2001) Thyrotropin-releasing hormone receptor (TRHR) gene is associated with essential hypertension. *Hypertension* 38(3 Pt 2):683–687. <https://doi.org/10.1161/01.hyp.38.3.683>
760. Wang X, Singh P, Zhou L, Sharafeldin N, Landier W, Hageman L, Burridge P, Yasui Y, Sapkota Y, Blanco JG et al (2023) Genome-wide association study identifies ROBO2 as a novel susceptibility gene for anthracycline-related cardiomyopathy in childhood cancer survivors. *J Clin Oncol* 41(9):1758–1769. <https://doi.org/10.1200/JCO.2022.01527>
761. Singh V, Kaur R, Kumari P, Pasricha C, Singh R (2023) ICAM-1 and VCAM-1: gatekeepers in various inflammatory and cardiovascular disorders. *Clin Chim Acta* 548:117487. <https://doi.org/10.1016/j.cca.2023.117487>
762. Ueland T, Gullestad L, Kou L, Aukrust P, Anand IS, Broughton MN, McMurray JJ, van Veldhuisen DJ, Warren DJ, Bolstad N (2018) Pro-gastrin-releasing peptide and outcome in patients with heart failure and anaemia: results from the RED-HF study. *ESC Heart Fail* 5(6):1052–1059. <https://doi.org/10.1002/ehf2.12312>
763. Saito N, Furuhashi M, Koyama M, Higashiura Y, Akasaka H, Tanaka M, Moniwa N, Ohnishi H, Saitoh S, Ura N et al (2021) Elevated circulating FABP4 concentration predicts cardiovascular death in a general population: a 12-year prospective study. *Sci Rep* 11(1):4008. <https://doi.org/10.1038/s41598-021-83494-5>
764. Tian X, Sun C, Wang X, Ma K, Chang Y, Guo Z, Si J (2020) ANO1 regulates cardiac fibrosis via ATI-mediated MAPK pathway. *Cell Calcium* 92:102306. <https://doi.org/10.1016/j.ceca.2020.102306>
765. Saleem M, Rahman S, Eljovich F, Laffer CL, Ertuglu LA, Masenga SK, Kirabo A (2022) Sox6, a potential target for MicroRNAs in cardiometabolic disease. *Curr Hypertens Rep* 24(5):145–156. <https://doi.org/10.1007/s11906-022-01175-8>
766. Zeng M, Wei X, He YL, Chen JX, Lin WT (2023) TFAP2C inhibits cell autophagy to alleviate myocardial ischemia/reperfusion injury by regulating miR-23a-5p/SFRP5/Wnt5a axis. *FASEB J* 37(6):e22959. <https://doi.org/10.1096/fj.202201962R>
767. Barrick CJ, Lenhart PM, Dackor RT, Nagle E, Caron KM (2012) Loss of receptor activity-modifying protein 3 exacerbates cardiac hypertrophy and transition to heart failure in a sex-dependent manner. *J Mol Cell Cardiol* 52(1):165–174. <https://doi.org/10.1016/j.yjmcc.2011.10.021>
768. Li Y, Jiang Y, Zhang Y, Li N, Yin Q, Liu L, Lv X, Liu Y, Li A, Fang B et al (2021) Abnormal upregulation of cardiovascular disease biomarker PLA2G7 induced by proinflammatory macrophages in COVID-19 patients. *Sci Rep* 11(1):6811. <https://doi.org/10.1038/s41598-021-85848-5>
769. Marcos-Jubilar M, Orbe J, Roncal C, Machado FJD, Rodriguez JA, Fernández-Montero A, Colina I, Rodil R, Pastrana JC, Páramo JA (2021) Association of SDF1 and MMP12 with atherosclerosis and inflammation: clinical and experimental study. *Life (Basel)* 11(5):414. <https://doi.org/10.3390/life11050414>
770. Corella D, Sorlí JV, González JJ, Ortega C, Fitó M, Bulló M, Martínez-González MA, Ros E, Arós F, Lapetra J et al (2014) Novel association of the obesity risk-allele near fas apoptotic inhibitory molecule 2 (FAIM2) gene with heart rate and study of its effects on myocardial infarction in diabetic participants of the PREDIMED trial. *Cardiovasc Diabetol* 13:5. <https://doi.org/10.1186/1475-2840-13-5>
771. Annema W, Gawinecka J, Muendlein A, Saely CH, Drexel H, von Eckardstein A (2022) Elevated levels of apolipoprotein D predict poor outcome in patients with suspected or established coronary artery disease. *Atherosclerosis* 341:27–33. <https://doi.org/10.1016/j.atherosclerosis.2021.12.011>
772. Jimenez J, Prabhu SD (2022) LAG3 blockade expands T cells within atherosclerotic plaque: an ongoing need for cardiovascular disease risk assessment. *JACC CardioOncol* 4(5):646–648. <https://doi.org/10.1016/j.jacc.2022.11.002>
773. Shi HY, Xie MS, Yang CX, Huang RT, Xue S, Liu XY, Xu YJ, Yang YQ (2022) Identification of SOX18 as a new gene predisposing to congenital heart disease. *Diagnostics (Basel)* 12(8):1917. <https://doi.org/10.3390/diagn12081917>
774. Wu Z, Geng J, Bai Y, Qi Y, Chang C, Jiao Y, Guo Z (2023) miR-125b-5p alleviates the damage of myocardial infarction by inhibiting the NFAT2 to reduce F2RL2 expression. *Regen Med* 18(7):543–559. <https://doi.org/10.2217/rme-2022-0150>

775. Chen C, Peng H, Zeng Y, Dong G (2020) CD14, CD163, and CCR1 are involved in heart and blood communication in ischemic cardiac diseases. *J Int Med Res* 48(9):300060520951649. <https://doi.org/10.1177/0300060520951649>
776. Mauricio R, Singh K, Sanghavi M, Ayers CR, Rohatgi A, Vongpatanasin W, de Lemos JA, Khera A (2022) Soluble Fms-like tyrosine kinase-1 (sFlt-1) is associated with subclinical and clinical atherosclerotic cardiovascular disease: the Dallas heart study. *Atherosclerosis* 346:46–52. <https://doi.org/10.1016/j.atherosclerosis.2022.02.026>
777. Ma T, Lin S, Wang B, Wang Q, Xia W, Zhang H, Cui Y, He C, Wu H, Sun F et al (2019) TRPC3 deficiency attenuates high salt-induced cardiac hypertrophy by alleviating cardiac mitochondrial dysfunction. *Biochem Biophys Res Commun* 519(4):674–681. <https://doi.org/10.1016/j.bbrc.2019.09.018>
778. Li Y, Wang DW, Chen Y, Chen C, Guo J, Zhang S, Sun Z, Ding H, Yao Y, Zhou L et al (2018) Genome-wide association and functional studies identify SCML4 and THSD7A as novel susceptibility genes for coronary artery disease. *Arterioscler Thromb Vasc Biol* 38(4):964–975. <https://doi.org/10.1161/ATVBAHA.117.310594>
779. Li XX, Mu B, Li X, Bie ZD (2022) circCELF1 inhibits myocardial fibrosis by regulating the expression of DKK2 through FTO/m6A and miR-636. *J Cardiovasc Transl Res* 15(5):998–1009. <https://doi.org/10.1007/s12265-022-10209-0>
780. Li Q, Park K, Li C, Rask-Madsen C, Mima A, Qi W, Mizutani K, Huang P, King GL (2013) Induction of vascular insulin resistance and endothelin-1 expression and acceleration of atherosclerosis by the overexpression of protein kinase C- β isoform in the endothelium. *Circ Res* 113(4):418–427. <https://doi.org/10.1161/CIRCRESAHA.113.301074>
781. Han F, Chen Q, Su J, Zheng A, Chen K, Sun S, Wu H, Jiang L, Xu X, Yang M et al (2019) MicroRNA-124 regulates cardiomyocyte apoptosis and myocardial infarction through targeting Dhcr24. *J Mol Cell Cardiol* 132:178–188. <https://doi.org/10.1016/j.yjmcc.2019.05.007>
782. Polidovitch N, Yang S, Sun H, Lakin R, Ahmad F, Gao X, Turnbull PC, Chiarello C, Perry CGR, Manganiello V et al (2019) Phosphodiesterase type 3A (PDE3A), but not type 3B (PDE3B), contributes to the adverse cardiac remodeling induced by pressure overload. *J Mol Cell Cardiol* 132:60–70. <https://doi.org/10.1016/j.yjmcc.2019.04.028>
783. Mu W, Qian S, Song Y, Yang L, Song S, Yang Q, Liu H, Liu Y, Pan D, Tang Y et al (2021) BMP4-mediated browning of perivascular adipose tissue governs an anti-inflammatory program and prevents atherosclerosis. *Redox Biol* 43:101979. <https://doi.org/10.1016/j.redox.2021.101979>
784. Lin JF, Wu S, Juang JJ, Chiang FT, Hsu LA, Teng MS, Cheng ST, Huang HL, Sun YC, Liu PY et al (2017) IL1RL1 single nucleotide polymorphism predicts sST2 level and mortality in coronary and peripheral artery disease. *Atherosclerosis* 257:71–77. <https://doi.org/10.1016/j.atherosclerosis.2016.12.020>
785. Wang T, Wang X, Teng Y, Wu L, Zhu F, Ma D, Wang H, Liu X (2024) APLAID complicated with arrhythmogenic dilated cardiomyopathy caused by a novel PLCG2 variant. *Immunol Res*. <https://doi.org/10.1007/s12026-024-09455-y>
786. Meyer T, Ruppert V, Ackermann S, Richter A, Perrot A, Sperling SR, Posch MG, Maisch B, Pankuweit S (2013) Novel mutations in the sarcomeric protein myopalladin in patients with dilated cardiomyopathy. *Eur J Hum Genet* 21(3):294–300. <https://doi.org/10.1038/ejhg.2012.173>
787. Glezer A, Santana MR, Bronstein MD, Donato J Jr, Jallad RS (2023) The interplay between prolactin and cardiovascular disease. *Front Endocrinol (Lausanne)* 13:1018090. <https://doi.org/10.3389/fendo.2022.1018090>
788. Tong S, Du Y, Ji Q, Dong R, Cao J, Wang Z, Li W, Zeng M, Chen H et al (2020) Expression of Sfrp5/Wnt5a in human epicardial adipose tissue and their relationship with coronary artery disease. *Life Sci* 245:117338. <https://doi.org/10.1016/j.lfs.2020.117338>
789. Schumacher D, Peisker F, Kramann R (2021) MEOX1: a novel drug-gable target that orchestrates the activation of fibroblasts in cardiac fibrosis. *Signal Transduct Target Ther* 6(1):440. <https://doi.org/10.1038/s41392-021-00842-7>
790. Chen H, Chen S, Ye H, Guo X (2022) Protective effects of circulating TIMP3 on coronary artery disease and myocardial infarction: a mendelian randomization study. *J Cardiovasc Dev Dis* 9(8):277. <https://doi.org/10.3390/jcdd9080277>
791. Guan H, Zhang J, Luan J, Xu H, Huang Z, Yu Q, Gou X, Xu L (2021) Secreted frizzled related proteins in cardiovascular and metabolic diseases. *Front Endocrinol (Lausanne)* 12:712217. <https://doi.org/10.3389/fendo.2021.712217>
792. Jia EZ, Wang J, Yang ZJ, Zhu TB, Wang LS, Wang H, Li CJ, Chen B, Cao KJ, Huang J et al (2009) Association of the mutation for the human carboxypeptidase E gene exon 4 with the severity of coronary artery atherosclerosis. *Mol Biol Rep* 36(2):245–254. <https://doi.org/10.1007/s11033-007-9173-4>
793. Wei M, Pan H, Guo K (2021) Association between plasma ADAMTS-9 levels and severity of coronary artery disease. *Angiology* 72(4):371–380. <https://doi.org/10.1177/0003319720979238>
794. Yang K, Song HF, He S, Yin WJ, Fan XM, Ru F, Gong H, Zhai XY, Zhang J, Peng ZX et al (2019) Effect of neuron-derived neurotrophic factor on rejuvenation of human adipose-derived stem cells for cardiac repair after myocardial infarction. *J Cell Mol Med* 23(9):5981–5993. <https://doi.org/10.1111/jcmm.14456>
795. Tuuminen R, Dashkevich A, Keränen MA, Raissadati A, Krebs R, Jokinen JJ, Arnaudova R, Rouvinen E, Ylä-Herttuala S, Nykänen AI et al (2016) Platelet-derived growth factor-B protects rat cardiac allografts from ischemia-reperfusion injury. *Transplantation* 100(2):303–313. <https://doi.org/10.1097/TP.0000000000000909>
796. Liu Q, Dong Y, Escames G, Wu X, Ren J, Yang W, Zhang S, Zhu Y, Tian Y, Acuña-Castroviejo D et al (2022) Identification of PIK3CG as a hub in septic myocardial injury using network pharmacology and weighted gene co-expression network analysis. *Bioeng Transl Med* 8(1):e10384. <https://doi.org/10.1002/btm2.10384>
797. Ference BA, Kastelein JJP, Ray KK, Ginsberg HN, Chapman MJ, Packard CJ, Laufs U, Oliver-Williams C, Wood AM, Butterworth AS et al (2019) Association of triglyceride-lowering LPL variants and LDL-C-lowering LDLR variants with risk of coronary heart disease. *JAMA* 321(4):364–373. <https://doi.org/10.1001/jama.2018.20045>
798. Saigusa R, Roy P, Freuchet A, Gulati R, Ghosheh Y, Suthahar SSA, Durant CP, Hanna DB, Kiosses WB, Orecchioni M et al (2022) Single cell transcriptomics and TCR reconstruction reveal CD4 T cell response to MHC-II-restricted APOB epitope in human cardiovascular disease. *Nat Cardiovasc Res* 1(5):462–475. <https://doi.org/10.1038/s44161-022-00063-3>
799. Li R, Liu R, Yan F, Zhuang X, Shi H, Gao X (2020) Inhibition of TRPA1 promotes cardiac repair in mice after myocardial infarction. *J Cardiovasc Pharmacol* 75(3):240–249. <https://doi.org/10.1097/FJC.00000000000000783>
800. Zhao X, Zhu L, Yin Q, Xu Z, Jia Q, Yang R, He K (2022) F2RL3 methylation in the peripheral blood as a potential marker for the detection of coronary heart disease: a case-control study. *Front Genet* 13:833923. <https://doi.org/10.3389/fgene.2022.833923>
801. Guan H, Shi T, Liu M, Wang X, Guo F (2022) C1QL1/CTRP14 is largely dispensable for atherosclerosis formation in apolipoprotein-E-deficient mice. *J Cardiovasc Dev Dis* 9(10):341. <https://doi.org/10.3390/jcdd9100341>
802. Wang Z, Ye D, Ye J, Wang M, Liu J, Jiang H, Xu Y, Zhang J, Chen J, Wan J (2019) ADAMTS-5 decreases in coronary arteries and plasma from patients with coronary artery disease. *Dis Markers* 2019:6129748. <https://doi.org/10.1155/2019/6129748>
803. Wan Q, Xu C, Zhu L, Zhang Y, Peng Z, Chen H, Rao H, Zhang E, Wang H, Chu F et al (2022) Targeting PDE4B (phosphodiesterase-4 subtype B) for cardioprotection in acute myocardial infarction via neutrophils and microcirculation. *Circ Res* 131(5):442–455. <https://doi.org/10.1161/CIRCRESAHA.122.321365>
804. Del Toro R, Chèvre R, Rodríguez C, Ordóñez A, Martínez-González J, Andrés V, Méndez-Ferrer S et al (2016) Nestin(+) cells direct inflammatory cell migration in atherosclerosis. *Nat Commun* 7:12706. <https://doi.org/10.1038/ncomms12706>
805. Penttilä O, Merikallio E, Pispala J, Klinge E, Siltanen P, Kyösola K (1978) Auricular tyrosine hydroxylase and dopamine-beta-hydroxylase activities and noradrenaline content in ischaemic heart disease. *Acta Med Scand* 203(3):161–166. <https://doi.org/10.1111/j.0954-6820.1978.tb14850.x>
806. Salminen A, Vlachopoulou E, Havulinna AS, Tervahartiala T, Sattler W, Lokki ML, Nieminen MS, Perola M, Salomaa V, Sinisalo J et al (2017) Genetic variants contributing to circulating matrix metalloproteinase 8 levels and their association with cardiovascular diseases: a

- genome-wide analysis. *Circ Cardiovasc Genet* 10(6):e001731. <https://doi.org/10.1161/CIRCGENETICS.117.001731>
807. Wang Y, Zheng Y, Zhang W, Yu H, Lou K, Zhang Y, Qin Q, Zhao B, Yang Y, Hui R (2007) Polymorphisms of KDR gene are associated with coronary heart disease. *J Am Coll Cardiol* 50(8):760–767. <https://doi.org/10.1016/j.jacc.2007.04.074>
808. Onrat ST, Dural İE, Yalim Z, Onrat E (2022) Correction to: Investigating changes in β -adrenergic gene expression (ADRB1 and ADRB2) in Takotsubo (stress) cardiomyopathy syndrome; a pilot study. *Mol Biol Rep* 49(4):3373–3375. <https://doi.org/10.1007/s11033-022-07258-8>
809. Duval V, Alayrac P, Silvestre JS, Levoye A (2022) Emerging roles of the atypical chemokine receptor 3 (ACKR3) in cardiovascular diseases. *Front Endocrinol (Lausanne)* 13:906586. <https://doi.org/10.3389/fendo.2022.906586>
810. Thude H, Gerlach K, Richartz B, Krack A, Brenke B, Pethig K, Figulla HR, Barz D (2005) No association between transmembrane protein-tyrosine phosphatase receptor type C (CD45) exon a point mutation (77C>G) and idiopathic dilated cardiomyopathy. *Hum Immunol* 66(9):1008–1012. <https://doi.org/10.1016/j.humimm.2005.07.004>
811. Martín-Núñez E, Pérez-Castro A, Tagua VG, Hernández-Carballo C, Ferri C, Pérez-Delgado N, Rodríguez-Ramos S, Cerro-López P, López-Castillo Á, Delgado-Molinós A et al (2022) Klotho expression in peripheral blood circulating cells is associated with vascular and systemic inflammation in atherosclerotic vascular disease. *Sci Rep* 12(1):8422. <https://doi.org/10.1038/s41598-022-12548-z>
812. Xu JP, Zeng RX, He MH, Lin SS, Guo LH, Zhang MZ (2022) Associations between serum soluble α -klotho and the prevalence of specific cardiovascular disease. *Front Cardiovasc Med* 9:899307. <https://doi.org/10.3389/fcvm.2022.899307>
813. Gupta KK, Donahue DL, Sandoval-Cooper MJ, Castellino FJ, Ploplis VA (2017) Plasminogen activator inhibitor-1 protects mice against cardiac fibrosis by inhibiting urokinase-type plasminogen activator-mediated plasminogen activation. *Sci Rep* 7(1):365. <https://doi.org/10.1038/s41598-017-00418-y>
814. Dai R, Yang X, He W, Su Q, Deng X, Li J (2023) LncRNA AC005332.7 inhibited ferroptosis to alleviate acute myocardial infarction through regulating miR-331-3p/CCND2 Axis. *Korean Circ J* 53(3):151–167. <https://doi.org/10.4070/kcj.2022.0242>
815. Lics N, Krivmane B, Latkovskis G, Erglis A (2011) A common promoter variant of the gene encoding cyclooxygenase-1 (PTGS1) is related to decreased incidence of myocardial infarction in patients with coronary artery disease. *Thromb Res* 127(6):600–602. <https://doi.org/10.1016/j.thromres.2010.12.019>
816. Liu X, Li Y, Wang L, Zhao Q, Lu X, Huang J, Fan Z, Gu D (2008) The INSIG1 gene, not the INSIG2 gene, associated with coronary heart disease: tag-SNPs and haplotype-based association study. The Beijing Atherosclerosis Study. *Thromb Haemostasis* 100(5):886–892
817. Wang K, Zhou M, Zhang Y, Du Y, Li P, Guan C, Huang Z (2023) IRX2 activated by jumonji domain-containing protein 2A is crucial for cardiac hypertrophy and dysfunction in response to the hypertrophic stimuli. *Int J Cardiol* 371:332–344. <https://doi.org/10.1016/j.ijcard.2022.09.070>
818. Clancy RM, Halushka M, Rasmussen SE, Lhakhang T, Chang M, Buyon JP (2019) Siglec-1 macrophages and the contribution of IFN to the development of autoimmune congenital heart block. *J Immunol* 202(1):48–55. <https://doi.org/10.1049/jimmunol.1800357>
819. Adão R, Santos-Ribeiro D, Rademaker MT, Leite-Moreira AF, Brás-Silva C (2015) Urocortin 2 in cardiovascular health and disease. *Drug Discov Today* 20(7):906–914. <https://doi.org/10.1016/j.drudis.2015.02.012>
820. Tantray JA, Reddy KP, Jamil K, Yerra SK (2019) Role of cytochrome epoxygenase (CYP2J2) in the pathophysiology of coronary artery disease in South Indian population. *Indian Heart J* 71(1):60–64. <https://doi.org/10.1016/j.ihj.2018.11.011>
821. Peng DD, Xie W, Yu ZX (2017) Impact of interaction between CYP1A1 genetic polymorphisms and smoking on coronary artery disease in the Han of China. *Clin Exp Hypertens* 39(4):339–343. <https://doi.org/10.1080/10641963.2016.1259326>
822. Burt O, Johnston KJA, Graham N, Cullen B, Lyall DM, Lyall LM, Pell JP, Ward J, Smith DJ, Strawbridge RJ (2021) Genetic variation in the ASTN2 locus in cardiovascular, metabolic and psychiatric traits: evidence for pleiotropy rather than shared biology. *Genes (Basel)* 12(8):1194. <https://doi.org/10.3390/genes12081194>
823. Schlegel M, Moore KJ (2020) A heritable netrin-1 mutation increases atherogenic immune responses. *Atherosclerosis* 301:82–83. <https://doi.org/10.1016/j.atherosclerosis.2020.04.003>
824. Lee C, Li X (2018) Platelet-derived growth factor-C and -D in the cardiovascular system and diseases. *Mol Aspects Med* 62:12–21. <https://doi.org/10.1016/j.mam.2017.09.005>
825. Nosalski R, Guzik TJ (2021) IL-15 and IL-7: keys to dysregulated inflammation in acute coronary syndromes. *Cardiovasc Res* 117(8):1806–1808. <https://doi.org/10.1093/cvr/cvab189>
826. Wang MX, Liu X, Li JM, Liu L, Lu W, Chen GC (2020) Inhibition of CACNA1H can alleviate endoplasmic reticulum stress and reduce myocardial cell apoptosis caused by myocardial infarction. *Eur Rev Med Pharmacol Sci* 24(24):12887–12895. https://doi.org/10.26355/eurev_202012_24192
827. Bobos D, Soufla G, Angouras DC, Lekakis I, Georgopoulos S, Melissari E (2023) Investigation of the role of BMP2 and -4 in ASD, VSD and complex congenital heart disease. *Diagnostics (Basel)* 13(16):2717. <https://doi.org/10.3390/diagnostics13162717>
828. Sun T, Han Y, Li JL, Wang S, Jing ZJ, Yan Z, Zhou L, Zuo L, Yang JL, Cao JM (2024) Synaptotagmin-7 mediates cardiac hypertrophy by targeting autophagy. *FEBS J* 291(3):489–509. <https://doi.org/10.1111/febs.16961>
829. Fu Y, Jia Q, Ren M, Bie H, Zhang X, Zhang Q, He S, Li C, Zhou H, Wang Y et al (2023) Circular RNA ZBTB46 depletion alleviates the progression of Atherosclerosis by regulating the ubiquitination and degradation of hnRNP2B1 via the AKT/mTOR pathway. *Immun Ageing* 20(1):66. <https://doi.org/10.1186/s12979-023-00386-0>
830. Theodoraki EV, Nikopensius T, Suhorutsenko J, Papamikos V, Kolovou GD, Peppas V, Panagiotakos D, Limberi S, Zakopoulos N, Metspalu A et al (2009) ROS1 Asp2213Asn polymorphism is not associated with coronary artery disease in a Greek case-control study. *Clin Chem Lab Med* 47(12):1471–1473. <https://doi.org/10.1515/CCLM.2009.155>
831. Marais AD (2019) Apolipoprotein E in lipoprotein metabolism, health and cardiovascular disease. *Pathology* 51(2):165–176. <https://doi.org/10.1016/j.pathol.2018.11.002>
832. Park HS, Kim IJ, Kim EG, Ryu CS, Lee JY, Ko EJ, Park HW, Sung JH, Kim NK (2020) A study of associations between CLUBN, HNF1A, and LIPC gene polymorphisms and coronary artery disease. *Sci Rep* 10(1):16294. <https://doi.org/10.1038/s41598-020-73048-6>
833. Zhang Y, Wang C, Sun M, Jin Y, Braz CU, Khatib H, Hacker TA, Liss M, Gotthardt M, Granzier H et al (2022) RBM20 phosphorylation and its role in nucleocytoplasmic transport and cardiac pathogenesis. *FASEB J* 36(5):e22302. <https://doi.org/10.1096/fj.202101811RR>
834. Zheng F, Gong Z, Xing S, Xing Q (2014) Overexpression of caspase-1 in aorta of patients with coronary atherosclerosis. *Heart Lung Circ* 23(11):1070–1074. <https://doi.org/10.1016/j.hlc.2014.04.256>
835. Besler C, Rommel KP, Kresoja KP, Mörbitz J, Kirsten H, Scholz M, Klingel K, Thiery J, Burkhardt R, Büttner P et al (2021) Evaluation of phosphodiesterase 9A as a novel biomarker in heart failure with preserved ejection fraction. *ESC Heart Fail* 8(3):1861–1872. <https://doi.org/10.1002/ehf2.13327>
836. Liu J, Yang W, Li Y, Wei Z, Dan X (2020) ABCG2 rs2231142 variant in hyperuricemia is modified by SLC2A9 and SLC22A12 polymorphisms and cardiovascular risk factors in an elderly community-dwelling population. *BMC Med Genet* 21(1):54. <https://doi.org/10.1186/s12881-020-0987-4>
837. Li H, Lewis A, Brodsky S, Rieger R, Iden C, Goligorsky MS (2002) Homocysteine induces 3-hydroxy-3-methylglutaryl coenzyme a reductase in vascular endothelial cells: a mechanism for development of atherosclerosis? *Circulation* 105(9):1037–1043. <https://doi.org/10.1161/hc0902.104713>
838. Ferrari R (2005) Angiotensin-converting enzyme inhibition in cardiovascular disease: evidence with perindopril. *Expert Rev Cardiovasc Ther* 3(1):15–29. <https://doi.org/10.1586/14779072.3.1.15>
839. Sanders LN, Schoenhard JA, Saleh MA, Mukherjee A, Ryzhov S, McMaster WG Jr, Nolan K, Gumina RJ, Thompson TB, Magnuson MA et al (2016) BMP antagonist gremlin 2 limits inflammation after myocardial infarction. *Circ Res* 119(3):434–449. <https://doi.org/10.1161/CIRCRESAHA.116.308700>
840. Thériault S, Gaudreault N, Lamontagne M, Rosa M, Boulanger MC, Messika-Zeitoun D, Clavel MA, Capoulade R, Dagenais F et al (2018) A transcriptome-wide association study identifies PALMD as a

- susceptibility gene for calcific aortic valve stenosis. *Nat Commun* 9(1):988. <https://doi.org/10.1038/s41467-018-03260-6>
841. Lin YJ, Liu X, Chang JS, Chien WK, Chen JH, Tsang H, Hung CH, Lin TH, Huang SM, Liao CC et al (2014) Coronary artery aneurysms occurrence risk analysis between Kawasaki disease and LRP1B gene in Taiwanese children. *Biomedicine (Taipei)* 4(2):10. <https://doi.org/10.7603/s40681-014-0010-5>
842. Ren Y, Li X, Wang S, Pan W, Lv H, Wang M, Zhou X, Xia Y, Yin D (2021) Serum alkaline phosphatase levels are associated with coronary artery calcification patterns and plaque vulnerability. *Catheter Cardiovasc Interv* 97(Suppl 2):1055–1062. <https://doi.org/10.1002/ccd.29642>
843. Federspiel JM, Gartner J, Lipp P, Schmidt P, Tschernig T (2023) Elderly with varying extents of cardiac disease show interindividual fluctuating myocardial TRPC6-immunoreactivity. *J Cardiovasc Dev Dis* 10(1):26. <https://doi.org/10.3390/jcdd10010026>
844. Biagioli M, Marchianò S, Di Giorgio C, Bordoni M, Urbani G, Bellini R, Massa C, Sami Ullah Khan R, Roselli R, Chiara Monti M et al (2023) Activation of GPBAR1 attenuates vascular inflammation and atherosclerosis in a mouse model of NAFLD-related cardiovascular disease. *Biochem Pharmacol* 218:115900. <https://doi.org/10.1016/j.bcp.2023.115900>
845. Ochoa JP, Lalaguna L, Mirelis JG, Dominguez F, Gonzalez-Lopez E, Salas C, Roustan G, McGurk KA, Zheng SL, Barton PJR et al (2024) Biallelic loss of function variants in myocardial zonula adherens protein gene (MYZAP) cause a severe recessive form of dilated cardiomyopathy. *Circ Heart Fail* 17(3):e011226. <https://doi.org/10.1161/CIRCHEARTFAILURE.123.011226>
846. Moreira JBN, Wohlwend M, Fenk S, Åmellem I, Flatberg A, Kraljevic J, Marinovic J, Ljubkovic M, Bjørkøy G, Wisløff U (2019) Exercise reveals proline dehydrogenase as a potential target in heart failure. *Prog Cardiovasc Dis* 62(2):193–202. <https://doi.org/10.1016/j.pcad.2019.03.002>
847. Chen X, Wang X, Zhang Z, Chen Y, Wang C (2021) Role of IL-9, IL-2RA, and IL-2RB genetic polymorphisms in coronary heart disease. Bedeutung der genetischen polymorphismen IL-9, IL-2RA und IL-2RB bei koronarer Herzkrankheit. *Herz* 46(6):558–566. <https://doi.org/10.1007/s00059-020-05004-z>
848. Alghamdi MA, Al-Eitan L, Alkhatib R, Al-Assi A, Almasri A, Aljamal H, Aman H, Khasawneh R (2021) Variants in CDHR3, CACNAC1, and LTA genes predisposing sensitivity and response to warfarin in patients with cardiovascular disease. *Int J Gen Med* 14:1093–1100. <https://doi.org/10.2147/IJGM.S298597>
849. Akuta N, Kawamura Y, Arase Y, Saitoh S, Fujiyama S, Sezaki H, Hosaka T, Kobayashi M, Kobayashi M, Suzuki Y et al (2021) PNPLA3 genotype and fibrosis-4 index predict cardiovascular diseases of Japanese patients with histopathologically-confirmed NAFLD. *BMC Gastroenterol* 21(1):434. <https://doi.org/10.1186/s12876-021-02020-z>
850. Yuan S, Bäck M, Bruzelius M, Mason AM, Burgess S, Larsson S (2019) Plasma phospholipid fatty acids, FADS1 and risk of 15 cardiovascular diseases: a Mendelian randomisation study. *Nutrients* 11(12):3001. <https://doi.org/10.3390/nu11123001>
851. Bonaccorsi I, Carrega P, Venanzi Rullo E, Ducatelli R, Falco M, Freni J, Miceli M, Cavaliere R, Fontana V, Versace A et al (2021) HLA-C*17 in COVID-19 patients: Hints for associations with severe clinical outcome and cardiovascular risk. *Immunol Lett* 234:44–46. <https://doi.org/10.1016/j.imlet.2021.04.007>
852. Yu Y, Shi H, Wang Y, Yu Y, Chen R (2024) A pilot study of S100A4, S100A8/A9, and S100A12 in dilated cardiomyopathy: novel biomarkers for diagnosis or prognosis? *ESC Heart Fail* 11(1):503–512. <https://doi.org/10.1002/ehf2.14605>
853. Wenwang L (2023) Susceptibility of MMP3 gene polymorphism to coronary artery disease: A meta-analysis. *J Med Biochem* 42(4):685–693. <https://doi.org/10.5937/jomb0-43315>
854. Wang D, Fang J, Lv J, Pan Z, Yin X, Cheng H, Guo X (2019) Novel polymorphisms in PDLIM3 and PDLIM5 gene encoding Z-line proteins increase risk of idiopathic dilated cardiomyopathy. *J Cell Mol Med* 23(10):7054–7062. <https://doi.org/10.1111/jcmm.14607>
855. Duan X, Zhu T, Chen C, Zhang G, Zhang J, Wang L, Zhang L, Wang M, Wang X (2018) Serum glial cell line-derived neurotrophic factor levels and postoperative cognitive dysfunction after surgery for rheumatic heart disease. *J Thorac Cardiovasc Surg* 155(3):958–965.e1. <https://doi.org/10.1016/j.jtcvs.2017.07.073>
856. Jeong A, Lim Y, Kook T, Kwon DH, Cho YK, Ryu J, Lee YG, Shin S, Choe N, Kim YS et al (2023) Circular RNA circSMAD4 regulates cardiac fibrosis by targeting miR-671-5p and FGFR2 in cardiac fibroblasts. *Mol Ther Nucleic Acids* 34:102071. <https://doi.org/10.1016/j.omtn.2023.102071>
857. Thanikachalam PV, Ramamurthy S, Mallapu P, Varma SR, Narayanan J, Abourehab MA, Kesharwani P (2023) Modulation of IL-33/ST2 signaling as a potential new therapeutic target for cardiovascular diseases. *Cytokine Growth Factor Rev* 71–72:94–104. <https://doi.org/10.1016/j.cytogfr.2023.06.003>
858. Pius-Sadowska E, Machaliński B (2021) Pleiotropic activity of nerve growth factor in regulating cardiac functions and counteracting pathogenesis. *ESC Heart Fail* 8(2):974–987. <https://doi.org/10.1002/ehf2.13138>
859. Yan T, Song S, Sun W, Ge Y (2024) HAPLN1 knockdown inhibits heart failure development via activating the PKA signaling pathway. *BMC Cardiovasc Disord* 24(1):197. <https://doi.org/10.1186/s12872-024-03861-8>
860. Wang R, Wu Y, Jiang S (2021) FOXC2 alleviates myocardial ischemia-reperfusion injury in rats through regulating Nrf2/HO-1 signaling pathway. *Dis Markers* 2021:9628521. <https://doi.org/10.1155/2021/9628521>
861. Fu Y, Kong W (2017) Cartilage oligomeric matrix protein: matricellular and matricrine signaling in cardiovascular homeostasis and disease. *Curr Vasc Pharmacol* 15(3):186–196. <https://doi.org/10.2174/157016115666170201121232>
862. Pan J, Nilsson J, Engström G, De Marinis Y (2024) Elevated circulating folistatin associates with increased risk of mortality and cardiometabolic disorders. *Nutr Metab Cardiovasc Dis* 34(2):418–425. <https://doi.org/10.1016/j.numecd.2023.09.012>
863. Bugiardini E, Nunes AM, Oliveira-Santos A, Dagda M, Fontelonga TM, Barraza-Flores P, Pittman AM, Morrow JM, Parton M, Houlden H et al (2022) Integrin $\alpha 7$ mutations are associated with adult-onset cardiac dysfunction in humans and mice. *J Am Heart Assoc* 11(23):e026494. <https://doi.org/10.1161/JAHA.122.026494>
864. Liu MM, Peng J, Guo YL, Zhu CG, Wu NQ, Xu RX, Dong Q, Cui CJ, Li JJ (2022) SORBS2 as a molecular target for atherosclerosis in patients with familial hypercholesterolemia. *J Transl Med* 20(1):233. <https://doi.org/10.1186/s12967-022-03381-z>
865. Yokoe S, Asahi M (2017) Phospholamban is downregulated by pVHL-mediated degradation through oxidative stress in failing heart. *Int J Mol Sci* 18(11):2232. <https://doi.org/10.3390/ijms18112232>
866. Wu H, Wu H, He Y, Sun W, Meng Y, Wen B, Chu M (2023) Functional characterization of GATA6 genetic variants associated with mild congenital heart defects. *Biochem Biophys Res Commun* 641:77–83. <https://doi.org/10.1016/j.bbrc.2022.12.004>
867. Xu Y, Jiang K, Su F, Deng R, Cheng Z, Wang D, Yu Y, Xiang Y (2023) A transient wave of Bhlhe41+ resident macrophages enables remodeling of the developing infarcted myocardium. *Cell Rep* 42(10):113174. <https://doi.org/10.1016/j.celrep.2023.113174>
868. Koch CD, Lee CM, Apte SS (2020) Aggrexin in cardiovascular development and disease. *J Histochem Cytochem* 68(11):777–795. <https://doi.org/10.1369/0022155420952902>
869. Hu H, Lin S, Wang S, Chen X (2020) The role of transcription factor 21 in epicardial cell differentiation and the development of coronary heart disease. *Front Cell Dev Biol* 8:457. <https://doi.org/10.3389/fcell.2020.00457>
870. Gao F, Zhao Y, Zhang B, Xiao C, Sun Z, Gao Y, Dou X (2022) Suppression of lncRNA Gm47283 attenuates myocardial infarction via miR-706/Ptgs2/ferroptosis axis. *Bioengineered* 13(4):10786–10802. <https://doi.org/10.1080/21655979.2022.2065743>
871. Hou J, Yue Y, Hu B, Xu G, Su R, Lv L, Huang J, Yao J, Guan Y, Wang K et al (2019) DACT1 involvement in the cytoskeletal arrangement of cardiomyocytes in atrial fibrillation by regulating Cx43. *Braz J Cardiovasc Surg* 34(6):711–722. <https://doi.org/10.21470/1678-9741-2019-0033>
872. Franco D, Campione M (2003) The role of Pitx2 during cardiac development. Linking left-right signaling and congenital heart diseases. *Trends Cardiovasc Med* 13(4):157–163. [https://doi.org/10.1016/s1050-1738\(03\)00039-2](https://doi.org/10.1016/s1050-1738(03)00039-2)
873. Liang J, Cao Y, He M, Li W, Huang G, Ma T, Li M, Huang Y, Huang X, Hu Y (2021) AKR1C3 and its transcription factor HOXB4 are promising diagnostic biomarkers for acute myocardial infarction. *Front Cardiovasc Med* 8:694238. <https://doi.org/10.3389/fcvm.2021.694238>

874. Fioranelli M, Rocchia MG, Przybylsek B, Garo ML (2023) The role of brain-derived neurotrophic factor (BDNF) in depression and cardiovascular disease: a systematic review. *Life (Basel)* 13(10):1967. <https://doi.org/10.3390/life13101967>
875. Zhao J, Zhao Q, Mao S (2021) N-myc downstream regulated gene 2 ameliorates myocardial remodeling and cardiac function in heart failure rats. *Hum Exp Toxicol* 40(8):1296–1307. <https://doi.org/10.1177/0960327121993208>
876. Lee SH, Yang DK, Choi BY, Lee YH, Kim SY, Jeong D, Hajjar RJ, Park WJ (2009) The transcription factor Eya2 prevents pressure overload-induced adverse cardiac remodeling. *J Mol Cell Cardiol* 46(4):596–605. <https://doi.org/10.1016/j.jmcc.2008.12.021>
877. Zhang S, Wang Y, Yu M, Shang Y, Chang Y, Zhao H, Kang Y, Zhao L, Xu L, Zhao X et al (2022) Discovery of herbacetin as a novel SGK1 inhibitor to alleviate myocardial hypertrophy. *Adv Sci (Weinh)* 9(2):e2101485. <https://doi.org/10.1002/adv.202101485>
878. Shan T, Li X, Xie W, Wang S, Gao Y, Zheng Y, Su G, Li Y, Zhao Z (2024) Rap1GAP exacerbates myocardial infarction by regulating the AMPK/SIRT1/NF- κ B signaling pathway. *Cell Signal* 117:111080. <https://doi.org/10.1016/j.cellsig.2024.111080>
879. Dill TL, Naya FJ (2018) A hearty dose of noncoding RNAs: the imprinted DLK1-DIO3 locus in cardiac development and disease. *J Cardiovasc Dev Dis* 5(3):37. <https://doi.org/10.3390/jcdd5030037>
880. Zhuang C, Chen R, Zheng Z, Lu J, Hong C (2022) Toll-like receptor 3 in cardiovascular diseases. *Heart Lung Circ* 31(7):e93–e109. <https://doi.org/10.1016/j.hlc.2022.02.012>
881. Katanasaka Y, Saito A, Sunagawa Y, Sari N, Funamoto M, Shimizu S, Shimizu K, Akimoto T, Ueki C, Kitano M et al (2022) ANGPTL4 expression is increased in epicardial adipose tissue of patients with coronary artery disease. *J Clin Med* 11(9):2449. <https://doi.org/10.3390/jcm11092449>
882. Su SA, Yang D, Wu Y, Xie Y, Zhu W, Cai Z, Shen J, Fu Z, Wang Y, Jia L et al (2017) EphrinB2 regulates cardiac fibrosis through modulating the interaction of stat3 and TGF- β /Smad3 signaling. *Circ Res* 121(6):617–627. <https://doi.org/10.1161/CIRCRESAHA.117.311045>
883. Xiao J, Zhang Y, Tang Y, Dai H, OuYang Y, Li C, Yu M (2021) hsa-miR-4443 inhibits myocardial fibroblast proliferation by targeting THBS1 to regulate TGF- β 1/ α -SMA/collagen signaling in atrial fibrillation. *Braz J Med Biol Res* 54(4):e10692. <https://doi.org/10.1590/1414-431X202010692>
884. Kondo T, Higo S, Shiba M, Kohama Y, Kameda S, Tabata T, Inoue H, Okuno S, Ogawa S, Nakamura S et al (2022) Human-induced pluripotent stem cell-derived cardiomyocyte model for TNNT2 Δ 160E-induced cardiomyopathy. *Circ Genom Precis Med* 15(5):e003522. <https://doi.org/10.1161/CIRCGEN.121.003522>
885. Vendrov AE, Vendrov KC, Smith A, Yuan J, Sumida A, Robidoux J, Runge MS, Madamanchi NR (2015) NOX4 NADPH oxidase-dependent mitochondrial oxidative stress in aging-associated cardiovascular disease. *Antioxid Redox Signal* 23(18):1389–1409. <https://doi.org/10.1089/ars.2014.6221>
886. Xiong F, Mao R, Zhao R, Zhang L, Tan K, Liu C, Wang S, Xu M, Li Y, Zhang T (2022) Plasma exosomal S1PR5 and CARN1 as potential non-invasive screening biomarkers of coronary heart disease. *Front Cardiovasc Med* 9:845673. <https://doi.org/10.3389/fcvm.2022.845673>
887. Li C, Lv LF, Qi-Li MG, Yang R, Wang YJ, Chen SS, Zhang MX, Li TY, Yu T, Zhou YH et al (2022) Endocytosis of peptidase inhibitor SerpinE2 promotes myocardial fibrosis through activating ERK1/2 and β -catenin signaling pathways. *Int J Biol Sci* 18(16):6008–6019. <https://doi.org/10.7150/ijbs.67726>
888. Wei X, Ma X, Lu R, Bai G, Zhang J, Deng R, Gu N, Feng N, Guo X (2014) Genetic variants in PCSK1 gene are associated with the risk of coronary artery disease in type 2 diabetes in a Chinese Han population: a case control study. *PLoS ONE* 9(1):e87168. <https://doi.org/10.1371/journal.pone.0087168>
889. Schwaneckamp JA, Lorts A, Sargent MA, York AJ, Grimes KM, Fischesser DM, Gokey JJ, Whitsett JA, Conway SJ, Molkentin JD (2017) TGFB1 functions similar to periostin but is uniquely dispensable during cardiac injury. *PLoS ONE* 12(7):e0181945. <https://doi.org/10.1371/journal.pone.0181945>
890. Watanabe D, Nakato D, Yamada M, Suzuki H, Takenouchi T, Miya F, Kosaki K (2024) SALL4 deletion and kidney and cardiac defects associated with VACTERL association. *Pediatr Nephrol*. <https://doi.org/10.1007/s00467-024-06306-8>
891. Williams T, Hundertmark M, Nordbeck P, Voll S, Arias-Loza PA, Oppelt D, Mühlfelder M, Schraut S, Elsner I, Czolbe M et al (2015) Eya4 induces hypertrophy via regulation of p27kip1. *Circ Cardiovasc Genet* 8(6):752–764. <https://doi.org/10.1161/CIRCGENETICS.115.001134>
892. Gonzalez-Parra E, Rojas-Rivera J, Tuñón J, Praga M, Ortiz A, Egido J (2012) Vitamin D receptor activation and cardiovascular disease. *Nephrol Dial Transplant* 27:iv17–iv21. <https://doi.org/10.1093/ndt/gfs534>
893. Muendlein A, Heinzele C, Leiberer A, Brandtner EM, Geiger K, Gaenger S, Fraunberger P, Mader A, Saely CH, Drexel H (2023) Circulating glypican-4 is a new predictor of all-cause mortality in patients with heart failure. *Clin Biochem* 121–122:110675. <https://doi.org/10.1016/j.clinbiochem.2023.110675>
894. Castillo-Avila RG, González-Castro TB, Tovilla-Zárate CA, Martínez-Magaña JJ, López-Narváez ML, Juárez-Rojop IE, Arias-Vázquez PI, Borgonio-Cuadra VM, Pérez-Hernández N, Rodríguez-Pérez JM (2023) Association between genetic variants of CELSR2-PSRC1-SORT1 and cardiovascular diseases: a systematic review and meta-analysis. *J Cardiovasc Dev Dis* 10(3):91. <https://doi.org/10.3390/jcdd10030091>
895. Li J, Dong W, Gao X, Chen W, Sun C, Li J, Gao S, Zhang Y, He J, Lu D et al (2021) EphA4 is highly expressed in the atria of heart and its deletion leads to atrial hypertrophy and electrocardiographic abnormalities in rats. *Life Sci* 278:119595. <https://doi.org/10.1016/j.lfs.2021.119595>
896. Guay SP, Légaré C, Brisson D, Mathieu P, Bossé Y, Gaudet D, Bouchard L (2016) Epigenetic and genetic variations at the TNNT1 gene locus are associated with HDL-C levels and coronary artery disease. *Epigenomics* 8(3):359–371. <https://doi.org/10.2217/epi.15.120>
897. Murphy NP, Lubbers ER, Mohler PJ (2020) Advancing our understanding of AnkrD1 in cardiac development and disease. *Cardiovasc Res* 116(8):1402–1404. <https://doi.org/10.1093/cvr/cvaa063>
898. Pu T, Liu Y, Xu R, Li F, Chen S, Sun K (2018) Identification of ZFPM2 mutations in sporadic conotruncal heart defect patients. *Mol Genet Genom* 293(1):217–223. <https://doi.org/10.1007/s00438-017-1373-6>
899. Rui L, Liu R, Jiang H, Liu K (2022) Sox9 promotes cardiomyocyte apoptosis after acute myocardial infarction by promoting miR-223-3p and inhibiting MEF2C. *Mol Biotechnol* 64(8):902–913. <https://doi.org/10.1007/s12033-022-00471-7>
900. Roumeliotis S, Dounousi E, Eleftheriadis T, Liakopoulos V (2019) Association of the inactive circulating matrix gla protein with vitamin K intake, calcification, mortality, and cardiovascular disease: a review. *Int J Mol Sci* 20(3):628. <https://doi.org/10.3390/ijms20030628>
901. Qian P, Cao X, Xu X, Duan M, Zhang Q, Huang G (2020) Contribution of CYP24A1 variants in coronary heart disease among the Chinese population. *Lipids Health Dis* 19(1):181. <https://doi.org/10.1186/s12944-020-01356-x>
902. Xia H, Huang X, Deng S, Xu H, Yang Y, Liu X, Yuan L, Deng H (2021) DNAH11 compound heterozygous variants cause heterotaxy and congenital heart disease. *PLoS ONE* 16(6):e0252786. <https://doi.org/10.1371/journal.pone.0252786>
903. Roshan MH, Tambo A, Pace NP (2016) The role of TLR2, TLR4, and TLR9 in the pathogenesis of atherosclerosis. *Int J Inflamm* 2016:1532832. <https://doi.org/10.1155/2016/1532832>
904. Kreiner FF, Kraaijenhof JM, von Herrath M, Hovingh GKK, von Scholten BJ (2022) Interleukin 6 in diabetes, chronic kidney disease, and cardiovascular disease: mechanisms and therapeutic perspectives. *Exp Rev Clin Immunol* 18(4):377–389. <https://doi.org/10.1080/1744666X.2022.2045952>
905. Ma T, Huang R, Xu Y, Lv Y, Liu Y, Pan X, Dong J, Gao D, Wang Z, Zhang F et al (2023) Plasma GAS6 predicts mortality risk in acute heart failure patients: insights from the DRAGON-HF trial. *J Transl Med* 21(1):21. <https://doi.org/10.1186/s12967-022-03859-w>
906. Ng A, Wong M, Viviano B, Erlich JM, Alba G, Pflederer C, Jay PY, Saunders S (2009) Loss of glypican-3 function causes growth factor-dependent defects in cardiac and coronary vascular development. *Dev Biol* 335(1):208–215. <https://doi.org/10.1016/j.ydbio.2009.08.029>
907. Werner P, Paluru P, Simpson AM, Latney B, Iyer R, Brodeur GM, Goldmuntz E (2014) Mutations in NTRK3 suggest a novel signaling pathway in human congenital heart disease. *Hum Mutat* 35(12):1459–1468. <https://doi.org/10.1002/humu.2268>
908. Ma X, Hu P, Chen H, Fang T (2019) Loss of AMIGO2 causes dramatic damage to cardiac preservation after ischemic injury. *Cardiol J* 26(4):394–404. <https://doi.org/10.5603/CJ.a2018.0049>

909. Cavusoglu E, Kornecki E, Sobocka MB, Babinska A, Ehrlich YH, Chopra V, Yanamadala S, Ruwende C, Salifu MO, Clark LT et al (2007) Association of plasma levels of F11 receptor/junctional adhesion molecule-A (F11R/JAM-A) with human atherosclerosis. *J Am Coll Cardiol* 50(18):1768–1776. <https://doi.org/10.1016/j.jacc.2007.05.051>
910. Yu P, Zhao J, Jiang H, Liu M, Yang X, Zhang B, Yu Y, Zhang L, Tong R, Liu G et al (2018) Neural cell adhesion molecule-1 may be a new biomarker of coronary artery disease. *Int J Cardiol* 257:238–242. <https://doi.org/10.1016/j.ijcard.2017.12.040>
911. Gao W, Guo N, Yan H, Zhao S, Sun Y, Chen Z (2024) Mycn ameliorates cardiac hypertrophy-induced heart failure in mice by mediating the USP2/JUP/Akt/ β -catenin cascade. *BMC Cardiovasc Disord* 24(1):82. <https://doi.org/10.1186/s12872-024-03748-8>
912. Dabravolski SA, Sukhorukov VN, Kalmykov VA, Grechko AV, Shakhpazyan NK, Orekhov AN (2022) The role of KLF2 in the regulation of atherosclerosis development and potential use of KLF2-targeted therapy. *Biomedicines* 10(2):254. <https://doi.org/10.3390/biomedicines10020254>
913. Hsu A, Duan Q, McMahon S, Huang Y, Wood SA, Gray NS, Wang B, Bruneau BG, Haldar SM (2020) Salt-inducible kinase 1 maintains HDAC7 stability to promote pathologic cardiac remodeling. *J Clin Invest* 130(6):2966–2977. <https://doi.org/10.1172/JCI133753>
914. Gao W, Guo N, Zhao S, Chen Z, Zhang W, Yan F, Liao H, Chi K (2020) FBXW7 promotes pathological cardiac hypertrophy by targeting EZH2-SIX1 signaling. *Exp Cell Res* 393(1):112059. <https://doi.org/10.1016/j.yexcr.2020.112059>
915. Wu JJ, Jin J, Li YH, Wang C, Bai J, Jiang QJ, He TX, Nie SJ, Li DJ, Qu LF (2023) LncRNA FGF7-5 and lncRNA GLRX3 together inhibits the formation of carotid plaque via regulating the miR-2681-5p/ERCC4 axis in atherosclerosis. *Cell Cycle* 22(2):165–182. <https://doi.org/10.1080/15384101.2022.2110446>
916. Hong L, Lai HL, Fang Y, Tao Y, Qiu Y (2018) Silencing CTGF/CCN2 inactivates the MAPK signaling pathway to alleviate myocardial fibrosis and left ventricular hypertrophy in rats with dilated cardiomyopathy. *J Cell Biochem* 119(11):9519–9531. <https://doi.org/10.1002/jcb.27268>
917. Douglas G, Mehta V, Al Haj Zen A, Akoumianakis I, Goel A, Rashbrook VS, Trelfa L, Donovan L, Drydale E, Chuaiphichai S et al (2020) A key role for the novel coronary artery disease gene JCAD in atherosclerosis via shear stress mechanotransduction. *Cardiovasc Res* 116(11):1863–1874. <https://doi.org/10.1093/cvr/cvz263>
918. Aneke-Nash CS, Xue X, Qi Q, Biggs ML, Cappola A, Kuller L, Pollak M, Psaty BM, Siscovick D, Mukamal K et al (2017) The association between IGF-1 and IGFBP-3 and incident diabetes in an older population of men and women in the cardiovascular health study. *J Clin Endocrinol Metab* 102(12):4541–4547. <https://doi.org/10.1210/jc.2017-01273>
919. Jiang X, Cui J, Yang C, Song Y, Yuan J, Liu S, Hu F, Yang W, Qiao S (2020) Elevated lymphatic vessel density measured by Lyve-1 expression in areas of replacement fibrosis in the ventricular septum of patients with hypertrophic obstructive cardiomyopathy (HOCM). *Heart Vessels* 35(1):78–85. <https://doi.org/10.1007/s00380-019-01463-5>
920. Massadeh S, Albeladi M, Albeshar N, Alhabshan F, Kampe KD, Chai-khouni F, Kabbani MS, Beetz C, Alaamery M (2021) Novel autosomal recessive splice-altering variant in PRKD1 is associated with congenital heart disease. *Genes (Basel)* 12(5):612. <https://doi.org/10.3390/genes12050612>
921. Scuruchi M, Poti F, Rodríguez-Carrijo J, Campo GM, Mandraffino G (2020) Biglycan and atherosclerosis: lessons from high cardiovascular risk conditions. *Biochim Biophys Acta Mol Cell Biol Lipids* 1865(2):158545. <https://doi.org/10.1016/j.bbalip.2019.158545>
922. Toprak K, Kaplangoray M, Palice A, Taşcanov MB, Altıparmak İH, Biçer A, Demirbağ R (2023) Ectodysplasin a is associated with the presence and severity of coronary artery disease and poor long-term clinical outcome in patients presenting with ST-elevation myocardial infarction. *Acta Clin Belg* 78(4):270–279. <https://doi.org/10.1080/17843286.2022.2140246>
923. Liu Y, Yin Z, Xu X, Liu C, Duan X, Song Q, Tuo Y, Wang C, Yang J, Yin S (2021) Crosstalk between the activated Slit2-Robo1 pathway and TGF- β 1 signalling promotes cardiac fibrosis. *ESC Heart Fail* 8(1):447–460. <https://doi.org/10.1002/ehf2.13095>
924. Chong JX, Childers MC, Marvin CT, Marcello AJ, Gonorazky H, Hazrati LN, Dowling JJ, Al Amrani F, Alanay Y, Nieto Y et al (2023) Variants in ACTC1 underlie distal arthrogyposis accompanied by congenital heart defects. *HGG Adv* 4(3):100213. <https://doi.org/10.1016/j.xhgg.2023.100213>
925. Dhupar R, Powers AA, Eisenberg SH, Gemmill RM, Bardawil CE, Udoh HM, Cubitt A, Nangle LA, Soloff AC (2024) Orchestrating resilience: how neuropilin-2 and macrophages contribute to cardiothoracic disease. *J Clin Med* 13(5):1446. <https://doi.org/10.3390/jcm13051446>
926. Tsantilas P, Lao S, Wu Z, Eberhard A, Winski G, Vaerst M, Nanda V, Wang Y, Kojima Y, Ye J et al (2021) Chitinase 3 like 1 is a regulator of smooth muscle cell physiology and atherosclerotic lesion stability. *Cardiovasc Res* 117(14):2767–2780. <https://doi.org/10.1093/cvr/cvab014>
927. Qi P, Zhai Q, Zhang X (2023) RUNX1 facilitates heart failure progression through regulating TGF- β -induced cardiac remodeling. *PeerJ* 11:e16202. <https://doi.org/10.7717/peerj.16202>
928. Gao S, Chen H (2023) Therapeutic potential of apelin and Elabela in cardiovascular disease. *Biomed Pharmacother* 166:115268. <https://doi.org/10.1016/j.biopha.2023.115268>
929. Ji M, Su L, Liu L, Zhuang M, Xiao J, Guan Y, Zhu S, Ma L, Pu H (2023) CaM-KII regulates the proteins TPM1 and MYOM2 and promotes diacetyl-morphine-induced abnormal cardiac rhythms. *Sci Rep* 13(1):5827. <https://doi.org/10.1038/s41598-023-32941-6>
930. Martin RI, Babaei MS, Choy MK, Owens WA, Chico TJ, Keenan D, Yonan N, Koref MS, Keavney BD (2015) Genetic variants associated with risk of atrial fibrillation regulate expression of PITX2, CAV1, MYOZ1, C9orf3 and FANCC. *J Mol Cell Cardiol* 85:207–214. <https://doi.org/10.1016/j.jmcc.2015.06.005>
931. Tulbah S, Alruwaili N, Alhashem A, Aljohany A, Alhadeq F, Brotons DCA, Alwadai A, Al-Hassnan ZN (2024) Variable phenotype of a null PPP1R13L allele in children with dilated cardiomyopathy. *Am J Med Genet A* 194(1):59–63. <https://doi.org/10.1002/ajmg.a.63402>
932. Hsu CH, Liu IF, Kuo HF, Li CY, Lian WS, Chang CY, Chen YH, Liu WL, Lu CY, Liu YR et al (2021) miR-29a-3p/THBS2 axis regulates PAH-induced cardiac fibrosis. *Int J Mol Sci* 22(19):10574. <https://doi.org/10.3390/ijms221910574>
933. Piesanen J, Kunnas T, Nikkari ST (2022) The gene variant for desmin rs1058261 may protect against combined cancer and cardiovascular death, the Tampere adult population cardiovascular risk study. *Medicine (Baltimore)* 101(40):e31005. <https://doi.org/10.1097/MD.000000000000031005>
934. Prasongsukarn K, Dechkhajorn W, Benjathummarak S, Maneerat Y (2021) TRPM2, PDLIM5, BCL3, CD14, GBA genes as feasible markers for premature coronary heart disease risk. *Front Genet* 12:598296. <https://doi.org/10.3389/fgene.2021.598296>
935. Cappola TP, Li M, He J, Ky B, Gilmore J, Qu L, Keating B, Reilly M, Kim CE, Glessner J et al (2010) Common variants in HSPB7 and FRMD4B associated with advanced heart failure. *Circ Cardiovasc Genet* 3(2):147–154. <https://doi.org/10.1161/CIRCGENETICS.109.898395>
936. Wang Y, Wei J, Zhang P, Zhang X, Wang Y, Chen W, Zhao Y, Cui X (2022) Neuregulin-1, a potential therapeutic target for cardiac repair. *Front Pharmacol* 13:945206. <https://doi.org/10.3389/fphar.2022.945206>
937. Künzel SR, Hoffmann M, Weber S, Künzel K, Kämmerer S, Günscht M, Klapproth E, Rausch JSE, Sadek MS, Kolanowski T et al (2021) Diminished PLK2 induces cardiac fibrosis and promotes atrial fibrillation. *Circ Res* 129(8):804–820. <https://doi.org/10.1161/CIRCRESAHA.121.319425>
938. Chen X, Li X, Wu X, Ding Y, Li Y, Zhou G, Wei Y, Chen S, Lu X, Xu J et al (2023) Integrin beta-like 1 mediates fibroblast-cardiomyocyte crosstalk to promote cardiac fibrosis and hypertrophy. *Cardiovasc Res* 119(10):1928–1941. <https://doi.org/10.1093/cvr/cvad104>
939. Kassiteridi C, Cole JE, Griseri T, Falck-Hansen M, Goddard ME, Seneviratne AN, Green PA, Park I, Shami AG, Pattarabanjird T et al (2021) CD200 limits monopoiesis and monocyte recruitment in atherosclerosis. *Circ Res* 129(2):280–295. <https://doi.org/10.1161/CIRCRESAHA.121.316062>
940. Ivanova AA, Maksimov VN, Ivanoshchuk DE, Orlov PS, Novoselov VP, Savchenko SV, Voevoda MI (2017) Association of polymorphism in SCN5A, GJA5, and KCNN3 gene with sudden cardiac death. *Bull Exp Biol Med* 163(1):73–77. <https://doi.org/10.1007/s10517-017-3741-y>
941. Ortega A, Tarazón E, Roselló-Lletí E, Gil-Cayuela C, Lago F, González-Juanatey JR, Cinca J, Jorge E, Martínez-Dolz L, Portolés M et al (2015) Patients with dilated cardiomyopathy and sustained monomorphic ventricular tachycardia show up-regulation of KCNN3 and KCNJ2 genes and CACNG8-linked left ventricular dysfunction. *PLoS ONE* 10(12):e0145518. <https://doi.org/10.1371/journal.pone.0145518>

942. Li J, Chen Y, Gao J, Chen Y, Zhou C, Lin X, Liu C, Zhao M, Xu Y, Ji L et al (2021) Eva1a ameliorates atherosclerosis by promoting re-endothelialization of injured arteries via Rac1/Cdc42/Arpc1b. *Cardiovasc Res* 117(2):450–461. <https://doi.org/10.1093/cvr/cvaa011>
943. Zhou H, Che Y, Fu X, Wei H, Gao X, Chen Y, Zhang S (2019) Interaction between tissue factor pathway inhibitor-2 gene polymorphisms and environmental factors associated with coronary atherosclerosis in a Chinese Han. *J Thromb Thrombol* 47(1):67–72. <https://doi.org/10.1007/s11239-018-1755-6>
944. Zhang X, He D, Xiang Y, Wang C, Liang B, Li B, Qi D, Deng Q, Yu H, Lu Z et al (2022) DYSF promotes monocyte activation in atherosclerotic cardiovascular disease as a DNA methylation-driven gene. *Transl Res* 247:19–38. <https://doi.org/10.1016/j.trsl.2022.04.001>
945. Giguère H, Dumont AA, Berthiaume J, Oliveira V, Laberge G, Auger-Messier M (2018) ADAP1 limits neonatal cardiomyocyte hypertrophy by reducing integrin cell surface expression. *Sci Rep* 8(1):13605. <https://doi.org/10.1038/s41598-018-31784-w>
946. Wang HH, Luo WY, Lin M, Li XJ, Xiang GD, Triganti D (2021) Plasma asprosin, CCDC80 and ANGPTL4 levels are associated with metabolic and cardiovascular risk in patients with inflammatory bowel disease. *Physiol Res* 70(2):203–211. <https://doi.org/10.33549/physiolres.934547>
947. Zhang J, Zhang J, Zhou B, Jiang X, Tang Y, Zhang Z (2022) Death-associated protein kinase 1 (DAPK1) protects against myocardial injury induced by myocardial infarction in rats via inhibition of inflammation and oxidative stress. *Dis Markers* 2022:9651092. <https://doi.org/10.1155/2022/9651092>
948. Xiong H, Yang Q, Zhang X, Wang P, Chen F, Liu Y, Wang P, Zhao Y, Li S, Huang Y et al (2019) Significant association of rare variant p.Gly8Ser in cardiac sodium channel β -4 subunit SCN4B with atrial fibrillation. *Ann Hum Genet* 83(4):239–248. <https://doi.org/10.1111/ahg.12305>
949. Jiang F, Dong Y, Wu C, Yang X, Zhao L, Guo J, Li Y, Dong J, Zheng GY, Cao H et al (2011) Fine mapping of chromosome 3q22.3 identifies two haplotype blocks in ESYT3 associated with coronary artery disease in female Han Chinese. *Atherosclerosis* 218(2):397–403. <https://doi.org/10.1016/j.atherosclerosis.2011.06.017>
950. Fan LL, Liu L, Wang CY, Guo T, Luo H (2022) A novel nonsense mutation of ABCA8 in a patient with reduced HDL-c levels and atherosclerosis. *QJM* 115(5):321–322. <https://doi.org/10.1093/qjmed/hcac009>
951. Tamargo IA, Baek KI, Xu C, Kang DW, Kim Y, Andueza A, Williams D, Demos C, Villa-Roel N, Kumar S et al (2024) HEG1 protects against atherosclerosis by regulating stable flow-induced KLF2/4 expression in endothelial cells. *Circulation* 149(15):1183–1201. <https://doi.org/10.1161/CIRCULATIONAHA.123.064735>
952. Zhou QL, Teng F, Zhang YS, Sun Q, Cao YX, Meng GW (2018) FPR1 gene silencing suppresses cardiomyocyte apoptosis and ventricular remodeling in rats with ischemia/reperfusion injury through the inhibition of MAPK signaling pathway. *Exp Cell Res* 370(2):506–518. <https://doi.org/10.1016/j.yexcr.2018.07.016>
953. Hwang HS, Kahmini AR, Pracsak J, Cejas-Carbonell A, Valera IC, Champion S, Corrigan M, Mumbi F, Parvatiyar MS (2023) Sarcospan deficiency increases oxidative stress and arrhythmias in hearts after acute ischemia-reperfusion injury. *Int J Mol Sci* 24(14):11868. <https://doi.org/10.3390/ijms241411868>
954. Latella MC, Di Castelnuovo A, de Lorgeril M, Arnout J, Cappuccio FP, Krogh V, Siani A, van Dongen M, Donati MB, de Gaetano G et al (2009) Genetic variation of alcohol dehydrogenase type 1C (ADH1C), alcohol consumption, and metabolic cardiovascular risk factors: results from the IMMIDIET study. *Atherosclerosis* 207(1):284–290. <https://doi.org/10.1016/j.atherosclerosis.2009.04.022>
955. Jiang DS, Zhang XF, Gao L, Zong J, Zhou H, Liu Y, Zhang Y, Bian ZY, Zhu LH, Fan GC et al (2014) Signal regulatory protein- α protects against cardiac hypertrophy via the disruption of toll-like receptor 4 signaling. *Hypertension* 63(1):96–104. <https://doi.org/10.1161/HYPERTENSIONAHA.113.0150>
956. García SI, Pirola CJ (2005) Thyrotropin-releasing hormone in cardiovascular pathophysiology. *Regul Pept* 128(3):239–246. <https://doi.org/10.1016/j.regpep.2005.01.002>
957. Buensuceso AV, Son AI, Zhou R, Paquet M, Withers BM, Deroo BJ (2016) Ephrin-A5 is required for optimal fertility and a complete ovulatory response to gonadotropins in the female mouse. *Endocrinology* 157(2):942–955. <https://doi.org/10.1210/en.2015-1216>
958. Barišić A, Perez N, Hodžić A, Kapović M, Peterlin B, Ostojić S (2017) Functional single nucleotide polymorphisms of matrix metalloproteinase 7 and 12 genes in idiopathic recurrent spontaneous abortion. *J Assist Reprod Genet* 34(3):365–371. <https://doi.org/10.1007/s10815-016-0848-4>
959. Karaer A, Cigremis Y, Celik E, Urhan GR (2014) Prokineticin 1 and leukemia inhibitory factor mRNA expression in the endometrium of women with idiopathic recurrent pregnancy loss. *Fertil Steril* 102(4):1091–1095. e1. <https://doi.org/10.1016/j.fertnstert.2014.07.010>
960. Yang H, Lin J, Li H, Liu Z, Chen X, Chen Q (2021) Prolactin is associated with insulin resistance and beta-cell dysfunction in infertile women with polycystic ovary syndrome. *Front Endocrinol (Lausanne)* 12:571229. <https://doi.org/10.3389/fendo.2021.571229>
961. Mu J, Wang W, Chen B, Wu L, Li B, Mao X, Zhang Z, Fu J, Kuang Y, Sun X et al (2019) Mutations in NLRP2 and NLRP5 cause female infertility characterised by early embryonic arrest. *J Med Genet* 56(7):471–480. <https://doi.org/10.1136/jmedgenet-2018-105936>
962. Chang HM, Wu HC, Sun ZG, Lian F, Leung PCK (2019) Neurotrophins and glial cell line-derived neurotrophic factor in the ovary: physiological and pathophysiological implications. *Hum Reprod Update* 25(2):224–242. <https://doi.org/10.1093/humupd/dmy047>
963. Pan Z, Wang W, Wu L, Yao Z, Wang W, Chen Y, Gu H, Dong J, Mu J, Zhang Z et al (2024) Bi-allelic missense variants in MEI4 cause preimplantation embryonic arrest and female infertility. *Hum Genet*. <https://doi.org/10.1007/s00439-023-02633-2>
964. Drosch M, Schmidt N, Markowski DN, Zollner TM, Koch M, Bullerdiek J (2014) The CD24hi smooth muscle subpopulation is the predominant fraction in uterine fibroids. *Mol Hum Reprod* 20(7):664–676. <https://doi.org/10.1093/molehr/gau022>
965. Zhang Z, Wu L, Diao F, Chen B, Fu J, Mao X, Yan Z, Li B, Mu J, Zhou Z et al (2020) Novel mutations in LHCGR (luteinizing hormone/choriogonadotropin receptor): expanding the spectrum of mutations responsible for human empty follicle syndrome. *J Assist Reprod Genet* 37(11):2861–2868. <https://doi.org/10.1007/s10815-020-01931-2>
966. Shakerian B, Irvani S, Mostafavi S, Moghtaderi M (2022) Quantitative serum determination of CD3, CD4, CD8, CD16, and CD56 in women with primary infertility: the role of cell-mediated immunity. *Turk J Obstet Gynecol* 19(3):242–245. <https://doi.org/10.4274/tjod.galenos.2022.47527>
967. Hu J, Ke H, Luo W, Yang Y, Liu H, Li G, Qin Y, Ma J, Zhao S (2020) A novel FOXL2 mutation in two infertile patients with blepharophimosis-ptosis-epicanthus inversus syndrome. *J Assist Reprod Genet* 37(1):223–229. <https://doi.org/10.1007/s10815-019-01651-2>
968. Honarvar N, Sheikha MH, Farshahi Yazd E, Pashaeifar H, Mohtaram S, Sazegari A, Feizollahi Z, Ghasemi N (2016) KDR gene polymorphisms and idiopathic recurrent spontaneous abortion. *J Matern Fetal Neonatal Med* 29(22):3737–3740. <https://doi.org/10.3109/14767058.2016.1142966>
969. Zangeneh FZ, Bagheri M, Shoushtari MS, Naghizadeh MM (2021) Expression of ADR- α 1, 2 and ADR- β 2 in cumulus cell culture of infertile women with polycystic ovary syndrome and poor responder who are a candidate for IVF: the novel strategic role of clonidine in this expression. *J Recept Signal Transduct Res* 41(3):263–272. <https://doi.org/10.1080/10799893.2020.1806320>
970. Heidarzadehpilehrood R, Pirhoushiaran M, Abdollahzadeh R, Binti Osman M, Sakinah M, Nordin N, Abdul HH (2022) A review on CYP11A1, CYP17A1, and CYP19A1 polymorphism studies: candidate susceptibility genes for polycystic ovary syndrome (PCOS) and infertility. *Genes (Basel)* 13(2):302. <https://doi.org/10.3390/genes13020302>
971. Bouilly J, Messina A, Papadakis G, Cassatella D, Xu C, Acierno JS, Tata B, Sykiotis G, Santini S, Sidis Y et al (2018) DCC/NTN1 complex mutations in patients with congenital hypogonadotropic hypogonadism impair GnRH neuron development. *Hum Mol Genet* 27(2):359–372. <https://doi.org/10.1093/hmg/ddx408>
972. Wang S, Fang L, Cong L, Chung JPW, Li TC, Chan DY (2022) Myostatin: a multifunctional role in human female reproduction and fertility - a short review. *Reprod Biol Endocrinol* 20(1):96. <https://doi.org/10.1186/s12958-022-00969-4>
973. Demiray SB, Yilmaz O, Goker ENT, Tavmergen E, Calimlioglu N, Sezerman U, Soykam HO, Oktem G (2017) Expression of the bone morphogenetic protein-2 (BMP2) in the human cumulus cells as a biomarker of oocytes

- and embryo quality. *J Hum Reprod Sci* 10(3):194–200. https://doi.org/10.4103/jhrs.JHRS_21_17
974. Li J, Chen Y, Wu H, Li L (2014) Apolipoprotein E (Apo E) gene polymorphisms and recurrent pregnancy loss: a meta-analysis. *J Assist Reprod Genet* 31(2):139–148. <https://doi.org/10.1007/s10815-013-0128-5>
 975. Al-Mutawa J (2018) Interaction with angiotensin-converting enzyme-encoding gene in female infertility: insertion and deletion polymorphism studies. *Saudi J Biol Sci* 25(8):1617–1621. <https://doi.org/10.1016/j.sjbs.2016.06.003>
 976. Rocha-Junior CV, Da Broi MG, Miranda-Furtado CL, Navarro PA, Ferriani RA, Meola J (2019) Progesterone receptor B (PGR-B) is partially methylated in eutopic endometrium from infertile women with endometriosis. *Reprod Sci* 26(12):1568–1574. <https://doi.org/10.1177/1933719119828078>
 977. Rydze RT, Patton BK, Briley SM, Salazar Torralba H, Gipson G, James R, Rajkovic A, Thompson T, Pangas SA (2021) Deletion of Grem1-2 alters estrous cyclicity and disrupts female fertility in mice†. *Biol Reprod* 105(5):1205–1220. <https://doi.org/10.1093/biolre/iaob148>
 978. Vatin M, Bouvier S, Bellazi L, Montagutelli X, Laissue P, Ziyat A, Serres C, De Mazancourt P, Dieudonné MN, Mornet E et al (2014) Polymorphisms of human placental alkaline phosphatase are associated with in vitro fertilization success and recurrent pregnancy loss. *Am J Pathol* 184(2):362–368. <https://doi.org/10.1016/j.ajpath.2013.10.024>
 979. Cardoso JV, Machado DE, da Silva MC, Berardo PT, Ferrari R, Abrão MS, Perini JA (2019) Matrix metalloproteinases 3 polymorphism increases the risk of developing advanced endometriosis and infertility: a case-control study. *Eur J Obstet Gynecol Reprod Biol X* 3:100041. <https://doi.org/10.1016/j.eurox.2019.100041>
 980. Chang HM, Wu HC, Sun ZG, Lian F, Leung PCK (2019) Neurotrophins and glial cell line-derived neurotrophic factor in the ovary: physiological and pathophysiological implications. *Hum Reprod Update* 25(2):224–242. <https://doi.org/10.1093/humupd/dmy047>
 981. Filant J, DeMayo FJ, Pru JK, Lydon JP, Spencer TE (2014) Fibroblast growth factor receptor two (FGFR2) regulates uterine epithelial integrity and fertility in mice. *Biol Reprod* 90(1):7. <https://doi.org/10.1095/biolreprod.113.114496>
 982. He B, Teng XM, Hao F, Zhao M, Chen ZQ, Li KM, Yan Q (2022) Decreased intracellular IL-33 impairs endometrial receptivity in women with adenomyosis. *Front Endocrinol (Lausanne)* 13:928024. <https://doi.org/10.3389/fendo.2022.928024>
 983. Palumbo MA, Giuffrida E, Gulino FA, Leonardi E, Cantarella G, Bernardini R (2013) Nerve growth factor (NGF) levels in follicular fluid of infertile patients undergoing to in vitro fertilization (IVF) cycle. *Gynecol Endocrinol* 29(11):1002–1004. <https://doi.org/10.3109/09513590.2013.829450>
 984. Mu Y, Zhou DN, Yan NN, Ding JL, Yang J (2019) Upregulation of ADAMTS-7 and downregulation of COMP are associated with spontaneous abortion. *Mol Med Rep* 19(4):2620–2626. <https://doi.org/10.3892/mmr.2019.9898>
 985. Norton KA, Niri F, Weatherill CB, Williams CE, Duong K, McDermid HE (2021) Implantation failure and embryo loss contribute to subfertility in female mice mutant for chromatin remodeler Ccr2†. *Biol Reprod* 104(4):835–849. <https://doi.org/10.1093/biolre/iaaa231>
 986. Lin SY, Craythorn RG, O'Connor AE, Matzuk MM, Girling JE, Morrison JR, de Kretser DM (2008) Female infertility and disrupted angiogenesis are actions of specific follistatin isoforms. *Mol Endocrinol* 22(2):415–429. <https://doi.org/10.1210/me.2006-05>
 987. Bennett J, Wu YG, Gossen J, Zhou P, Stocco C (2012) Loss of GATA-6 and GATA-4 in granulosa cells blocks folliculogenesis, ovulation, and follicle stimulating hormone receptor expression leading to female infertility. *Endocrinology* 153(5):2474–2485. <https://doi.org/10.1210/en.2011-1969>
 988. Sadeu JC, Doedée AM, Neal MS, Hughes EG, Foster WG (2012) Neurotrophins (BDNF and NGF) in follicular fluid of women with different infertility diagnoses. *Reprod Biomed Online* 24(2):174–179. <https://doi.org/10.1016/j.rbmo.2011.11.011>
 989. Salker MS, Christian M, Steel JH, Nautiyal J, Lavery S, Trew G, Webster Z, Al-Sabbagh M, Puchchakayala G, Föller M et al (2011) Deregulation of the serum- and glucocorticoid-inducible kinase SGK1 in the endometrium causes reproductive failure. *Nat Med* 17(11):1509–1513. <https://doi.org/10.1038/nm.2498>
 990. Li M, Hu J, Yao L, Gao M (2020) Decreased ANGPTL4 impairs endometrial angiogenesis during peri-implantation period in patients with recurrent implantation failure. *J Cell Mol Med* 24(18):10730–10743. <https://doi.org/10.1111/jcmm.15696>
 991. Bender HR, Campbell GE, Aytoda P, Mathiesen AH, Duffy DM (2019) Thrombospondin 1 (THBS1) promotes follicular angiogenesis, luteinization, and ovulation in primates. *Front Endocrinol (Lausanne)* 10:727. <https://doi.org/10.3389/fendo.2019.00727>
 992. Ding JL, Diao LH, Yin TL, Huang CY, Yin B, Chen C, Zhang Y, Li J, Cheng YX, Zeng Y et al (2017) Aberrant expressions of endometrial Id3 and CTLA-4 are associated with unexplained repeated implantation failure and recurrent miscarriage. *Am J Reprod Immunol*. <https://doi.org/10.1111/aji.12632>.doi:10.1111/aji.12632
 993. Maraldi T, Resca E, Nicoli A, Beretti F, Zavatti M, Capodanno F, Morini D, Palomba S, La Sala GB, De Pol A (2016) NADPH oxidase-4 and MATER expressions in granulosa cells: relationships with ovarian aging. *Life Sci* 162:108–114. <https://doi.org/10.1016/j.lfs.2016.08.007>
 994. Mu H, Cai S, Wang X, Li H, Zhang L, Li H, Xiang W (2022) RNA binding protein IGF2BP1 mediates oxidative stress-induced granulosa cell dysfunction by regulating MDM2 mRNA stability in a m6A-dependent manner. *Redox Biol* 57:102492. <https://doi.org/10.1016/j.redox.2022.102492>
 995. Wang Q, Li D, Cai B, Chen Q, Li C, Wu Y, Jin L, Wang X, Zhang X, Zhang F (2019) Whole-exome sequencing reveals SALL4 variants in premature ovarian insufficiency: an update on genotype-phenotype correlations. *Hum Genet* 138(1):83–92. <https://doi.org/10.1007/s00439-018-1962-4>
 996. Ashraf M, Khan HN, Ibrahim R, Shahid M, Khan S, Fatima A, Ullah S, Rehman R (2024) Genetic association of vitamin D receptor gene with female infertility. *Nucleosides Nucleotides Nucleic Acids* 43(2):116–133. <https://doi.org/10.1080/15257770.2023.2236167>
 997. Taghizadeh E, Kalantar SM, Mahdian R, Sheikha MH, Farashahi-Yazd E, Ghasemi S, Shahbazi Z (2015) SULF 1 gene polymorphism, rs6990375 is in significant association with fetus failure in IVF technique. *Iran J Reprod Med* 13(4):215–220
 998. Koval H, Chopiak V, Kamyshnyi A (2015) mRNA tlr2 and tlr4 expression in the endometrium tissue in women with endometriosis associated with infertility. *Georgian Med News* 244–245:7–11
 999. Incognito GG, Di Guardo F, Gulino FA, Genovese F, Benvenuto D, Lello C, Palumbo M (2023) Interleukin-6 as a useful predictor of endometriosis-associated infertility: a systematic review. *Int J Fertil Steril* 17(4):226–230. <https://doi.org/10.22074/ijfs.2023.557683.1329>
 1000. Gokce S, Herkiloglu D, Cevik O, Turan V (2023) Evaluation of intrafollicular syndecan 1, glypican 3, and spermidine levels in women with diminished ovarian reserve. *Reprod Sci* 30(2):569–575. <https://doi.org/10.1007/s43032-022-01085-9>
 1001. Núñez-Ollé M, Jung C, Terré B, Balsiger NA, Plata C, Roset R, Pardo-Pastor C, Garrido M, Rojas S, Alameda F et al (2017) Constitutive Cyclin O deficiency results in penetrant hydrocephalus, impaired growth and infertility. *Oncotarget* 8(59):99261–99273. <https://doi.org/10.18632/oncotarget.21818>
 1002. Yovich JL, Zaidi S, Nguyen MDK, Hincliffe PM (2020) Measuring IGF-1 and IGFBP-3 profiles in women seeking assisted reproduction; relationship to clinical parameters (Study 1). *J Pers Med* 10(3):122. <https://doi.org/10.3390/jpm10030122>
 1003. Chen Y, Sun T, Niu Y, Wang D, Liu K, Wang T, Wang S, Xu H, Liu J (2021) Cell adhesion molecule L1 like plays a role in the pathogenesis of idiopathic hypogonadotropic hypogonadism. *J Endocrinol Invest* 44(8):1739–1751. <https://doi.org/10.1007/s40618-020-01485-1>
 1004. Dorfman MD, Garcia-Rudaz C, Alderman Z, Kerr B, Lomniczi A, Dissen GA, Castellano JM, Garcia-Galiano D, Gaytan F, Xu B et al (2014) Loss of Ntrk2/Kiss1r signaling in oocytes causes premature ovarian failure. *Endocrinology* 155(8):3098–3111. <https://doi.org/10.1210/en.2014-1111>
 1005. Wu J, Fang Z, Wang X, Zeng W, Zhao Y, Jiang F, Chen DN, Zheng R, Li J, Men M et al (2022) SLIT2 rare sequencing variants identified in idiopathic hypogonadotropic hypogonadism. *Horm Res Paediatr* 95(4):384–392. <https://doi.org/10.1159/000525769>
 1006. Roche J, Ramé C, Reverchon M, Mellouk N, Cornuau M, Guerif F, Froment P, Dupont J (2016) Apelin (APLN) and apelin receptor (APLNR) in human ovary: expression, signaling, and regulation of steroidogenesis in primary human luteinized granulosa cells. *Biol Reprod* 95(5):104. <https://doi.org/10.1095/biolreprod.116.141754>

1007. Li L, Feng F, Zhao M, Li T, Yue W, Ma X, Wang B, Yin C (2020) NOTCH2 variant D1853H is mutated in two non-syndromic premature ovarian insufficiency patients from a Chinese pedigree. *J Ovarian Res* 13(1):41. <https://doi.org/10.1186/s13048-020-00645-4>
1008. Bender HR, Trau HA, Duffy DM (2018) Placental growth factor is required for ovulation, luteinization, and angiogenesis in primate ovulatory follicles. *Endocrinology* 159(2):710–722. <https://doi.org/10.1210/en.2017-00739>
1009. Margioulas-Siakou C, Prapas Y, Petousis S, Miliadis S, Ravanos K, Dagklis T, Kalogiannidis I, Mavromatidis G, Haitoglou C, Prapas N et al (2017) LIF endometrial expression is impaired in women with unexplained infertility while LIF-R expression in all infertility sub-groups. *Cytokine* 96:166–172. <https://doi.org/10.1016/j.cyto.2017.04.009>
1010. Clark DA (2009) Cell-surface CD200 may predict efficacy of paternal mononuclear leukocyte immunotherapy in treatment of human recurrent pregnancy loss. *Am J Reprod Immunol* 61(1):75–84. <https://doi.org/10.1111/j.1600-0897.2008.00665.x>
1011. Altmäe S, Salumets A, Bjuresten K, Kallak TK, Wänggren K, Landgren BM, Hovatta O, Stavreus-Evers A (2011) Tissue factor and tissue factor pathway inhibitors TFPI and TFPI2 in human secretory endometrium—possible link to female infertility. *Reprod Sci* 18(7):666–678. <https://doi.org/10.1177/19337191114400633>
1012. Lee YJ, Kim CH, Kwack JY, Ahn JW, Kim SH, Chae HD, Kang BM (2014) Subclinical hypothyroidism diagnosed by thyrotropin-releasing hormone stimulation test in infertile women with basal thyroid-stimulating hormone levels of 2.5 to 5.0 mIU/L. *Obstet Gynecol Sci* 57(6):507–512. <https://doi.org/10.5468/ogs.2014.57.6.507>
1013. Gokulakrishnan K, Pandey GK, Sathishkumar C, Sundararajan S, Durairaj P, Manickam N, Mohan V, Balasubramanyam M (2021) Augmentation of RBP4/STRA6 signaling leads to insulin resistance and inflammation and the plausible therapeutic role of vildagliptin and metformin. *Mol Biol Rep* 48(5):4093–4106. <https://doi.org/10.1007/s11033-021-06420-y>
1014. Rhyu HJ, Bae SH, Jung J, Hyun YM (2020) Cochlin-cleaved LCLL is a dual-armed regulator of the innate immune response in the cochlea during inflammation. *BMB Rep* 53(9):449–452. <https://doi.org/10.5483/BMBRep.2020.53.9.104>
1015. Petronilho F, Danielski LG, Roesler R, Schwartzmann G, Dal-Pizzol F (2013) Gastrin-releasing peptide as a molecular target for inflammatory diseases: an update. *Inflamm Allergy Drug Targets* 12(3):172–177. <https://doi.org/10.2174/1871528111312030003>
1016. Te Velde AA, Pronk I, de Kort F, Stokkers PC (2008) Glutathione peroxidase 2 and aquaporin 8 as new markers for colonic inflammation in experimental colitis and inflammatory bowel diseases: an important role for H2O2? *Eur J Gastroenterol Hepatol* 20(6):555–560. <https://doi.org/10.1097/MEG.0b013e3282f45751>
1017. Mao H, Han B, Li H, Tao Y, Wu W (2021) FABP4 knockdown suppresses inflammation, apoptosis and extracellular matrix degradation in IL-1 β -induced chondrocytes by activating PPAR γ to regulate the NF- κ B signaling pathway. *Mol Med Rep* 24(6):855. <https://doi.org/10.3892/mmr.2021.12495>
1018. Dai WJ, Qiu J, Sun J, Ma CL, Huang N, Jiang Y, Zeng J, Ren BC, Li WC, Li YH (2019) Downregulation of microRNA-9 reduces inflammatory response and fibroblast proliferation in mice with idiopathic pulmonary fibrosis through the ANO1-mediated TGF- β -Smad3 pathway. *J Cell Physiol* 234(3):2552–2565. <https://doi.org/10.1002/jcp.26961>
1019. Xu Y, Zhan X (2022) lncRNA KCNQ1OT1 regulated high glucose-induced proliferation, oxidative stress, extracellular matrix accumulation, and inflammation by miR-147a/SOX6 in diabetic nephropathy (DN). *Endocr J* 69(5):511–522. <https://doi.org/10.1507/endocrj.EJ21-0514>
1020. Zhang H, Ren C, Liu Q, Wang Q, Wang D (2023) TFAP2C exacerbates psoriasis-like inflammation by promoting Th17 and Th1 cells activation through regulating TEAD4 transcription. *Allergol Immunopathol (Madr)* 51(3):124–134. <https://doi.org/10.15586/aei.v51i3.854>
1021. Dong X, Tang Y (2022) Ntrk1 promotes mesangial cell proliferation and inflammation in rat glomerulonephritis model by activating the STAT3 and p38/ERK MAPK signaling pathways. *BMC Nephrol* 23(1):413. <https://doi.org/10.1186/s12882-022-03001-4>
1022. Le Quintrec M, Teisseyre M, Bec N, Delmont E, Szwarc J, Perrochia H, Machet MC, Chauvin A, Mavroudakos N, Taieb G et al (2021) Contactin-1 is a novel target antigen in membranous nephropathy associated with chronic inflammatory demyelinating polyneuropathy. *Kidney Int* 100(6):1240–1249. <https://doi.org/10.1016/j.kint.2021.08.014>
1023. Peng W, Song Y, Zhu G, Zeng Y, Cai H, Lu C, Abuduxukuer Z, Song X, Gao X, Ye L et al (2024) FGF10 attenuates allergic airway inflammation in asthma by inhibiting PI3K/AKT/NF- κ B pathway. *Cell Signal* 113:110964. <https://doi.org/10.1016/j.cellsig.2023.110964>
1024. Wolf M, Clay SM, Zheng S, Pan P, Chan MF (2019) MMP12 inhibits corneal neovascularization and inflammation through regulation of CCL2. *Sci Rep* 9(1):11579. <https://doi.org/10.1038/s41598-019-47831-z>
1025. Fang L, Shen R, Lu Y, Xu X, Huang F (2024) Tetrandrine alleviates inflammation and promotes macrophage M2 polarization in gouty arthritis by NF- κ B-mediated Lcp1. *Cell Mol Biol (Noisy-le-grand)* 70(2):205–211. <https://doi.org/10.14715/cmb/2024.70.2.29>
1026. Forloni G (2023) Alpha synuclein: neurodegeneration and inflammation. *Int J Mol Sci* 24(6):5914. <https://doi.org/10.3390/ijms24065914>
1027. Fyfe-Desmarais G, Desmarais F, Rassart É, Mounier C (2023) Apolipoprotein D in oxidative stress and inflammation. *Antioxidants (Basel)* 12(5):1027. <https://doi.org/10.3390/antiox12051027>
1028. Mair MJ, Kiesel B, Feldmann K, Widhalm G, Dieckmann K, Wöhrer A, Müllauer L, Preusser M, Berghoff AS et al (2021) LAG-3 expression in the inflammatory microenvironment of glioma. *J Neurooncol* 152(3):533–539. <https://doi.org/10.1007/s11060-021-03721-x>
1029. Berahovich RD, Miao Z, Wang Y, Premack B, Howard MC, Schall TJ (2005) Proteolytic activation of alternative CCR1 ligands in inflammation. *J Immunol* 174(11):7341–7351. <https://doi.org/10.4049/jimmunol.174.11.7341>
1030. Du L, Xu C, Tang K, Shi J, Tang L, Lisha X, Lei C, Liu H, Liang Y, Guo Y (2023) Epithelial CST1 promotes airway eosinophilic inflammation in asthma via the AKT signaling pathway. *Allergy Asthma Immunol Res* 15(3):374–394. <https://doi.org/10.4168/air.2023.15.3.374>
1031. Zhang X, Yang Z, Pan T, Long X, Sun Q, Wang PH, Li X, Kuang E (2022) SARS-CoV-2 ORF3a induces RETREG1/FAM134B-dependent reticulophagy and triggers sequential ER stress and inflammatory responses during SARS-CoV-2 infection. *Autophagy* 18(11):2576–2592. <https://doi.org/10.1080/15548627.2022.2039992>
1032. Xu F, Ren ZX, Zhong XM, Zhang Q, Zhang JY, Yang J (2019) Intrauterine inflammation damages placental angiogenesis via Wnt5a-Flt1 activation. *Inflammation* 42(3):818–825. <https://doi.org/10.1007/s10753-018-0936-y>
1033. Hou Y, Wei D, Bossila EA, Zhang Z, Li S, Bao J, Xu H, Zhang L, Zhao Y (2022) FABP5 deficiency impaired macrophage inflammation by regulating AMPK/NF- κ B signaling pathway. *J Immunol* 209(11):2181–2191. <https://doi.org/10.4049/jimmunol.2200182>
1034. Zhao Q, Xia N, Xu J, Wang Y, Feng L, Su D, Cheng Z (2023) Pro-Inflammatory of PRDM1/SIRT2/NLRP3 axis in monosodium urate-induced acute gouty arthritis. *J Innate Immun* 15(1):614–628. <https://doi.org/10.1159/000530966>
1035. Nishiyama K, Toyama C, Kato Y, Tanaka T, Nishimura A, Nagata R, Mori Y, Nishida M (2021) Deletion of TRPC3 or TRPC6 fails to attenuate the formation of inflammation and fibrosis in non-alcoholic steatohepatitis. *Biol Pharm Bull* 44(3):431–436. <https://doi.org/10.1248/bpb.b20-00903>
1036. Gorowiec MR, Catalano RD, Norman JE, Denison FC, Jabbour HN (2011) Prokineticin 1 induces inflammatory response in human myometrium: a potential role in initiating term and preterm parturition. *Am J Pathol* 179(6):2709–2719. <https://doi.org/10.1016/j.ajpath.2011.08.029>
1037. Liu X, Li X, Hua B, Yang X, Zheng J, Liu S (2021) WNT16 is upregulated early in mouse TMJ osteoarthritis and protects fibrochondrocytes against IL-1 β induced inflammatory response by regulation of RUNX2/MMP13 cascade. *Bone* 143:115793. <https://doi.org/10.1016/j.bone.2020.115793>
1038. Kaartinen MT, Arora M, Heinonen S, Hang A, Barry A, Lundbom J, Hakkarainen A, Lundholm N, Rissanen A, Kaprio J et al (2021) F13A1 transglutaminase expression in human adipose tissue increases in acquired excess weight and associates with inflammatory status of adipocytes. *Int J Obes (Lond)* 45(3):577–587. <https://doi.org/10.1038/s41366-020-00722-0>
1039. Zhou E, Ge X, Nakashima H, Li R, van der Zande HJP, Liu C, Li Z, Müller C, Bracher F, Mohammed Y et al (2023) Inhibition of DHCR24 activates LXRs to ameliorate hepatic steatosis and inflammation. *EMBO Mol Med* 15(8):e16845. <https://doi.org/10.15252/emmm.202216845>

1040. Zuo Y, Xu H, Li Y, Zhang Z, Tao R, Wang M (2023) Hsa_circ_0007707 participates in PDE3B-mediated apoptosis inhibition and inflammation promotion in fibroblast-like synoviocytes. *Int Immunopharmacol* 119:110157. <https://doi.org/10.1016/j.intimp.2023.110157>
1041. Yu GI, Song DK, Shin DH (2020) Associations of IL1RAP and IL1RL1 gene polymorphisms with obesity and inflammation mediators. *Inflamm Res* 69(2):191–202. <https://doi.org/10.1007/s00011-019-01307-y>
1042. Tsai AP, Dong C, Lin PB, Messenger EJ, Casali BT, Moutinho M, Liu Y, Oblak AL, Lamb BT, Landreth GE et al (2022) PLCG2 is associated with the inflammatory response and is induced by amyloid plaques in Alzheimer's disease. *Genome Med* 14(1):17. <https://doi.org/10.1186/s13073-022-01022-0>
1043. Ramos-Martinez E, Ramos-Martínez I, Molina-Salinas G, Zepeda-Ruiz WA, Carbon M (2021) The role of prolactin in central nervous system inflammation. *Rev Neurosci* 32(3):323–340. <https://doi.org/10.1515/revneuro-2020-0082>
1044. Zhuang J, Chen Z, Cai P, Wang R, Yang Q, Li L, Yang H, Zhu R (2020) Targeting MicroRNA-125b promotes neurite outgrowth but represses cell apoptosis and inflammation via blocking PTGS2 and CDK5 in a FOXQ1-dependent way in alzheimer disease. *Front Cell Neurosci* 14:587747. <https://doi.org/10.3389/fncel.2020.587747>
1045. Hayashi T, Nukui T, Piao JL, Sugimoto T, Anada R, Matsuda N, Yamamoto M, Konishi H, Dougu N, Nakatsuyi Y (2021) Serum neurofilament light chain in chronic inflammatory demyelinating polyneuropathy. *Brain Behav* 11(5):e02084. <https://doi.org/10.1002/brb3.2084>
1046. Huang Y, Xue Q, Chang J, Wang X, Miao C (2023) Wnt5a: A promising therapeutic target for inflammation, especially rheumatoid arthritis. *Cytokine* 172:156381. <https://doi.org/10.1016/j.cyto.2023.156381>
1047. Li Y, Zhang T, Tian W, Hu H, Xin Z, Ma X, Ye C, HZhao J, ang K, Han X, et al (2020) Loss of TIMP3 expression induces inflammation, matrix degradation, and vascular ingrowth in nucleus pulposus: a new mechanism of intervertebral disc degeneration. *FASEB J* 34(4):5483–5498. <https://doi.org/10.1096/fj.201902364RR>
1048. Vaheer H, Kivihall A, Runnel T, Raam L, Prans E, Maslovskaja J, Abram K, Kaldvee B, Mrowietz U, Weidinger S et al (2020) SERPINB2 and miR-146a/b are coordinately regulated and act in the suppression of psoriasis-associated inflammatory responses in keratinocytes. *Exp Dermatol* 29(1):51–60. <https://doi.org/10.1111/exd.14049>
1049. Huang X, Zhong L, van Helvoort E, Lafeber F, Mastbergen S, Hendriks J, Post JN, Karperien M (2021) The expressions of dickkopf-related protein 1 and frizzled-related protein are negatively correlated to local inflammation and osteoarthritis severity. *Cartilage* 12(4):496–504. <https://doi.org/10.1177/1947603519841676>
1050. Matsuoka Y, Yamashita A, Matsuda M, Kawai K, Sawa T, Amaya F (2019) NLRP2 inflammasome in dorsal root ganglion as a novel molecular platform that produces inflammatory pain hypersensitivity. *Pain* 160(9):2149–2160. <https://doi.org/10.1097/j.pain.0000000000001611>
1051. Dahm PH, Richards JB, Karmouty-Quintana H, Cromar KR, Sur S, Price RE, Malik F, Spencer CY, Barreno RX, Hashmi SS et al (2014) Effect of antigen sensitization and challenge on oscillatory mechanics of the lung and pulmonary inflammation in obese carboxypeptidase E-deficient mice. *Am J Physiol Regul Integr Comp Physiol* 307(6):R621–R633. <https://doi.org/10.1152/ajpregu.00205.2014>
1052. Yang S, Yin W, Ding Y, Liu F (2020) Lnc RNA ZFAS1 regulates the proliferation, apoptosis, inflammatory response and autophagy of fibroblast-like synoviocytes via miR-2682-5p/ADAMTS9 axis in rheumatoid arthritis. *Biosci Rep* 40(8):BSR20201273. <https://doi.org/10.1042/BSR20201273>
1053. Xing Y, Liu Y, Deng M, Wang HP, Abdul M, Zhang FF, Zhang Z, Cao JL (2021) The synergistic effects of opioid and neuropeptide B/W in rat acute inflammatory and neuropathic pain models. *Eur J Pharmacol* 898:173979. <https://doi.org/10.1016/j.ejphar.2021.173979>
1054. Harada M, Kamimura D, Arima Y, Kohsaka H, Nakatsuiji Y, Nishida M, Atsumi T, Meng J, Bando H, Singh R et al (2015) Temporal expression of growth factors triggered by epiregulin regulates inflammation development. *J Immunol* 194(3):1039–1046. <https://doi.org/10.4049/jimmunol.1400562>
1055. Xu D, Lian D, Wu J, Liu Y, Zhu M, Sun J, He D, Li L (2017) Brain-derived neurotrophic factor reduces inflammation and hippocampal apoptosis in experimental *Streptococcus pneumoniae* meningitis. *J Neuroinflamm* 14(1):156. <https://doi.org/10.1186/s12974-017-0930-6>
1056. Duan XL, Guo Z, He YT, Li YX, Liu YN, Bai HH, Li HL, Hu XD, Suo ZW (2020) SNAP25/syntaxin4/VAMP2/Munc18-1 complexes in spinal dorsal horn contributed to inflammatory pain. *Neuroscience* 429:203–212. <https://doi.org/10.1016/j.neuroscience.2020.01.003>
1057. Cortés J, Hidalgo J, Aguilera S, Castro I, Brito M, Urrea H, Pérez P, Barrera MJ, Carvajal P, Urzúa U et al (2019) Synaptotagmin-1 overexpression under inflammatory conditions affects secretion in salivary glands from Sjögren's syndrome patients. *J Autoimmun* 97:88–99. <https://doi.org/10.1016/j.jaut.2018.10.019>
1058. Sampath H, Ntambi JM (2011) The role of stearoyl-CoA desaturase in obesity, insulin resistance, and inflammation. *Ann NY Acad Sci* 1243:47–53. <https://doi.org/10.1111/j.1749-6632.2011.06303.x>
1059. Wang M, Wei J, Shang F, Zang K, Ji T (2019) Platelet-derived growth factor B attenuates lethal sepsis through inhibition of inflammatory responses. *Int Immunopharmacol* 75:105792. <https://doi.org/10.1016/j.intimp.2019.105792>
1060. Portugal LR, Fernandes LR, Pietra Pedroso VS, Santiago HC, Gazzinelli RT, Alvarez-Leite JI (2008) Influence of low-density lipoprotein (LDL) receptor on lipid composition, inflammation and parasitism during *Toxoplasma gondii* infection. *Microbes Infect* 10(3):276–284. <https://doi.org/10.1016/j.micinf.2007.12.001>
1061. Williams GP, Schonhoff AM, Jurkuvenaite A, Gallups NJ, Standaert DG, Harms AS (2021) CD4 T cells mediate brain inflammation and neurodegeneration in a mouse model of Parkinson's disease. *Brain* 144(7):2047–2059. <https://doi.org/10.1093/brain/awab103>
1062. Emgård J, Kammoun H, García-Cassani B, Chesné J, Parigi SM, Jacob JM, Cheng HW, Evren E, Das S, Czarnewski P et al (2018) Oxysterol sensing through the receptor GPR183 promotes the lymphoid-tissue-inducing function of innate lymphoid cells and colonic inflammation. *Immunity* 48(1):120–132.e8. <https://doi.org/10.1016/j.immuni.2017.11.020>
1063. Na YR, Jung D, Stakenborg M, Jang H, Gu GJ, Jeong MR, Suh SY, Kim HJ, Kwon YH, Sung TS et al (2021) Prostaglandin E2 receptor PTGER4-expressing macrophages promote intestinal epithelial barrier regeneration upon inflammation. *Gut* 70(12):2249–2260. <https://doi.org/10.1136/gutjnl-2020-322146>
1064. Sharma N, Drobinski P, Kaye A, Chen Z, Kjelgaard-Petersen CF, Gantzel T, Karsdal MA, Michaelis M, Ladell C, Bay-Jensen AC et al (2020) Inflammation and joint destruction may be linked to the generation of cartilage metabolites of ADAMTS-5 through activation of toll-like receptors. *Osteoarthritis Cartilage* 28(5):658–668. <https://doi.org/10.1016/j.joca.2019.11.002>
1065. Wizenty J, Müllerke S, Kolesnichenko M, Heuberger J, Lin M, Fischer AS, Mollenkopf HJ, Berger H, Tacke F, Sigal M (2022) Gastric stem cells promote inflammation and gland remodeling in response to *Helicobacter pylori* via Rspo3-Lgr4 axis. *EMBO J* 41(13):e109996. <https://doi.org/10.15252/embj.2021109996>
1066. Wang X, Niu L, Kang A, Pang Y, Zhang Y, Wang W, Zhang Y, Huang X, Liu Q, Geng Z et al (2022) Effects of ambient PM2.5 on development of psoriasisiform inflammation through KRT17-dependent activation of AKT/mTOR/HIF-1 α pathway. *Ecotoxicol Environ Saf* 243:114008. <https://doi.org/10.1016/j.ecoenv.2022.114008>
1067. Ng YH, Zhu H, Pallen CJ, Leung PC, MacCalman CD (2006) Differential effects of interleukin-1 β and transforming growth factor- β 1 on the expression of the inflammation-associated protein, ADAMTS-1, in human decidual stromal cells in vitro. *Hum Reprod* 21(8):1990–1999. <https://doi.org/10.1093/humrep/del108>
1068. Su Y, Ding J, Yang F, He C, Xu Y, Zhu X, Zhou H, Li H (2022) The regulatory role of PDE4B in the progression of inflammatory function study. *Front Pharmacol* 13:982130. <https://doi.org/10.3389/fphar.2022.982130>
1069. Uncini A, Notturmo F, Pace M, Caporale CM (2011) Polymorphism of CD1 and SH2D2A genes in inflammatory neuropathies. *J Peripher Nerv Syst* 16(Suppl 1):48–51. <https://doi.org/10.1111/j.1529-8027.2011.00307.x>
1070. Ong LK, Briggs GD, Guan L, Dunkley PR, Dickson PW (2021) Peripheral inflammation induces long-term changes in tyrosine hydroxylase activation in the substantia nigra. *Neurochem Int* 146:105022. <https://doi.org/10.1016/j.neuint.2021.105022>
1071. van Wageningen TA, Vlaar E, Kooij G, Jongenelen CAM, Geurts JGG, van Dam AM (2019) Regulation of microglial TMEM119 and P2RY12 immunoreactivity in multiple sclerosis white and grey matter lesions is dependent on their inflammatory environment. *Acta Neuropathol Commun* 7(1):206. <https://doi.org/10.1186/s40478-019-0850-z>

1072. Sirniö P, Tuomisto A, Tervahartiala T, Sorsa T, Klintrup K, Karhu T, Herzig KH, Mäkelä J, Karttunen TJ, Salo T et al (2018) High-serum MMP-8 levels are associated with decreased survival and systemic inflammation in colorectal cancer. *Br J Cancer* 119(2):213–219. <https://doi.org/10.1038/s41416-018-0136-4>
1073. Petkevicius K, Bidault G, Virtue S, Newland SA, Dale M, Dugourd A, Saez-Rodriguez J, Mallat Z, Vidal-Puig A (2021) Macrophage beta2-adrenergic receptor is dispensable for the adipose tissue inflammation and function. *Mol Metab* 48:101220. <https://doi.org/10.1016/j.molmet.2021.1012>
1074. Cebo M, Dittrich K, Fu X, Manke MC, Emschermann F, Rheinlaender J, von Eysmond H, Ferreiros N, Sudman J, Witte A et al (2022) Platelet ACKR3/CXCR7 favors antiplatelet lipids over an atherothrombotic lipodome and regulates thromboinflammation. *Blood* 139(11):1722–1742. <https://doi.org/10.1182/blood.2021013097>
1075. Klasić M, Markulin D, Vojta A, Samaržija I, Biruš I, Dobrinčić P, Ventham NT, Trbojević-Akmačić I, Šimurina M, Štambuk J et al (2018) Promoter methylation of the MGAT3 and BACH2 genes correlates with the composition of the immunoglobulin G glycome in inflammatory bowel disease. *Clin Epigenetics* 10:75. <https://doi.org/10.1186/s13148-018-0507-y>
1076. Yu B, Zeng A, Liu H, Yang Z, Fu M (2022) MiR-654-3p, reduced by the excessive ALKBH5, alleviated the inflammation in OA by targeting TNFRSF9, the trigger of the NF- κ B pathway. *Biochem Biophys Res Commun* 634:30–39. <https://doi.org/10.1016/j.bbrc.2022.09.103>
1077. Mihara S, Suzuki N (2007) Role of Txk, a member of the Tec family of tyrosine kinases, in immune-inflammatory diseases. *Int Rev Immunol* 26(5–6):333–348. <https://doi.org/10.1080/08830180701690835>
1078. Rasmussen LJH, Petersen JEV, Eugen-Olsen J (2021) Soluble urokinase plasminogen activator receptor (suPAR) as a biomarker of systemic chronic inflammation. *Front Immunol* 12:780641. <https://doi.org/10.3389/fimmu.2021.780641>
1079. Dong Y, Lan W, Wu W, Huang Z, Zhao J, Peng L, Wang J (2013) Increased expression of EphA7 in inflamed human dental pulp. *J Endod* 39(2):223–227. <https://doi.org/10.1016/j.joen.2012.11.020>
1080. Wu PB, Zhang Y, Nie G, Huang X, Yu YJ, Yin AN, Zhou R, He CP, Wang P (2021) Association between genetic variants in ZNF365 and inflammatory bowel disease risk in Caucasians: a meta-analysis and trial sequential analysis. *Expert Rev Clin Immunol* 17(8):915–921. <https://doi.org/10.1080/1744666X.2021.1939012>
1081. Huang S, Zhen Y, Yin X, Yang Z, Li X, Wang R, Wen H, Zhong H, Yan J, Sun Q (2023) KMT2C induced by FAP5P3 aggravates keratinocyte hyperproliferation and psoriasisiform skin inflammation by upregulating the transcription of PIK3R3. *J Invest Dermatol* 143(1):37–47.e8. <https://doi.org/10.1016/j.jid.2022.06.02>
1082. Hu Y, Wu L, Jiang L, Liang N, Zhu X, He Q, Qin H, Chen W (2021) Notoginsenoside R2 reduces A β 25–35-induced neuronal apoptosis and inflammation via miR-27a/SOX8/ β -catenin axis. *Hum Exp Toxicol* 40:S347–S358. <https://doi.org/10.1177/09603271211041996>
1083. Zheng Z, Wang X, Zheng Y, Wu H (2024) Enhanced expression of miR-204 attenuates LPS stimulated inflammatory injury through inhibiting the Wnt/ β -catenin pathway via targeting CCND2. *Int Immunopharmacol* 126:111334. <https://doi.org/10.1016/j.intimp.2023.111334>
1084. Plaza-Díaz J, Robles-Sánchez C, Abadía-Molina F, Morón-Calvente V, Sáez-Lara MJ, Ruiz-Bravo A, Jiménez-Valera M, Gil Á, Gómez-Llorente C, Fontana L (2017) Adamdec1, Ednrb and Ptgsl/Cox1, inflammation genes upregulated in the intestinal mucosa of obese rats, are down-regulated by three probiotic strains. *Sci Rep* 7(1):1939. <https://doi.org/10.1038/s41598-017-02203-3>
1085. Kayama H, Tani H, Kitada S, Opasawatchai A, Okumura R, Motooka D, Nakamura S, Takeda K (2019) BATF2 prevents T-cell-mediated intestinal inflammation through regulation of the IL-23/IL-17 pathway. *Int Immunol* 31(6):371–383. <https://doi.org/10.1093/intimm/dxz014>
1086. Yamawaki H, Futagami S, Sakasegawa N, Murakami M, Agawa S, Ikeda G, Noda H, Kirita K, Gudis K, Higuchi K et al (2020) Acotiamide attenuates central urocortin 2-induced intestinal inflammatory responses, and urocortin 2 treatment reduces TNF- α productions in LPS-stimulated macrophage cell lines. *Neurogastroenterol Motil* 32(8):e13813. <https://doi.org/10.1111/nmo.13813>
1087. Zhou C, Huang J, Chen J, Lai J, Zhu F, Xu X, Wang DW (2016) CYP2J2-derived EETs attenuated angiotensin II-induced adventitial remodeling via reduced inflammatory response. *Cell Physiol Biochem* 39(2):721–739. <https://doi.org/10.1159/000445663>
1088. Kim Y, Lee S, Zhang H, Lee S, Kim H, Kim Y, Won MH, Kim YM, Kwon YG (2020) CLEC14A deficiency exacerbates neuronal loss by increasing blood-brain barrier permeability and inflammation. *J Neuroinflamm* 17(1):48. <https://doi.org/10.1186/s12974-020-1727-6>
1089. Kyoreva M, Li Y, Hoosenally M, Hardman-Smart J, Morrison K, Tosi I, Tolaini M, Barinaga G, Stockinger B, Mrowietz U et al (2021) CYP1A1 enzymatic activity influences skin inflammation via regulation of the AHR pathway. *J Invest Dermatol* 141(6):1553–1563.e3. <https://doi.org/10.1016/j.jid.2020.11.024>
1090. Mentxaka A, Gómez-Ambrosi J, Ramírez B, Rodríguez A, Becerril S, Neira G, Valentí V, Moncada R, Silva C, Unamuno X et al (2022) Netrin-1 promotes visceral adipose tissue inflammation in obesity and is associated with insulin resistance. *Nutrients* 14(20):4372. <https://doi.org/10.3390/nu14204372>
1091. Verzola D, Milanese S, Viazzi F, Ansaldo F, Saio M, Garibaldi S, Carta A, Costigliolo F, Salvidio G, Barisione C et al (2020) Enhanced myostatin expression and signalling promote tubulointerstitial inflammation in diabetic nephropathy. *Sci Rep* 10(1):6343. <https://doi.org/10.1038/s41598-020-62875-2>
1092. Allbrand M, Eklund D, Cao Y, Nilsson K, Lodefalk M (2022) Gene expression of leptin, leptin receptor isoforms and inflammatory cytokines in placentas of obese women - Associations to birth weight and fetal sex. *Placenta* 117:64–71. <https://doi.org/10.1016/j.placenta.2021.10.002>
1093. Chang TY, Hong YK, Kuo YL, Wu HL, Shieh SJ (2024) CD248 regulates inflammation and encapsulation in silicone-related capsule formation. *Plast Reconstr Surg* 153(1):109–120. <https://doi.org/10.1097/PRS.00000000000010464>
1094. Kaito T, Morimoto T, Mori Y, Kanayama S, Makino T, Takenaka S, Sakai Y, Otsuru S, Yoshioka Y, Yoshikawa H (2018) BMP-2/7 heterodimer strongly induces bone regeneration in the absence of increased soft tissue inflammation. *Spine J* 18(1):139–146. <https://doi.org/10.1016/j.spinee.2017.07.171>
1095. Budylev A, Solar I, Kessner R, Aizic A (2022) ROS1-positive inflammatory myofibroblastic tumor of the small bowel causing obstruction: a case report. *J Radiol Case Rep* 16(1):14–21. <https://doi.org/10.3941/jrcr.v16i1.3928>
1096. Wang X, Song R, Li Z (2023) Salvicolone protects against high glucose-induced proliferation, oxidative stress, inflammation, and fibrosis of human renal mesangial cells by upregulating membrane metalloendopeptidase expression. *Chem Biol Drug Des* 101(4):819–828. <https://doi.org/10.1111/cbdd.14183>
1097. Li K, Ching D, Luk FS, Raffai RL (2015) Apolipoprotein E enhances microRNA-146a in monocytes and macrophages to suppress nuclear factor- κ B-driven inflammation and atherosclerosis. *Circ Res* 117(1):e1–e11. <https://doi.org/10.1161/CIRCRESAHA.117.305844>
1098. Ji H, Zhang X, Du Y, Liu H, Li S, Li L (2012) Polydatin modulates inflammation by decreasing NF- κ B activation and oxidative stress by increasing Gli1, Ptch1, SOD1 expression and ameliorates blood-brain barrier permeability for its neuroprotective effect in pMCAO rat brain. *Brain Res Bull* 87(1):50–59. <https://doi.org/10.1016/j.brainresbull.2011.09.021>
1099. Man SM, Karki R, Briard B, Burton A, Gingras S, Pelletier S, Kanneganti TD (2017) Differential roles of caspase-1 and caspase-11 in infection and inflammation. *Sci Rep* 7:45126. <https://doi.org/10.1038/srep45126>
1100. Guindon J, Gujjarro A, Piomelli D, Hohmann AG (2011) Peripheral antinociceptive effects of inhibitors of monoacylglycerol lipase in a rat model of inflammatory pain. *Br J Pharmacol* 163(7):1464–1478. <https://doi.org/10.1111/j.1476-5381.2010.01192.x>
1101. Zhao J, Yuan G, Cendan CM, Nassar MA, Lagerström MC, Kullander K, Gavazzi I, Wood JN (2010) Nociceptor-expressed ephrin-B2 regulates inflammatory and neuropathic pain. *Mol Pain* 6:77. <https://doi.org/10.1186/1744-8069-6-77>
1102. Zhang D, Ding Y, Wang X, Xin W, Du W, Chen W, Zhang X, Li P (2020) Effects of ABCG2 and SLCO1B1 gene variants on inflammation markers in patients with hypercholesterolemia and diabetes mellitus treated with rosuvastatin. *Eur J Clin Pharmacol* 76(7):939–946. <https://doi.org/10.1007/s00228-020-02882-4>
1103. Chen Y, Ku H, Zhao L, Wheeler DC, Li LC, Li Q, Varghese Z, Moorhead JF, Powis SH, Huang A et al (2014) Inflammatory stress induces statin resistance by disrupting 3-hydroxy-3-methylglutaryl-CoA reductase feedback regulation. *Arterioscler Thromb Vasc Biol* 34(2):365–376. <https://doi.org/10.1161/ATVBAHA.113.301301>

1104. Gaddam RR, Chambers S, Bhatia M (2014) ACE and ACE2 in inflammation: a tale of two enzymes. *Inflamm Allergy Drug Targets* 13(4):224–234. <https://doi.org/10.2174/1871528113666140713164506>
1105. Park CJ, Lin PC, Zhou S, Barakat R, Bashir ST, Choi JM, Cacioppo JA, Oakley OR, Duffy DM, Lydon JP et al (2020) Progesterone receptor serves the ovary as a trigger of ovulation and a terminator of inflammation. *Cell Rep* 31(2):107496. <https://doi.org/10.1016/j.celrep.2020.03.060>
1106. Basak M, Das K, Mahata T, Kumar D, Nagar N, Poluri KM, Kumar P, Das P, Stewart A, Maity B (2023) RGS7 balances acetylation/de-acetylation of p65 to control chemotherapy-dependent cardiac inflammation. *Cell Mol Life Sci* 80(9):255. <https://doi.org/10.1007/s00018-023-04895-5>
1107. Kumagai T, Kiwamoto S, Brummet ME, Wu F, Aoki K, Zhu Z, Bochner BS, Tiemeyer M (2018) Airway glycomic and allergic inflammatory consequences resulting from keratan sulfate galactose 6-O-sulfotransferase (CHST1) deficiency. *Glycobiology* 28(6):406–417. <https://doi.org/10.1093/glycob/cwy025>
1108. Shinozaki T, Watanabe H, Takagishi K, Pritzker KP (1998) Allotype immunoglobulin enhances alkaline phosphatase activity: implications for the inflammatory response. *J Lab Clin Med* 132(4):320–328. [https://doi.org/10.1016/s0022-2143\(98\)90046-4](https://doi.org/10.1016/s0022-2143(98)90046-4)
1109. Liu L, Chen M, Lin X, Xiang X, Yang J, Zheng Y, Xiong X, Zhu S (2021) TRPC6 attenuates cortical astrocytic apoptosis and inflammation in cerebral ischemic/reperfusion injury. *Front Cell Dev Biol* 8:594283. <https://doi.org/10.3389/fcell.2020.594283>
1110. Ashkar S, Mesentsev A, Zhang WX, Mastuyugin V, Dunn MW, Laniado-Schwartzman M (2004) Retinoic acid induces corneal epithelial CYP4B1 gene expression and stimulates the synthesis of inflammatory 12-hydroxyeicosanoids. *J Ocul Pharmacol Ther* 20(1):65–74. <https://doi.org/10.1089/108076804772745473>
1111. Tocci S, Ibeawuchi SR, Das S, Sayed IM (2022) Role of ELMO1 in inflammation and cancer-clinical implications. *Cell Oncol (Dordr)* 45(4):505–525. <https://doi.org/10.1007/s13402-022-00680-x>
1112. Aslamiy A, Oh E, Olson EM, Zhang J, Ahn M, Moin ASM, Tunduguru R, Salunkhe VA, Veluthakal R, Thurmond DC (2018) Doc2b protects β -cells against inflammatory damage and enhances function. *Diabetes* 67(7):1332–1344. <https://doi.org/10.2337/db17-1352>
1113. González-Domínguez É, Samaniego R, Flores-Sevilla JL, Campos-Campos SF, Gómez-Campos G, Salas A, Campos-Peña V, Corbí AL, Sánchez-Mateos P, Sánchez-Torres C (2015) CD163L1 and CLEC5A discriminate subsets of human resident and inflammatory macrophages in vivo. *J Leukoc Biol* 98(4):453–466. <https://doi.org/10.1189/jlb.3H1114-531R>
1114. Nakanishi T, Sakiyama S, Takashima H, Honda R, Shumba MN, Nakamura Y, Kasahara K, Tamai I (2020) Toxicological implication of prostaglandin transporter SLC02A1 inhibition by cigarette smoke in exacerbation of lung inflammation. *Toxicol Appl Pharmacol* 405:115201. <https://doi.org/10.1016/j.taap.2020.115201>
1115. Bouzid D, Amouri A, Fourati H, Marques I, Abida O, Tahrir N, Gonçalves CP, Masmoudi H (2013) Polymorphisms in the IL2RA and IL2RB genes in inflammatory bowel disease risk. *Genet Test Mol Biomarkers* 17(11):833–839. <https://doi.org/10.1089/gtmb.2013.0291>
1116. Xu R, Singhal N, Serinagaoglu Y, Chandrasekharan K, Joshi M, Bauer JA, Janssen PM, Martin PT (2015) Deletion of Galgt2 (B4Galnt2) reduces muscle growth in response to acute injury and increases muscle inflammation and pathology in dystrophin-deficient mice. *Am J Pathol* 185(10):2668–2684. <https://doi.org/10.1016/j.ajpath.2015.06.008>
1117. Wang Z, Zhao Q, Nie Y, Yu Y, Misra BB, Zabalawi M, Chou JW, Key CC, Molina AJ, Quinn MA et al (2020) Solute carrier family 37 member 2 (SLC37A2) negatively regulates murine macrophage inflammation by controlling glycolysis. *iScience* 23(5):101125. <https://doi.org/10.1016/j.isci.2020.101125>
1118. Yuan S, Liu H, Yuan D, Xu J, Chen Y, Xu X, Xu F, Liang H (2020) PNPLA3 I148M mediates the regulatory effect of NF- κ B on inflammation in PA-treated HepG2 cells. *J Cell Mol Med* 24(2):1541–1552. <https://doi.org/10.1111/jcmm.14839>
1119. Vaitinen M, Lankinen MA, Käkälä P, Ågren J, Wheelock CE, Laakso M, Schwab U, Pihlajamäki J (2022) The FADS1 genotypes modify the effect of linoleic acid-enriched diet on adipose tissue inflammation via pro-inflammatory eicosanoid metabolism. *Eur J Nutr* 61(7):3707–3718. <https://doi.org/10.1007/s00394-022-02922-y>
1120. Castro-Santos P, Moro-García MA, Marcos-Fernández R, Alonso-Arias R, Díaz-Peña R (2017) ERAP1 and HLA-C interaction in inflammatory bowel disease in the Spanish population. *Innate Immun* 23(5):476–481. <https://doi.org/10.1177/1753425917716527>
1121. Strelakova T, Svirin E, Veniaminova E, Kopeikina E, Veremeyko T, Yung AWY, Proshin A, Walitza S, Anthony DC, Lim LW et al (2021) ASD-like behaviors, a dysregulated inflammatory response and decreased expression of PLP1 characterize mice deficient for sialyltransferase ST3GAL5. *Brain Behav Immun Health* 16:100306. <https://doi.org/10.1016/j.bbih.2021.100306>
1122. Ambartsumian N, Klingelhöfer J, Grigorian M (2019) The multifaceted S100A4 protein in cancer and inflammation. *Methods Mol Biol* 1929:339–365. https://doi.org/10.1007/978-1-4939-9030-6_22
1123. Kobelt D, Zhang C, Clayton-Lucey IA, Glauben R, Voss C, Siegmund B, Stein U (2020) Pro-inflammatory TNF- α and IFN- γ promote tumor growth and metastasis via induction of MACC1. *Front Immunol* 11:980. <https://doi.org/10.3389/fimmu.2020.00980>
1124. Barriga M, Benitez R, Ferraz-de-Paula V, Garcia-Frutos M, Caro M, Robledo G, O'Valle F, Campos-Salinas J, Delgado M (2021) Protective role of cortistatin in pulmonary inflammation and fibrosis. *Br J Pharmacol* 178(21):4368–4388. <https://doi.org/10.1111/bph.15615>
1125. Van Hove I, Lefevère E, De Groef L, Sergeys J, Salinas-Navarro M, Libert C, Vandenbroucke R, Moons L (2016) MMP-3 deficiency alleviates endotoxin-induced acute inflammation in the posterior eye segment. *Int J Mol Sci* 17(11):1825. <https://doi.org/10.3390/ijms17111825>
1126. Zhang DK, He FQ, Li TK, Pang XH, Cui DJ, Xie Q, Huang XL, Gan HT (2010) Glial-derived neurotrophic factor regulates intestinal epithelial barrier function and inflammation and is therapeutic for murine colitis. *J Pathol* 222(2):213–222. <https://doi.org/10.1002/path.2749>
1127. Olsavszky V, Ulbrich F, Singh S, Dieltm M, Sticht C, Schmid CD, Zierow J, Wohlfeil SA, Schledzewski K, Dooley S et al (2017) GATA4 and LMO3 balance angiocrine signaling and autocrine inflammatory activation by BMP2 in liver sinusoidal endothelial cells. *Gene* 627:491–499. <https://doi.org/10.1016/j.gene.2017.06.051>
1128. Delmont E, Manso C, Querol L, Cortese A, Berardinelli A, Lozza A, Belghazi M, Malissart P, Labauge P, Taieb G et al (2017) Autoantibodies to nodal isoforms of neurofascin in chronic inflammatory demyelinating polyneuropathy. *Brain* 140(7):1851–1858. <https://doi.org/10.1093/brain/awx124>
1129. Thakur M, Rho O, Khandelwal A, Nathan CO, DiGiovanni J (2024) Inducible keratinocyte specific FGFR2 deficiency inhibits UVB-induced signaling, proliferation, inflammation, and skin carcinogenesis. *J Invest Dermatol* 144(2):341–350.e7. <https://doi.org/10.1016/j.jid.2023.08.013>
1130. Chan BCL, Lam CWK, Tam LS, Wong CK (2019) IL33: roles in allergic inflammation and therapeutic perspectives. *Front Immunol* 10:364. <https://doi.org/10.3389/fimmu.2019.00364>
1131. Villoslada P, Genain CP (2004) Role of nerve growth factor and other trophic factors in brain inflammation. *Prog Brain Res* 146:403–414. [https://doi.org/10.1016/S0079-6123\(03\)46025-1](https://doi.org/10.1016/S0079-6123(03)46025-1)
1132. Chen Y, Wang B, Chen Y, Wu Q, Lai WF, Wei L, Nandakumar KS, Liu D (2022) HAPLN1 affects cell viability and promotes the pro-inflammatory phenotype of fibroblast-like synoviocytes. *Front Immunol* 13:888612. <https://doi.org/10.3389/fimmu.2022.888612>
1133. Cui H, Zhang J, Li Z, Chen F, Cui H, Du X, Liu H, Wang J, Diwan AD, Zheng Z (2021) Growth differentiation factor-6 attenuates inflammatory and pain-related factors and degenerated disc-induced pain behaviors in rat model. *J Orthop Res* 39(5):959–970. <https://doi.org/10.1002/jor.24793>
1134. Yang L, Li T, Zha L (2020) Foxc2 alleviates Ox-LDL-induced lipid accumulation, inflammation, and apoptosis of macrophage via regulating the expression of Angptl2. *Inflammation* 43(4):1397–1410. <https://doi.org/10.1007/s10753-020-01217-w>
1135. Zivanović S, Rackov LP, Zivanović A, Jevtić M, Nikolić S, Kocić S (2011) Cartilage oligomeric matrix protein - inflammation biomarker in knee osteoarthritis. *Bosn J Basic Med Sci* 11(1):27–32. <https://doi.org/10.17305/bjbm.2011.2619>
1136. Tang R, Harasymowicz NS, Wu CL, Collins KH, Choi YR, Oswald SJ, Guilak F (2020) Gene therapy for follistatin mitigates systemic metabolic inflammation and post-traumatic arthritis in high-fat diet-induced obesity. *Sci Adv* 6(19):eaaz7492. <https://doi.org/10.1126/sciadv.aaz7492>
1137. Ba MA, Aiyuk A, Hernández K, Evasovic JM, Wuebbles RD, Burkin DJ, Singer CA (2022) Transgenic overexpression of $\alpha 7$ integrin in smooth muscle attenuates allergen-induced airway inflammation in a murine

- model of asthma. *FASEB Bioadv* 4(11):724–740. <https://doi.org/10.1096/fba.2022-00050>
1138. Sosnovski KE, Braun T, Amir A, Moshel D, BenShoshan M, VanDussen KL, Levhar N, Abbas-Egbariya H, Beider K, Ben-Yishay R et al (2023) GATA6-AS1 regulates intestinal epithelial mitochondrial functions, and its reduced expression is linked to intestinal inflammation and less favourable disease course in ulcerative colitis. *J Crohns Colitis* 17(6):960–971. <https://doi.org/10.1093/ecco-jcc/jjad006>
 1139. Fraser A, Fearon U, Billingham RC, Ionescu M, Reece R, Barwick T, Emery P, Poole AR, Veale DJ (2003) Turnover of type II collagen and aggrecan in cartilage matrix at the onset of inflammatory arthritis in humans: relationship to mediators of systemic and local inflammation. *Arthritis Rheum* 48(11):3085–3095. <https://doi.org/10.1002/art.11331>
 1140. Ouyang Y, Tang Y, Fu L, Peng S, Wu W, Tan D, Fu X (2020) Exosomes secreted by chronic hepatitis B patients with PNALT and liver inflammation grade \geq A2 promoted the progression of liver cancer by transferring miR-25-3p to inhibit the co-expression of TCF21 and HHIP. *Cell Prolif* 53(7):e12833. <https://doi.org/10.1111/cpr.12833>
 1141. Martín-Vázquez E, Cobo-Vuilleumier N, López-Noriega L, Lorenzo PI, Gauthier BR (2023) The PTGS2/COX2-PGE2 signaling cascade in inflammation: Pro or anti? A case study with type 1 diabetes mellitus. *Int J Biol Sci* 19(13):4157–4165. <https://doi.org/10.7150/ijbs.86492>
 1142. Chen JX, Wang YP, Zhang X, Li GX, Zheng K, Duan CZ (2020) lncRNA Mtss1 promotes inflammatory responses and secondary brain injury after intracerebral hemorrhage by targeting miR-709 in mice. *Brain Res Bull* 162:20–29. <https://doi.org/10.1016/j.brainresbull.2020.04.017>
 1143. Zolfaghari R, Chen Q, Ross AC (2012) DHRS3, a retinal reductase, is differentially regulated by retinoic acid and lipopolysaccharide-induced inflammation in THP-1 cells and rat liver. *Am J Physiol Gastrointest Liver Physiol* 303(5):G578–G588. <https://doi.org/10.1152/ajpgi.00234.2012>
 1144. Le Magnen C, Virk RK, Dutta A, Kim JY, Panja S, Lopez-Bujanda ZA, Califano A, Drake CG, Mitrofanova A, Abate-Shen C (2018) Cooperation of loss of NKX3.1 and inflammation in prostate cancer initiation. *Dis Model Mech* 11(11):dmm035139. <https://doi.org/10.1242/dmm.035139>
 1145. Burgaletto C, Platania CBM, Di Benedetto G, Munafò A, Giurdanella G, Federico C, Caltabiano R, Saccone S, Conti F, Bernardini R et al (2021) Targeting the miRNA-155/TNFSF10 network restrains inflammatory response in the retina in a mouse model of Alzheimer's disease. *Cell Death Dis* 12(10):905. <https://doi.org/10.1038/s41419-021-04165-x>
 1146. Carniel BP, da Rocha NS (2021) Brain-derived neurotrophic factor (BDNF) and inflammatory markers: Perspectives for the management of depression. *Prog Neuropsychopharmacol Biol Psych* 108:110151. <https://doi.org/10.1016/j.pnpb.2020.110151>
 1147. Xu J, Ji T, Li G, Zhang H, Zheng Y, Li M, Ma J, Li Y, Chi G (2022) Lactate attenuates astrocytic inflammation by inhibiting ubiquitination and degradation of NDRG2 under oxygen-glucose deprivation conditions. *J Neuroinflamm* 19(1):314. <https://doi.org/10.1186/s12974-022-02678-6>
 1148. Xu LJ, Wang HN, Zhou H, Li SY, Li F, Miao Y, Lei B, Sun XH, Gao F, Wang Z (2023) EphA4/ephrinA3 reverse signaling induced Müller cell gliosis and production of pro-inflammatory cytokines in experimental glaucoma. *Brain Res* 1801:148204. <https://doi.org/10.1016/j.brainres.2022.148204>
 1149. Koganesawa M, Dwyer DF, Alhallak K, Nagai J, Zaleski K, Samuchiwal S, Hiroaki H, Nishida A, Hirsch TI, Brennan PJ et al (2024) Pla2g5 contributes to viral-like-induced lung inflammation through macrophage proliferation and LA/Ffar1 lung cell recruitment. *Immunology* 172(1):144–162. <https://doi.org/10.1111/imm.13766>
 1150. Chen Y, Ying Y, Wang M, Ma C, Jia M, Shi L, Wang S, Zheng X, Chen W, Shu XS (2023) A distal super-enhancer activates oncogenic ETS2 via recruiting MECOM in inflammatory bowel disease and colorectal cancer. *Cell Death Dis* 14(1):8. <https://doi.org/10.1038/s41419-022-05513-1>
 1151. Yamamoto H, Yoshida N, Kihara S (2022) Esaxerenone blocks vascular endothelial inflammation through SGK1. *J Cardiovasc Pharmacol* 80(4):583–591. <https://doi.org/10.1097/FJC.0000000000001316>
 1152. Wu Y, Quan Y, Liu Y, Liu K, Li H, Jiang Z, Zhang T, Lei H, Radek KA, Li D et al (2016) Hyperglycaemia inhibits REG3A expression to exacerbate TLR3-mediated skin inflammation in diabetes. *Nat Commun* 7:13393. <https://doi.org/10.1038/ncomms13393>
 1153. Zuo Y, He Z, Chen Y, Dai L (2023) Dual role of ANGPL4 in inflammation. *Inflamm Res* 72(6):1303–1313. <https://doi.org/10.1007/s00011-023-01753-9>
 1154. Ernst AS, Böhler LI, Hagenston AM, Hoffmann A, Heiland S, Sticht C, Bendzus M, Hecker M, Bading H, Marti HH et al (2019) EphB2-dependent signaling promotes neuronal excitotoxicity and inflammation in the acute phase of ischemic stroke. *Acta Neuropathol Commun* 7(1):15. <https://doi.org/10.1186/s40478-019-0669-7>
 1155. Chen B, Sheng D, Wang C, Liu W, Hu A, Xiao X, Gajendran B, Gao J, Hu J, Sample KM et al (2022) FL11 regulates inflammation-associated genes to accelerate leukemogenesis. *Cell Signal* 92:110269. <https://doi.org/10.1016/j.cellsig.2022.110269>
 1156. Hassan HM, Liang X, Xin J, Lu Y, Cai Q, Shi D, Ren K, Li J, Chen Q, Li J et al (2024) Thrombospondin 1 enhances systemic inflammation and disease severity in acute-on-chronic liver failure. *BMC Med* 22(1):95. <https://doi.org/10.1186/s12916-024-03318-x>
 1157. D'Aversa TG, Eugenin EA, Lopez L, Berman JW (2013) Myelin basic protein induces inflammatory mediators from primary human endothelial cells and blood-brain barrier disruption: implications for the pathogenesis of multiple sclerosis. *Neuropathol Appl Neurobiol* 39(3):270–283. <https://doi.org/10.1111/j.1365-2990.2012.01279.x>
 1158. Misiou A, Garmey JC, Hensien JM, Harmon DB, Osinski V, McSkimming C, Marshall MA, Fischer JW, Grandoch M, McNamara CA (2021) Helix-loop-helix factor Id3 (inhibitor of differentiation 3): a novel regulator of hyaluronan-mediated adipose tissue inflammation. *Arterioscler Thromb Vasc Biol* 41(2):796–807. <https://doi.org/10.1161/ATVBAHA.120.315588>
 1159. Gray SP, Di Marco E, Kennedy K, Chew P, Okabe J, El-Osta A, Calkin AC, Biessen EA, Touyz RM, Cooper ME et al (2016) Reactive oxygen species can provide atheroprotection via NOX4-dependent inhibition of inflammation and vascular remodeling. *Arterioscler Thromb Vasc Biol* 36(2):295–307. <https://doi.org/10.1161/ATVBAHA.115.307012>
 1160. Taylor Meadows KR, Steinberg MW, Clemons B, Stokes ME, Opitck GJ, Peach R, Scott FL (2018) Ozanimod (RPC1063), a selective S1PR1 and S1PR5 modulator, reduces chronic inflammation and alleviates kidney pathology in murine systemic lupus erythematosus. *PLoS ONE* 13(4):e0193236. <https://doi.org/10.1371/journal.pone.0193236>
 1161. Sun Y, Lin S, Wang H, Wang L, Qiu Y, Zhang F, Hao N, Wang F, Tan W (2024) Regulatory role of PL16 in autoimmune arthritis and intestinal inflammation: implications for Treg cell differentiation and function. *J Transl Med* 22(1):327. <https://doi.org/10.1186/s12967-024-05082-1>
 1162. Mao Y, Jiang F, Xu XJ, Zhou LB, Jin R, Zhuang LL, Juan CX, Zhou GP (2023) Inhibition of IGF2BP1 attenuates renal injury and inflammation by alleviating m6A modifications and E2F1/MLF pathway. *Int J Biol Sci* 19(2):593–609. <https://doi.org/10.7150/ijbs.78348>
 1163. Zhao B, Wang Y, Tan X, Ke K, Zheng X, Wang F, Lan S, Liao N, Cai Z, Shi Y et al (2019) Inflammatory micro-environment contributes to stemness properties and metastatic potential of HCC via the NF- κ B/miR-497/SALL4 axis. *Mol Ther Oncolytics* 15:79–90. <https://doi.org/10.1016/j.omto.2019.08.009>
 1164. Nguyen A, Tao H, Metrione M, Hajri T (2014) Very low density lipoprotein receptor (VLDLR) expression is a determinant factor in adipose tissue inflammation and adipocyte-macrophage interaction. *J Biol Chem* 289(3):1688–1703. <https://doi.org/10.1074/jbc.M113.515320>
 1165. Garg M, Royce SG, Tikellis C, Shallue C, Sluka P, Wardan H, Hosking P, Monagle S, Thomas M, Lubel JS et al (2019) The intestinal vitamin D receptor in inflammatory bowel disease: inverse correlation with inflammation but no relationship with circulating vitamin D status. *Therap Adv Gastroenterol* 12:1756284818822566. <https://doi.org/10.1177/1756284818822566>
 1166. Sriwattanapong K, Theerapanon T, Khamwachirapitak C, Sae-Ear P, Sa-Ard-Iam N, Shotelersuk V, Porntaveetus T (2024) In-depth investigation of FAM20A insufficiency effects on deciduous dental pulp cells: Altered behaviours, osteogenic differentiation, and inflammatory gene expression. *Int Endod J*. <https://doi.org/10.1111/iej.14056>
 1167. Kowalski EA, Chen J, Hazy A, Fritsch LE, Gudenschwager-Basso EK, Chen M, Wang X, Qian Y, Zhou M et al (2019) Peripheral loss of EphA4 ameliorates TBI-induced neuroinflammation and tissue damage. *J Neuroinflamm* 16(1):210. <https://doi.org/10.1186/s12974-019-1605-2>
 1168. Liu XH, Bauman WA, Cardozo C (2015) ANKRD1 modulates inflammatory responses in C2C12 myoblasts through feedback inhibition of NF- κ B signaling activity. *Biochem Biophys Res Commun* 464(1):208–213. <https://doi.org/10.1016/j.bbrc.2015.06.118>
 1169. Raffaghello L, Principi E, Baratto S, Panicucci C, Pintus S, Antonini F, Del Zotto G, Benzi A, Bruzzone S, Studieri P et al (2022) P2X7 receptor

- antagonist reduces fibrosis and inflammation in a mouse model of alpha-sarcoglycan muscular dystrophy. *Pharmaceuticals* (Basel) 15(1):89. <https://doi.org/10.3390/ph15010089>
1170. Scharf GM, Kilian K, Cordero J, Wang Y, Grund A, Hofmann M, Froese N, Wang X, Kispert A, Kist R et al (2019) Inactivation of Sox9 in fibroblasts reduces cardiac fibrosis and inflammation. *JCI Insight* 5(15):e126721. <https://doi.org/10.1172/jci.insight.12672>
 1171. Brnic D, Martinovic D, Zivkovic PM, Tokic D, Vilovic M, Rusic D, Tadin Hadjina I, Libers C, Glumac S, Supe-Domic D et al (2020) Inactive matrix Gla protein is elevated in patients with inflammatory bowel disease. *World J Gastroenterol* 26(32):4866–4877. <https://doi.org/10.3748/wjg.v26.i32.486>
 1172. Chen XQ, Mao JY, Wang CS, Li WB, Han TT, Lv K, Li JN (2022) CYP24A1 involvement in inflammatory factor regulation occurs via the wnt signaling pathway. *Curr Med Sci* 42(5):1022–1032. <https://doi.org/10.1007/s11596-022-2564-x>
 1173. Kim J, Durai P, Jeon D, Jung ID, Lee SJ, Park YM, Kim Y (2018) Phloretin as a potent natural TLR2/1 inhibitor suppresses TLR2-induced inflammation. *Nutrients* 10(7):868. <https://doi.org/10.3390/nu1007086>
 1174. Hirano T (2021) IL-6 in inflammation, autoimmunity and cancer. *Int Immunol* 33(3):127–148. <https://doi.org/10.1093/intimm/dxaa078>
 1175. Owlett LD, Karaahmet B, Le L, Belcher EK, Dionisio-Santos D, Olschowka JA, Elliott MR, O'Banion MK (2022) Gas6 induces inflammation and reduces plaque burden but worsens behavior in a sex-dependent manner in the APP/PS1 model of Alzheimer's disease. *J Neuroinflamm* 19(1):38. <https://doi.org/10.1186/s12974-022-02397-y>
 1176. Han Q, Zhang Z, He X, Chen M, Pang X, Chen C, Du T, Zhang H (2023) Primary inflammatory myofibroblastic tumour of the liver: a clinicopathological and genetic study including a subset with ETV6::NTRK3 fusion. *Histopathology* 82(6):925–936. <https://doi.org/10.1111/his.14881>
 1177. Liu Y, Bockermann R, Hadi M, Safari I, Carrion B, Kveiborg M, Issazadeh-Navikas S (2021) ADAM12 is a costimulatory molecule that determines Th1 cell fate and mediates tissue inflammation. *Cell Mol Immunol* 18(8):1904–1919. <https://doi.org/10.1038/s41423-020-0486-8>
 1178. Pansieri J, Ostojić L, Iashchishyn IA, Magzoub M, Wallin C, Wärmländer SKTS, Gräslund A, Nguyen Ngoc M, Smirnovas V, Svedružić Ž et al (2019) Pro-inflammatory S100A9 protein aggregation promoted by NCAM1 peptide constructs. *ACS Chem Biol* 14(7):1410–1417. <https://doi.org/10.1021/acscchembio.9b00394>
 1179. Itakura T, Peters DM, Fini ME (2015) Glaucomatous MYOC mutations activate the IL-1/NF-κB inflammatory stress response and the glaucoma marker SELE in trabecular meshwork cells. *Mol Vis* 21:1071–1084
 1180. Liu X, Zhang G, Liu L, Xiong G, Liu J, Wei W (2024) USP2 promotes the proliferation and inflammation of fibroblast-like synovial cells in rheumatoid arthritis through deubiquitination of TRAF2. *Biochem Genet*. <https://doi.org/10.1007/s10528-024-10737-1>
 1181. Shi J, Zhou LR, Wang XS, Du JF, Jiang MM, Song Z, Han GC, Mai ZT (2018) KLF2 attenuates bleomycin-induced pulmonary fibrosis and inflammation with regulation of AP-1. *Biochem Biophys Res Commun* 495(1):20–26. <https://doi.org/10.1016/j.bbrc.2017.10.114>
 1182. Babbe H, Sundberg TB, Tichenor M, Seierstad M, Bacani G, Berstler J, Chai W, Chang L, Chung M, Coe K et al (2024) Identification of highly selective SIK1/2 inhibitors that modulate innate immune activation and suppress intestinal inflammation. *Proc Natl Acad Sci USA* 121(1):e2307086120. <https://doi.org/10.1073/pnas.2307086120>
 1183. Zhan H, Chen H, Tang Z, Liu S, Xie K, Wang H (2022) SIX1 attenuates inflammation and rheumatoid arthritis by silencing MyD88-dependent TLR1/2 signaling. *Int Immunopharmacol* 106:108613. <https://doi.org/10.1016/j.intimp.2022.108613>
 1184. Moura J, Sørensen A, Leal EC, Svendsen R, Carvalho L, Willemoes RJ, Jørgensen PT, Jenssen H, Wengel J, Dalgaard LT et al (2019) microRNA-155 inhibition restores Fibroblast Growth Factor 7 expression in diabetic skin and decreases wound inflammation. *Sci Rep* 9(1):5836. <https://doi.org/10.1038/s41598-019-42309-4>
 1185. Rayego-Mateos S, Marquez-Exposito L, Basantes P, Tejedor-Santamaria L, Sanz AB, Nguyen TQ, Goldschmieding R, Ortiz A, Ruiz-Ortega M (2023) CCN2 activates RIPK3, NLRP3 inflammasomes, and Nrf2/oxidative pathways linked to kidney inflammation. *Antioxidants* (Basel) 12(8):1541. <https://doi.org/10.3390/antiox12081541>
 1186. Lohr J, Grotevendt A, Nauck M, Völzke H, Wallaschofski H, Friedrich N (2014) Relation of insulin-like growth factor-I and IGF binding protein 3 with markers of inflammation: results of a population-based study. *Clin Endocrinol* (Oxf) 80(1):148–154. <https://doi.org/10.1111/cen.12241>
 1187. Johnson LA, Prevo R, Clasper S, Jackson DG (2007) Inflammation-induced uptake and degradation of the lymphatic endothelial hyaluronan receptor LYVE-1. *J Biol Chem* 282(46):33671–33680. <https://doi.org/10.1074/jbc.M702889200>
 1188. Schönfelder J, Seibold T, Morawe M, Sroka R, Schneider N, Cai J, Golomejic J, Schütte L, Armacki M, Huber-Lang M et al (2023) Endothelial Protein kinase D1 is a major regulator of post-traumatic hyperinflammation. *Front Immunol* 14:1093022. <https://doi.org/10.3389/fimmu.2023.1093022>
 1189. Vaxevanis CK, Bauer M, Subbarayan K, Friedrich M, Massa C, Biehler K, Al-Ali HK, Wickenhauser C, Seliger B (2022) Biglycan as a mediator of proinflammatory response and target for MDS and sAAML therapy. *Oncimmunology* 12(1):2152998. <https://doi.org/10.1080/2162402X.2022.2152998>
 1190. Wang L, Zheng J, Pathak JL, Chen Y, Liang D, Yang L, Sun H, Zhong M, Wu L, Li L et al (2020) SLIT2 overexpression in periodontitis intensifies inflammation and alveolar bone loss, possibly via the activation of MAPK pathway. *Front Cell Dev Biol* 8:593. <https://doi.org/10.3389/fcell.2020.00593>
 1191. Yao J, Wu D, Zhang C, Yan T, Zhao Y, Shen H, Xue K, Huang X, Wang Z, Qiu Y (2021) Macrophage IRX3 promotes diet-induced obesity and metabolic inflammation. *Nat Immunol* 22(10):1268–1279. <https://doi.org/10.1038/s41590-021-01023-y>
 1192. Frade AF, Teixeira PC, Ianni BM, Pissetti CW, Saba B, Wang LH, Kuramoto A, Nogueira LG, Buck P, Dias F et al (2013) Polymorphism in the alpha cardiac muscle actin 1 gene is associated to susceptibility to chronic inflammatory cardiomyopathy. *PLoS ONE* 8(12):e83446. <https://doi.org/10.1371/journal.pone.0083446>
 1193. Mócsai G, Gáspár K, Nagy G, Irinyi B, Kapitány A, Bíró T, Gyimesi E, Tóth B, Maródi L, Szegedi A (2014) Severe skin inflammation and filaggrin mutation similarly alter the skin barrier in patients with atopic dermatitis. *Br J Dermatol* 170(3):617–624. <https://doi.org/10.1111/bjd.12743>
 1194. Zhou W, Wang F, Qian X, Luo S, Wang Z, Gao X, Kong X, Zhang J, Chen S (2023) Quercetin protects endothelial function from inflammation induced by localized disturbed flow by inhibiting NRP2-VEGFC complex. *Int Immunopharmacol* 116:109842. <https://doi.org/10.1016/j.intimp.2023.109842>
 1195. Liu A, Guo M, He L, Martínez MA, Martínez M, Lopez-Torres B, Martínez-Larrañaga MR, Wang X, Anadón A, Ares I (2022) Nicotinamide N-methyltransferase protects against deoxynivalenol-induced growth inhibition by suppressing pro-inflammatory cytokine expression. *Food Chem Toxicol* 163:112969. <https://doi.org/10.1016/j.fct.2022.112969>
 1196. Song Y, Hao D, Jiang H, Huang M, Du Q, Lin Y, Liu F, Chen B (2021) Nrf2 regulates CHI3L1 to suppress inflammation and improve post-traumatic osteoarthritis. *J Inflamm Res* 14:4079–4088. <https://doi.org/10.2147/JIR.S310831>
 1197. Zezulín AU, Yen D, Ye D, Howell ED, Bresciani E, Diemer J, Ren JG, Ahmad MH, Castilla LH, Touw IP et al (2023) RUNX1 is required in granulocyte-monocyte progenitors to attenuate inflammatory cytokine production by neutrophils. *Genes Dev* 37(13–14):605–620. <https://doi.org/10.1101/gad.350418.123>
 1198. Sun H, Li N, Tan J, Li H, Zhang J, Qu L, Lamont SJ (2022) Transcriptional regulation of RIP2 gene by NFIB is associated with cellular immune and inflammatory response to APEC infection. *Int J Mol Sci* 23(7):3814. <https://doi.org/10.3390/ijms23073814>
 1199. Wang X, Zhang L, Li P, Zheng Y, Yang Y, Ji S (2022) Apelin/APJ system in inflammation. *Int Immunopharmacol* 109:108822. <https://doi.org/10.1016/j.intimp.2022.108822>
 1200. Strelakova T, Svirin E, Veniaminova E, Kopeikina E, Veremeyko T, Yung AWY, Proshin A, Walitza S, Anthony DC, Lim LW et al (2021) ASD-like behaviors, a dysregulated inflammatory response and decreased expression of PLP1 characterize mice deficient for sialyltransferase ST3GAL5. *Brain Behav Immun Health* 16:100306. <https://doi.org/10.1016/j.bbih.2021.100306>
 1201. Wang P, Wang Y, Ma B (2023) Long noncoding RNA NAV2-AS5 relieves chondrocyte inflammation by targeting miR-8082/TNIP2 in

- osteoarthritis. *Cell Cycle* 22(7):796–807. <https://doi.org/10.1080/15384101.2022.2154554>
1202. Canalis E, Yu J, Singh V, Mocarska M, Schilling L (2023) NOTCH2 sensitizes the chondrocyte to the inflammatory response of tumor necrosis factor α . *J Biol Chem* 299(12):105372. <https://doi.org/10.1016/j.jbc.2023.105372>
 1203. Oura H, Bertoncini J, Velasco P, Brown LF, Carmeliet P, Detmar M (2003) A critical role of placental growth factor in the induction of inflammation and edema formation. *Blood* 101(2):560–567. <https://doi.org/10.1182/blood-2002-05-1516>
 1204. Zhao F, Dong J, Guo J, Bi L (2020) Inhibiting role of long non-coding RNA LINC01197 in inflammation in rheumatoid arthritis through the microRNA-150/THBS2 axis. *Exp Cell Res* 394(2):112136. <https://doi.org/10.1016/j.jcyexr.2020.112136>
 1205. Gao YZ, Wu XM, Zhou ZQ, Liu PM, Yang JJ, Ji MH (2023) Dysfunction of NRG1/ErbB4 signaling in the hippocampus might mediate long-term memory decline after systemic inflammation. *Mol Neurobiol* 60(6):3210–3226. <https://doi.org/10.1007/s12035-023-03278-y>
 1206. Nguyen HN, Noss EH, Mizoguchi F, Huppertz C, Wei KS, Watts GFM, Brenner MB (2017) Autocrine loop involving IL-6 family member LIF, LIF Receptor, and STAT4 drives sustained fibroblast production of inflammatory mediators. *Immunity* 46(2):220–232. <https://doi.org/10.1016/j.immuni.2017.01.004>
 1207. Zou HH, Yang PP, Huang TL, Zheng XX, Xu GS (2017) PLK2 plays an essential role in high D-glucose-induced apoptosis, ROS generation and inflammation in podocytes. *Sci Rep* 7(1):4261. <https://doi.org/10.1038/s41598-017-00686-8>
 1208. Wang J, Yang Y, Liu J, Qiu J, Zhang D, Ou M, Kang Y, Zhu T, Zhou C (2023) Loss of sodium leak channel (NALCN) in the ventral dentate gyrus impairs neuronal activity of the glutamatergic neurons for inflammation-induced depression in male mice. *Brain Behav Immun* 110:13–29. <https://doi.org/10.1016/j.bbi.2023.02.013>
 1209. Patoine D, Bouchard K, Lemay AM, Bissonnette EY, Lauzon-Joset JF (2022) Specificity of CD200/CD200R pathway in LPS-induced lung inflammation. *Front Immunol* 13:1092126. <https://doi.org/10.3389/fimmu.2022.1092126>
 1210. Dolga AM, Letsche T, Gold M, Doti N, Bacher M, Chiamvimonvat N, Dodel R, Culmsee C (2012) Activation of KCNN3/SK3/K(Ca)2.3 channels attenuates enhanced calcium influx and inflammatory cytokine production in activated microglia. *Glia* 60(12):2050–2064. <https://doi.org/10.1002/glia.22419>
 1211. Canham L, Sendac S, Diagbouga MR, Wolodimeroff E, Pirri D, Tardajos Ayllon B, Feng S, Souilhol C, Chico TJA, Evans PC et al (2023) EVA1A (Eva-1 Homolog A) promotes endothelial apoptosis and inflammatory activation under disturbed flow via regulation of autophagy. *Arterioscler Thromb Vasc Biol* 43(4):547–561. <https://doi.org/10.1161/ATVBAHA.122.318110>
 1212. Gerecke C, Scholtka B, Löwenstein Y, Fait I, Gottschalk U, Rogoll D, Melcher R, Kleuser B (2015) Hypermethylation of ITGA4, TFPI2 and VIMENTIN promoters is increased in inflamed colon tissue: putative risk markers for colitis-associated cancer. *J Cancer Res Clin Oncol* 141(12):2097–2107. <https://doi.org/10.1007/s00432-015-1972-8>
 1213. Tang J, Song X, Ji G, Wu H, Sun S, Lu S, Li Y, Zhang C, Zhang H (2018) A novel mutation in the DYSF gene in a patient with a presumed inflammatory myopathy. *Neuropathology*. <https://doi.org/10.1111/neup.12474>
 1214. Xie X, Yuan Y, Huang Y, Hong X, Hong S, Chen G, Chen Y, Lin Y, Lu W, Fu W et al (2023) Effects of COL1A1 and SYTL2 on inflammatory cell infiltration and poor extracellular matrix remodeling of the vascular wall in thoracic aortic aneurysm. *Chin Med J (Engl)*. <https://doi.org/10.1097/CM9.0000000000002808>
 1215. Choteau L, Vancaeyneste H, Le Roy D, Dubuquoy L, Romani L, Jouault T, Poulain D, Sendid B, Calandra T, Roger T et al (2017) Role of TLR1, TLR2 and TLR6 in the modulation of intestinal inflammation and *Candida albicans* elimination. *Gut Pathog* 9:9. <https://doi.org/10.1186/s13099-017-0158-0>
 1216. Zhang X, Wang Y, Lv J (2024) STAT4 targets KISS1 to inhibit the oxidative damage, inflammation and neuronal apoptosis in experimental PD models by inactivating the MAPK pathway. *Neurochem Int* 175:105683. <https://doi.org/10.1016/j.neuint.2024.105683>
 1217. Vanhove B, Hofer-Warbinek R, Kapetanopoulos A, Hofer E, Bach FH, de Martin R (1997) Gem, a GTP-binding protein from mitogen-stimulated T cells, is induced in endothelial cells upon activation by inflammatory cytokines. *Endothelium* 5(1):51–61. <https://doi.org/10.3109/10623329709044158>
 1218. Lamichhane S, Mo JS, Sharma G, Choi TY, Chae SC (2021) MicroRNA 452 regulates IL20RA-mediated JAK1/STAT3 pathway in inflammatory colitis and colorectal cancer. *Inflamm Res* 70(8):903–914. <https://doi.org/10.1007/s00011-021-01486-7>
 1219. Lee SN, Yoon SA, Song JM, Kim HC, Cho HJ, Choi AMK, Yoon JH (2022) Cell-type-specific expression of hyaluronan synthases HAS2 and HAS3 promotes goblet cell hyperplasia in allergic airway inflammation. *Am J Respir Cell Mol Biol* 67(3):360–374. <https://doi.org/10.1165/rcmb.2021-0527OC>
 1220. Chan DD, Xiao WF, Li J, de la Motte CA, Sandy JD, Plaas A (2015) Deficiency of hyaluronan synthase 1 (Has1) results in chronic joint inflammation and widespread intra-articular fibrosis in a murine model of knee joint cartilage damage. *Osteoarthritis Cartilage* 23(11):1879–1889. <https://doi.org/10.1016/j.joca.2015.06.021>
 1221. Xiang D, Li H, Pan J, Chen Y (2024) SLC4A4 moulds the inflammatory tumor microenvironment and predicts therapeutic expectations in colorectal cancer. *Curr Med Chem*. <https://doi.org/10.2174/0109298673277357231218070812>
 1222. Geng R, Zhao Y, Xu W, Ma X, Jiang Y, Han X, Zhao L, Li Y (2024) SIRPB1 regulates inflammatory factor expression in the glioma microenvironment via SYK: functional and bioinformatics insights. *J Transl Med* 22(1):338. <https://doi.org/10.1186/s12967-024-05149-z>
 1223. Zou X, Liu C, Huang Z, Xiang S, Li K, Yuan Y, Hao Y, Zhou F (2023) Inhibition of STEAP1 ameliorates inflammation and ferroptosis of acute lung injury caused by sepsis in LPS-induced human pulmonary microvascular endothelial cells. *Mol Biol Rep* 50(7):5667–5674. <https://doi.org/10.1007/s11033-023-08403-7>
 1224. Murcia JDG, Weinert A, Freitas CMT, Arens DK, Ferrel MN, Grose JH, Ridge PG, Wilson E, Kauwe JSK, Weber KS (2020) Atypical chemokine receptor ACKR2-V41A has decreased CCL2 binding, scavenging, and activation, supporting sustained inflammation and increased Alzheimer's disease risk. *Sci Rep* 10(1):8019. <https://doi.org/10.1038/s41598-020-64755-1>
 1225. Yi X, Tran E, Odiba JO, Qin CX, Ritchie RH, Baell JB (2024) The formyl peptide receptors FPR1 and FPR2 as targets for inflammatory disorders: recent advances in the development of small-molecule agonists. *Eur J Med Chem* 265:115989. <https://doi.org/10.1016/j.ejmech.2023.115989>
 1226. Dull K, Fazekas F, Deák D, Kovács D, Pólska S, Szegedi A, Zouboulis CC, Törőcsik D (2021) miR-146a modulates TLR1/2 and 4 induced inflammation and links it with proliferation and lipid production via the indirect regulation of GNG7 in human SZ95 sebocytes. *Sci Rep* 11(1):21510. <https://doi.org/10.1038/s41598-021-00907-1>
 1227. Pan L, Cho KS, Wei X, Xu F, Lennikov A, Hu G, Tang J, Guo S, Chen J, Kriukov E et al (2023) IGFBP1 is a master driver of microglia homeostasis and resolution of neuroinflammation in glaucoma and brain tauopathy. *Cell Rep* 42(8):112889. <https://doi.org/10.1016/j.celrep.2023.112889>
 1228. Zhang A, Yan S, Cao M, Wu D, Zhou J, Yu Z, Wu M, Liu Y, Lu S, Hu G et al (2020) Abnormal methylation of PIK3AP1 was involved in regulating the immune inflammatory response of GES-1 cells induced by *Helicobacter pylori*. *Biochem Biophys Res Commun* 524(1):36–42. <https://doi.org/10.1016/j.bbrc.2020.01.007>
 1229. Song F, Zhang Y, Pan Z, Hu X, Zhang Q, Huang F, Ye X, Huang P (2021) The role of alcohol dehydrogenase 1C in regulating inflammatory responses in ulcerative colitis. *Biochem Pharmacol* 192:114691. <https://doi.org/10.1016/j.bcp.2021.114691>
 1230. Li Y, Huang B, Yang H, Kan S, Yao Y, Liu X, Pu S, He G, Khan TM, Qi G et al (2020) Latexin deficiency in mice up-regulates inflammation and aggravates colitis through HECTD1/Rps3/NF- κ B pathway. *Sci Rep* 10(1):9868. <https://doi.org/10.1038/s41598-020-66789-x>
 1231. Kamath J (2012) Cancer-related fatigue, inflammation and thyrotropin-releasing hormone. *Curr Aging Sci* 5(3):195–202. <https://doi.org/10.2174/1874609811205030005>
 1232. Kanmaz-Özer M, Vural P, Doğru-Abbasoğlu S, Gedikbaşı A, Çil E, Karadağ B, Uysal M (2012) Polymorphisms of vascular cell adhesion molecule1 (VCAM1) in polycystic ovary syndrome determined by quantitative real-time polymerase chain reaction and melting curve analysis. *Eur J*

- Obstet Gynecol Reprod Biol 160(2):174–178. <https://doi.org/10.1016/j.ejogrb.2011.11.013>
1233. Xiong Z, Li B, Wang L, Zeng X, Li B, Sha X, Liu H (2019) AQP8 and AQP9 expression in patients with polycystic ovary syndrome and its association with in vitro fertilization-embryo transfer outcomes. *Exp Ther Med* 18(1):755–760. <https://doi.org/10.3892/etm.2019.7592>
1234. Abali R, Temel Yuksel I, Yuksel MA, Bulut B, Imamoglu M, Emirir V, Unal F, Guzel S, Celik C (2016) Implications of circulating irisin and Fbpl4 levels in patients with polycystic ovary syndrome. *J Obstet Gynaecol* 36(7):897–901. <https://doi.org/10.3109/01443615.2016.1174200>
1235. Liu J, Li J, Wu X, Zhang M, Yan G, Sun H, Li D (2024) High levels of fatty acid-binding protein 5 excessively enhances fatty acid synthesis and proliferation of granulosa cells in polycystic ovary syndrome. *J Ovarian Res* 17(1):44. <https://doi.org/10.1186/s13048-024-01368-6>
1236. Khalaf M, Morera J, Bourret A, Reznik Y, Denoual C, Herlicoviez M, Mitre H, Benhaim A (2013) BMP system expression in GCs from polycystic ovary syndrome women and the in vitro effects of BMP4, BMP6, and BMP7 on GC steroidogenesis. *Eur J Endocrinol* 168(3):437–444. <https://doi.org/10.1530/EJE-12-0891>
1237. Zhao Y, Zhang C, Huang Y, Yu Y, Li R, Li M, Liu N, Liu P, Qiao J (2015) Up-regulated expression of WNT5a increases inflammation and oxidative stress via PI3K/AKT/NF- κ B signaling in the granulosa cells of PCOS patients. *J Clin Endocrinol Metab* 100(1):201–211. <https://doi.org/10.1210/jc.2014>
1238. GohariTaban S, Amiri I, Soleimani Asl S, Saidjiam M, Yavangi M, Khan-larzadeh E, Mohammadpour N, Shabab N, Artimani T (2019) Abnormal expressions of ADAMTS-1, ADAMTS-9 and progesterone receptors are associated with lower oocyte maturation in women with polycystic ovary syndrome. *Arch Gynecol Obstet* 299(1):277–286. <https://doi.org/10.1007/s00404-018-4967-2>
1239. Zhang Y, Chen D, Wang D, Wang L, Weng Y, Wang H, Wu X, Wang Y (2022) Moderate aerobic exercise regulates follicular dysfunction by initiating brain-derived neurotrophic factor (BDNF)-mediated anti-apoptotic signaling pathways in polycystic ovary syndrome. *J Clin Med* 11(19):5584. <https://doi.org/10.3390/jcm11195584>
1240. Makhdoomi MJ, Shah IA, Rashid R, Rashid A, Singh S, Shah ZA, Ganie MA (2023) Effect modification of LHCGR gene variant (rs2293275) on clinico-biochemical profile, and levels of luteinizing hormone in polycystic ovary syndrome patients. *Biochem Genet* 61(4):1418–1432. <https://doi.org/10.1007/s10528-022-10327-z>
1241. Yu L, Wang C, Zhang D, Liu M, Liu T, Pan B, Che Q, Liu S, Wang B, Dong X et al (2023) Exosomal circ_0008285 in follicle fluid regulates the lipid metabolism through the miR-4644/ LDLR axis in polycystic ovary syndrome. *J Ovarian Res* 16(1):113. <https://doi.org/10.1186/s13048-023-01199-x>
1242. Nasri F, Doroudchi M, Namavar Jahromi B, Ghahesi-Fard B (2018) T helper cells profile and CD4+CD25+Foxp3+regulatory T cells in polycystic ovary syndrome. *Iran J Immunol* 15(3):175–185. <https://doi.org/10.22034/IJI.2018.39387>
1243. Karakose M, Demircan K, Tural E, Demirci T, Arslan MS, Sahin M, Celik HT, Kazanci F, Karakaya J, Cakal E et al (2016) Clinical significance of ADAMTS1, ADAMTS5, ADAMTS9 aggrecanases and IL-17A, IL-23, IL-33 cytokines in polycystic ovary syndrome. *J Endocrinol Invest* 39(11):1269–1275. <https://doi.org/10.1007/s40618-016-0472-2>
1244. Tu P, Yan S, Zhang F (2023) Circ_0005925 promotes granulosa cell growth by targeting MiR-324-3p to upregulate MAP2K6 in Polycystic ovary syndrome. *Biochem Genet* 61(1):21–34. <https://doi.org/10.1007/s10528-022-10238-z>
1245. Yang G, Yao G, Xu Z, Fan H, Liu X, He J, Kong Y, Kong D, Bai Y, He Q et al (2021) Expression level of ADAMTS1 in granulosa cells of PCOS patients is related to granulosa cell function, oocyte quality, and embryo development. *Front Cell Dev Biol* 9:647522. <https://doi.org/10.3389/fcell.2021.647522>
1246. Jin LY, Yu JE, Xu HY, Chen B, Yang Q, Liu Y, Guo MX, Zhou CL, Cheng Y, Pang HY et al (2024) Overexpression of Pde4d in rat granulosa cells inhibits maturation and atresia of antral follicles to induce polycystic ovary. *Biochim Biophys Acta Mol Basis Dis* 1870(1):166869. <https://doi.org/10.1016/j.bbadis.2023.166869>
1247. Manni L, Holmång A, Cajander S, Lundeberg T, Aloe L, Stener-Victorin E (2006) Effect of anti-NGF on ovarian expression of alpha1- and beta2-adrenoceptors, TrkA, p75NTR, and tyrosine hydroxylase in rats with steroid-induced polycystic ovaries. *Am J Physiol Regul Integr Comp Physiol* 290(3):R826–R835. <https://doi.org/10.1152/ajpregu.00078.2005>
1248. Akcalı A, Bostanci N, Özçaka Ö, Öztürk-Ceyhan B, Gümüş P, Tervahartala T, Husu H, Buduneli N, Sorsa T, Bellibasakis GN (2015) Levated matrix metalloproteinase-8 in saliva and serum in polycystic ovary syndrome and association with gingival inflammation. *Innate Immun* 21(6):619–625. <https://doi.org/10.1177/1753425915572172>
1249. Biyik I, Erten O, Isiklar OO, Ince O, Soysal C, Berikten D, Mammadli S, Oztas E (2021) Comparison of serum human Klotho levels and thiol/disulfide homeostasis in women with polycystic ovary syndrome and in healthy women. *Taiwan J Obstet Gynecol* 60(3):487–491. <https://doi.org/10.1016/j.tjog.2021.03.017>
1250. Li S, Zhai J, Liu J, Di F, Sun Y, Li W, Chen ZJ, Du Y (2018) Erythropoietin-producing hepatocellular A7 triggering ovulation indicates a potential beneficial role for polycystic ovary syndrome. *EBioMedicine* 36:539–552. <https://doi.org/10.1016/j.ebiom.2018.09.046>
1251. Temur M, Yılmaz Ö, Aksun S, Calan M, Özin Özbay P, Kumbasar S, Sever E (2017) The relationship of urocortin-2 with insulin resistance patients having PCOS. *Gynecol Endocrinol* 33(2):124–127. <https://doi.org/10.1080/09513590.2016.1240772>
1252. Dallel M, Douma Z, Finan RR, Hachani F, Letaifa DB, Mahjoub T, Almawi WY (2021) Contrasting association of Leptin receptor polymorphisms and haplotypes with polycystic ovary syndrome in Bahraini and Tunisian women: a case-control study. *Biosci Rep* 41(1):BSR20202726. <https://doi.org/10.1042/BSR20202726>
1253. Liu Y, Li Z, Wang Y, Cai Q, Liu H, Xu C, Zhang F (2022) IL-15 participates in the pathogenesis of polycystic ovary syndrome by affecting the activity of granulosa cells. *Front Endocrinol (Lausanne)* 13:787876. <https://doi.org/10.3389/fendo.2022.787876>
1254. Chugh RM, Park HS, Esfandyari S, Elsharoud A, Ulin M, Al-Hendy A (2021) Mesenchymal stem cell-conditioned media regulate steroidogenesis and inhibit androgen secretion in a PCOS cell model via BMP-2. *Int J Mol Sci* 22(17):9184. <https://doi.org/10.3390/ijms22179184>
1255. Liu HW, Zhang F, Fan P, Bai H, Zhang JX, Wang Y (2013) Effects of apolipoprotein E genotypes on metabolic profile and oxidative stress in southwest Chinese women with polycystic ovary syndrome. *Eur J Obstet Gynecol Reprod Biol* 170(1):146–151. <https://doi.org/10.1016/j.ejogrb.2013.04.016>
1256. Cabus U, Kabukcu C, Fenkci S, Caner V, Oztekin O, Fenkci V, Enli Y (2020) Serum Caspase-1 levels in women with polycystic ovary syndrome. *Taiwan J Obstet Gynecol* 59(2):207–210. <https://doi.org/10.1016/j.tjog.2020.01.007>
1257. Li J, Lin Z, Wang S, Shi Q (2023) Angiotensin converting enzyme (ACE) gene I/D polymorphism is significantly associated with insulin resistance and polycystic ovary syndrome: a meta-analysis. *Gynecol Obstet Invest* 88(3):174–184. <https://doi.org/10.1159/000530089>
1258. Hu M, Li J, Zhang Y et al (2018) Endometrial progesterone receptor isoforms in women with polycystic ovary syndrome. *Am J Transl Res* 10(8):2696–2705
1259. Koroglu N, Aydogan Mathyk B, Tola EN, Aslan Cetin B, Temel Yuksel I, Dag I, Yetkin YG (2019) Gremlin-1 and gremlin-2 levels in polycystic ovary syndrome and their clinical correlations. *Gynecol Endocrinol* 35(7):604–607. <https://doi.org/10.1080/09513590.2019.1566452>
1260. Hrovat A, Kravos NA, Goričar K, Jensterle Sever M, Janež A, Dolžan V (2016) SORCS1 polymorphism and insulin secretion in obese women with polycystic ovary syndrome. *Gynecol Endocrinol* 32(5):395–398. <https://doi.org/10.3109/09513590.2015.1126818>
1261. Cong P, Shang B, Zhang L, Wu Z, Wang Y, Li J, Zhang L (2024) New insights into the treatment of polycystic ovary syndrome: HKDC1 promotes the growth of ovarian granulocyte cells by regulating mitochondrial function and glycolysis. *J Mol Histol* 55(2):187–199. <https://doi.org/10.1007/s10735-024-10183-8>
1262. Tian Y, Zhang W, Zhao S, Sun Y, Bian Y, Chen T, Du Y, Zhang J, Wang Z, Huang T et al (2016) FADS1-FADS2 gene cluster confers risk to polycystic ovary syndrome. *Sci Rep* 6:21195. <https://doi.org/10.1038/srep21195>
1263. Malpique R, Sánchez-Infantes D, García-Beltrán C, Taxerás SD, López-Bermejo A, de Zegher F, Ibáñez L (2019) Towards a circulating marker of hepato-visceral fat excess: S100A4 in adolescent girls with polycystic ovary syndrome - Evidence from randomized clinical trials. *Pediatr Obes* 14(5):e12500. <https://doi.org/10.1111/jipo.12500>

1264. Zangeneh FZ, Naghizadeh MM, Bagheri M, Jafarabadi M (2017) Are CRH & NGF as psychoneuroimmune regulators in women with polycystic ovary syndrome? *Gynecol Endocrinol* 33(3):227–233. <https://doi.org/10.1080/09513590.2016.1250152>
1265. Kabakchieva P, Georgiev T, Gateva A, Hristova J, Kamenov Z (2021) Polycystic ovary syndrome and (pre)osteoarthritis: assessing the link between hyperandrogenism in young women and cartilage oligomeric matrix protein as a marker of cartilage breakdown. *Clin Rheumatol* 40(10):4217–4223. <https://doi.org/10.1007/s10067-021-05753-0>
1266. Sang Q, Zhang S, Zou S, Wang H, Feng R, Li Q, Jin L, He L, Xing Q, Wang L (2013) Quantitative analysis of follistatin (FST) promoter methylation in peripheral blood of patients with polycystic ovary syndrome. *Reprod Biomed Online* 26(2):157–163. <https://doi.org/10.1016/j.rbmo.2012.10.011>
1267. Ho CK, Wood JR, Stewart DR, Ewens K, Ankener W, Wickenheisser J, Nelson-Degrave V, Zhang Z, Legro RS, Dunaif A et al (2005) Increased transcription and increased messenger ribonucleic acid (mRNA) stability contribute to increased GATA6 mRNA abundance in polycystic ovary syndrome theca cells. *J Clin Endocrinol Metab* 90(12):6596–6602. <https://doi.org/10.1210/endojc.2005-0890>
1268. Tola EN, Koroglu ND, Yalcin SE, Oral HB (2018) The role of serum ADAMTS-1 and aggrecan on polycystic ovary syndrome in adolescents and younger-aged females. *Arch Gynecol Obstet* 297(2):487–493. <https://doi.org/10.1007/s00404-017-4578-3>
1269. Paulukinas RD, Mesaros CA, Penning TM (2022) Conversion of classical and 11-oxygenated androgens by insulin-induced AKR1C3 in a model of human PCOS adipocytes. *Endocrinology* 163(7):bqac068. <https://doi.org/10.1210/endo/bqac068>
1270. Russo N, Russo M, Daino D, Buccì F, Pluchino N, Casarosa E, Artini PG, Cela V, Luisi M, Genazzani AR (2012) Polycystic ovary syndrome: brain-derived neurotrophic factor (BDNF) plasma and follicular fluid levels. *Gynecol Endocrinol* 28(4):241–244. <https://doi.org/10.3109/09513590.2011.613969>
1271. Jiang Q, Miao R, Wang Y, Wang W, Zhao D, Niu Y, Ding Q, Li Y, Leung PCK, Wei D et al (2023) ANGPTL4 inhibits granulosa cell proliferation in polycystic ovary syndrome by EGFR/JAK1/STAT3-mediated induction of p21. *FASEB J* 37(2):e22693. <https://doi.org/10.1096/fj.2022011246RR>
1272. Li Y, Xu J, Li L, Bai L, Wang Y, Zhang J, Wang H (2022) Inhibition of Nicotinamide adenine dinucleotide phosphate oxidase 4 attenuates cell apoptosis and oxidative stress in a rat model of polycystic ovary syndrome through the activation of Nrf-2/HO-1 signaling pathway. *Mol Cell Endocrinol* 550:111645. <https://doi.org/10.1016/j.mce.2022.111645>
1273. Albahlol IA, Neamatallah M, Serria MS, El-Gilany AH, Setate YA, Alkasaby NM, Mostafa SA, Abdelaziz M, Elazab H, Ammar OA (2023) Vitamin D receptor gene polymorphism and polycystic ovary syndrome susceptibility. *BMC Med Genom* 16(1):108. <https://doi.org/10.1186/s12920-023-01541-8>
1274. Chen Z, Zeng H, Huang Q, Lin C, Li X, Sun S, Liu JP (2024) Increased GPC4 and clusterin associated with insulin resistance in patients with PCOS. *Endocr Connect* 13(3):e230428. <https://doi.org/10.1530/EC-23-0428>
1275. Alshammari AF, Alshammari AM, Farzan R, Alsobaie SF, Alageel AA, Ali KI (2023) A study on the immunological vitality of an inflammatory biomarker explored with rs5743708 polymorphism in TLR2 gene among Saudi women confirmed with polycystic ovarian syndrome. *Saudi J Biol Sci* 30(7):103687. <https://doi.org/10.1016/j.sjbs.2023.103687>
1276. Borthakur A, Prabhu D (2020) Role of IL-6 signalling in polycystic ovarian syndrome associated inflammation. *J Reprod Immunol* 141:103155. <https://doi.org/10.1016/j.jri.2020.103155>
1277. Govahi Kakhki F, Sargazi S, Montazerifar F, Majidpour M, Karajibani A, Karajibani M, Ghasemi M et al (2024) IGF2BP2 and IGFBP3 genotypes, haplotypes, and genetic models studies in polycystic ovary syndrome. *J Clin Lab Anal* 38(5):e25021. <https://doi.org/10.1002/jcla.25021>
1278. Li D, Jiao J, Zhou YM, Wang XX (2015) Epigenetic regulation of traf2- and Nck-interacting kinase (TNIK) in polycystic ovary syndrome. *Am J Transl Res* 7(6):1152–1160
1279. Liu Q, Jiang J, Shi Y, Mo Z, Li M (2020) Apelin/apelin receptor: a new therapeutic target in polycystic ovary syndrome. *Life Sci* 260:118310. <https://doi.org/10.1016/j.lfs.2020.118310>
1280. Tal R, Seifer DB, Grazi RV, Malter HE (2014) Follicular fluid placental growth factor is increased in polycystic ovarian syndrome: correlation with ovarian stimulation. *Reprod Biol Endocrinol* 12:82. <https://doi.org/10.1186/1477-7827-12-82>
1281. Arpacı H (2019) Circulating neuregulin-1 levels in polycystic ovary syndrome. *J Obstet Gynaecol* 39(4):504–509. <https://doi.org/10.1080/01443615.2018.1519530>
1282. Javidan M, Changaei M, Ramezani Tehrani F, Mosaffa N, Noroozadeh M, Hosseinzadeh R, Rajaei S (2022) Altered expression of leukemia inhibitory factor (LIF), LIFR, gp130, and IL11 in the embryo implantation site of rat model with prenatal androgen-induced polycystic ovary syndrome. *Biochem Biophys Res Commun* 605:24–30. <https://doi.org/10.1016/j.bbrc.2022.03.053>
1283. Liu Z, Liu C, Hao C, Xue Q, Huang X, Zhang N, Bao H, Qu Q (2016) Aberrant expression of angiopoietin-like proteins 1 and 2 in cumulus cells is potentially associated with impaired oocyte developmental competence in polycystic ovary syndrome. *Gynecol Endocrinol* 32(7):557–561. <https://doi.org/10.3109/09513590.2016.1138463>
1284. McCarthy EA, Dischino D, Maguire C, Leon S, Talbi R, Cheung E, Schteingart CD, Rivière PJM, Reed SD, Steiner RA et al (2022) Inhibiting kiss1 neurons with kappa opioid receptor agonists to treat polycystic ovary syndrome and vasomotor symptoms. *J Clin Endocrinol Metab* 107(1):e328–e347. <https://doi.org/10.1210/clinem/dgab602>
1285. Park JM, Gu BH, Lee EJ, Kim JY, Choi SW, Baek KH (2008) A single nucleotide polymorphism in exon 7 of sorbin and SH3-domain-containing-1 (SORBS1) in Korean PCOS patients. *Mol Med Rep* 1(1):93–97
1286. Kaltsas T, Pontikides N, Krassas GE, Seferiadis K, Lolis D, Messinis IE (1999) Growth hormone response to thyrotrophin releasing hormone in women with polycystic ovarian syndrome. *Hum Reprod* 14(11):2704–2708. <https://doi.org/10.1093/humrep/14.11.2704>
1287. Yu GI, Jun SE, Shin DH (2017) Associations of VCAM-1 gene polymorphisms with obesity and inflammation markers. *Inflamm Res* 66(3):217–225. <https://doi.org/10.1007/s00011-016-1006-2>
1288. Boaghi A, Pop RM, Vasilache SL, Banescu C, Hutanu A, Marginean OC, Pascanu IM (2020) Plasma RBP4 level in association with body composition, metabolic profile, STRA6 and RBP4 gene polymorphisms in obese romanian children. *Diabetes Metab Syndr Obes* 13:4643–4650. <https://doi.org/10.2147/DMSO.S273146>
1289. Xiang M, Qian X, Han L, Wang H, Wang J, Liu W, Gu Y, Yao S, Yang J, Zhang Y et al (2023) Aquaporin-8 ameliorates hepatic steatosis through farnesoid X receptor in obese mice. *iScience* 26(4):106561. <https://doi.org/10.1016/j.isci.2023.106561>
1290. Osorio-Conles O, Ibarzabal A, Balibrea JM, Vidal J, Ortega E, de Hollanda A (2023) FABP4 expression in subcutaneous adipose tissue is independently associated with circulating triglycerides in obesity. *J Clin Med* 12(3):1013. <https://doi.org/10.3390/jcm12031013>
1291. Correa-Rodríguez M, Schmidt-RioValle J, Rueda-Medina B (2018) SOX6 rs7117858 polymorphism is associated with osteoporosis and obesity-related phenotypes. *Eur J Clin Invest* 48(10):e13011. <https://doi.org/10.1111/eci.13011>
1292. Liu T, Kamiyoshi A, Tanaka M, Iida S, Sakurai T, Ichikawa-Shindo Y, Kawate H, Hirabayashi K, Dai K, Cui N et al (2018) RAMP3 deficiency enhances postmenopausal obesity and metabolic disorders. *Peptides* 110:10–18. <https://doi.org/10.1016/j.peptides.2018.10.006>
1293. Wang CY, Zhang CP, Li BJ, Jiang SS, He WH, Long SY, Tian Y (2020) MMP-12 as a potential biomarker to forecast ischemic stroke in obese patients. *Med Hypotheses* 136:109524. <https://doi.org/10.1016/j.mehy.2019.109524>
1294. Conroy MJ, Galvin KC, Kavanagh ME, Mongan AM, Doyle SL, Gilmarin N, O'Farrelly C, Reynolds JV, Lysaght J (2016) CCR1 antagonism attenuates T cell trafficking to omentum and liver in obesity-associated cancer. *Immunol Cell Biol* 94(6):531–537. <https://doi.org/10.1038/icb.2016.26>
1295. Lopez-Yus M, Casamayor C, Soriano-Godes JJ, Borlan S, Gonzalez-Irazabal Y, Garcia-Sobreveia MP, Garcia-Rodríguez B, Del Moral-Bergos R, Calmarza P, Artigas JM et al (2023) Isthmin-1 (ISM1), a novel adipokine that reflects abdominal adipose tissue distribution in individuals with obesity. *Cardiovasc Diabetol* 22(1):335. <https://doi.org/10.1186/s12933-023-02075-0>
1296. Makey KL, Patterson SG, Robinson J, Loftin M, Waddell DE, Miele L, Chinchar E, Huang M, Smith AD, Weber M et al (2013) Increased plasma levels of soluble vascular endothelial growth factor receptor 1 (sFlt-1) in women by moderate exercise and increased plasma levels of vascular

- endothelial growth factor in overweight/obese women. *Eur J Cancer Prev* 22(1):83–89. <https://doi.org/10.1097/CEJ.0b013e328353ed81>
1297. Shibue K, Yamane S, Harada N, Hamasaki A, Suzuki K, Joo E, Iwasaki K, Nasteska D, Harada T, Hayashi Y et al (2015) atty acid-binding protein 5 regulates diet-induced obesity via GIP secretion from enteroendocrine K cells in response to fat ingestion. *Am J Physiol Endocrinol Metab* 308(7):E583–E591. <https://doi.org/10.1152/ajpendo.00543.2014>
 1298. Nizamuddin S, Govindaraj P, Saxena S, Kashyap M, Mishra A, Singh S, Rotti H, Raval R, Nayak J, Bhat BK et al (2016) Reply to "Lack of replication of association of THSD7A with obesity." *Int J Obes (Lond)* 40(4):727–728. <https://doi.org/10.1038/ijo.2016.16>
 1299. Sekar R, Chow BK (2014) Secretin receptor-knockout mice are resistant to high-fat diet-induced obesity and exhibit impaired intestinal lipid absorption. *FASEB J* 28(8):3494–3505. <https://doi.org/10.1096/fj.13-247536>
 1300. Zhu J, Qiu J, Magrane G, Abedalthagafi M, Zanko A, Golabi M, Chehab FF (2012) Duplication of C7orf58, WNT16 and FAM3C in an obese female with a t(7;22)(q32.1;q11.2) chromosomal translocation and clinical features resembling coffin-siris syndrome. *PLoS ONE* 7(12):e52353. <https://doi.org/10.1371/journal.pone.0052353>
 1301. Shu Y, Gumma N, Hassan F, Branch DA, Baer LA, Ostrowski MC, Stanford KI, Baskin KK, Mehta KD (2023) Hepatic protein kinase Cbeta deficiency mitigates late-onset obesity. *J Biol Chem* 299(8):104917. <https://doi.org/10.1016/j.jbc.2023.104917>
 1302. Mowers J, Uhm M, Reilly SM, Simon J, Leto D, Chiang SH, Chang L, Saltiel AR (2013) Inflammation produces catecholamine resistance in obesity via activation of PDE3B by the protein kinases IKKε and TBK1. *Elife* 2:e01119. <https://doi.org/10.7554/eLife.01119>
 1303. Liu J, Wang Q, Zhang L, Fu J, An Y, Meng H, Wang G (2021) Increased prolactin is an adaptive response to protect against metabolic disorders in obesity. *Endocr Pract* 27(7):728–735. <https://doi.org/10.1016/j.eprac.2021.01.002>
 1304. Koutaki D, Michos A, Bacopoulou F, Charmandari E (2021) The emerging role of Sfrp5 and Wnt5a in the pathogenesis of obesity: implications for a healthy diet and lifestyle. *Nutrients* 13(7):2459. <https://doi.org/10.3390/nu13072459>
 1305. Edwards TL, Velez Edwards DR, Villegas R, Cohen SS, Buchowski MS, Fowke JH, Schlundt D, Long J, Cai Q, Zheng W et al (2012) HTR1B, ADIPOR1, PPARGC1A, and CYP19A1 and obesity in a cohort of Caucasians and African Americans: an evaluation of gene-environment interactions and candidate genes. *Am J Epidemiol* 175(1):11–21. <https://doi.org/10.1093/aje/kwr272>
 1306. Monroy A, Kamath S, Chavez AO, Centonze VE, Veerasamy M, Barrentine A, Wewer JJ, Coletta DK, Jenkinson C, Jhingan RM et al (2009) Impaired regulation of the TNF-alpha converting enzyme/tissue inhibitor of metalloproteinase 3 proteolytic system in skeletal muscle of obese type 2 diabetic patients: a new mechanism of insulin resistance in humans. *Diabetologia* 52(10):2169–2181. <https://doi.org/10.1007/s00125-009-1451-3>
 1307. Song NJ, Lee A, Yasmeen R, Shen Q, Yang K, Kumar SB, Muhanna D, Arni-palli S, Noria SF, Needleman BJ et al (2022) Epiregulin as an alternative ligand for leptin receptor alleviates glucose intolerance without change in obesity. *Cells* 11(3):425. <https://doi.org/10.3390/cells11030425>
 1308. Sandrini L, Di Minno A, Amadio P, Ieraci A, Tremoli E, Barbieri SS (2018) Association between obesity and circulating brain-derived neurotrophic factor (BDNF) levels: systematic review of literature and meta-analysis. *Int J Mol Sci* 19(8):2281. <https://doi.org/10.3390/ijms19082281>
 1309. Ceddia RP, Zurawski Z, Thompson Gray A, Adegboye F, McDonald-Boyer A, Shi F, Liu D, Maldonado J, Feng J, Li Y et al (2023) Gβγ-SNAP25 exocytotic brake removal enhances insulin action, promotes adipocyte browning, and protects against diet-induced obesity. *J Clin Invest* 133(19):e160617. <https://doi.org/10.1172/JCI160617>
 1310. Shapira S, Kazanov D, Dankner R, Fishman S, Stern N, Arber N (2021) High expression level of PPARγ in CD24 knockout mice and gender-specific metabolic changes: a model of insulin-sensitive obesity. *J Pers Med* 11(1):50. <https://doi.org/10.3390/jpm11010050>
 1311. Onogi Y, Wada T, Okekawa A, Matsuzawa T, Watanabe E, Ikeda K, Nakano M, Kitada M, Koya D, Tsuneki H et al (2020) Pro-inflammatory macrophages coupled with glycolysis remodel adipose vasculature by producing platelet-derived growth factor-B in obesity. *Sci Rep* 10(1):670. <https://doi.org/10.1038/s41598-019-57368-w>
 1312. Du Y, Li S, Cui CJ, Zhang Y, Yang SH, Li JJ (2016) Leptin decreases the expression of low-density lipoprotein receptor via PCSK9 pathway: linking dyslipidemia with obesity. *J Transl Med* 14(1):276. <https://doi.org/10.1186/s12967-016-1032-4>
 1313. Shirakawa K, Sano M (2023) Drastic transformation of visceral adipose tissue and peripheral CD4 T cells in obesity. *Front Immunol* 13:1044737. <https://doi.org/10.3389/fimmu.2022.1044737>
 1314. Mahajan N, Khare P, Kondepudi KK, Bishnoi M (2021) TRPA1: Pharmacology, natural activators and role in obesity prevention. *Eur J Pharmacol* 912:174553. <https://doi.org/10.1016/j.ejphar.2021.174553>
 1315. Lee M, Lee Y, Kang I, Shin J, Sorn SR (2021) RMR-related MAP2K6 gene variation on the risk of overweight/obesity in children: a 3-year panel study. *J Pers Med* 11(2):91. <https://doi.org/10.3390/jpm11020091>
 1316. Clapcote SJ (2017) Phosphodiesterase-4B as a therapeutic target for cognitive impairment and obesity-related metabolic diseases. *Adv Neurobiol* 17:103–131. https://doi.org/10.1007/978-3-319-58811-7_5
 1317. Bittencourt A, Brum PO, Ribeiro CT, Gasparotto J, Bortolin RC, de Vargas AR, Heimfarth L, de Almeida RF, Moreira JCF, de Oliveira J et al (2022) High fat diet-induced obesity causes a reduction in brain tyrosine hydroxylase levels and non-motor features in rats through metabolic dysfunction, neuroinflammation and oxidative stress. *Nutr Neurosci* 25(5):1026–1040. <https://doi.org/10.1080/1028415X.2020.1831261>
 1318. Lauhio A, Färkkilä E, Pietiläinen KH, Åström P, Winkelmann A, Tervahartiala T, Pirilä E, Rissanen A, Kaprio J, Sorsa TA et al (2016) Association of MMP-8 with obesity, smoking and insulin resistance. *Eur J Clin Invest* 46(9):757–765. <https://doi.org/10.1111/eci.12649>
 1319. Mitra SR, Tan PY, Amini F (2019) Association of ADRB2 rs1042713 with obesity and obesity-related phenotypes and its interaction with dietary fat in modulating glycaemic indices in Malaysian adults. *J Nutr Metab* 2019:8718795. <https://doi.org/10.1155/2019/8718795>
 1320. Carreras-Badosa G, Puerto-Carranza E, Mas-Parés B, Gómez-Vilarrubla A, Gómez-Herrera B, Díaz-Roldán F, Riera-Pérez E, de Zegher F, Ibañez L, Bassols J et al (2023) Higher levels of serum α-Klotho are longitudinally associated with less central obesity in girls experiencing weight gain. *Front Endocrinol (Lausanne)* 14:1218949. <https://doi.org/10.3389/fendo.2023.1218949>
 1321. Kostopoulou E, Kalavrizioti D, Davoulou P, Sinopidis X, Papachristou E, Goumenos DS, Dimitriou G, Spiliotis BE, Papanotiriou M (2024) Soluble urokinase plasminogen activator receptor (suPAR) in children with obesity or type 1 diabetes as a marker of endothelial dysfunction: a cross-sectional study. *Eur J Pediatr*. <https://doi.org/10.1007/s00431-024-05496-5>
 1322. Carobbio S, Hagen RM, Lelliott CJ, Slawik M, Medina-Gomez G, Tan CY, Sicard A, Atherton HJ, Barbarroja N, Bjursell M et al (2013) Adaptive changes of the Insig1/SREBP1/SCD1 set point help adipose tissue to cope with increased storage demands of obesity. *Diabetes* 62(11):3697–3708. <https://doi.org/10.2337/db12-1748>
 1323. Zhong S, Chen L, Li X, Wang X, Ji G, Sun C, Liu Z (2023) Bmp8a deletion leads to obesity through regulation of lipid metabolism and adipocyte differentiation. *Commun Biol* 6(1):824. <https://doi.org/10.1038/s42003-023-05194-2>
 1324. Borg ML, Massart J, De Castro BT, Archilla-Ortega A, Smith JAB, Lanner JT, Alsina-Fernandez J, Yaden B, Culver AE, Karlsson HKR et al (2021) Modified UCN2 peptide treatment improves skeletal muscle mass and function in mouse models of obesity-induced insulin resistance. *J Cachexia Sarcopenia Muscle* 12(5):1232–1248. <https://doi.org/10.1002/jcsm.12746>
 1325. Zhang ZB, Ruan CC, Lin JR, Xu L, Chen XH, Du YN, Fu MX, Kong LR, Zhu DL, Gao PJ (2018) Perivascular adipose tissue-derived PDGF-D contributes to aortic aneurysm formation during obesity. *Diabetes* 67(8):1549–1560. <https://doi.org/10.2337/db18-0098>
 1326. Amor M, Itariu BK, Moreno-Viedma V, Keindl M, Jürets A, Prager G, Langer F, Grablowitz V, Zeyda M, Stulnig TM (2019) Serum myostatin is upregulated in obesity and correlates with insulin resistance in humans. *Exp Clin Endocrinol Diabetes* 127(8):550–556. <https://doi.org/10.1055/a-0641-5546>
 1327. Pérez-López A, Valadés D, Vázquez Martínez C, de Cos Blanco AI, Buján J, García-Honduvilla N (2018) Serum IL-15 and IL-15Ra levels are decreased in lean and obese physically active humans. *Scand J Med Sci Sports* 28(3):1113–1120. <https://doi.org/10.1111/sms.12983>

1328. Ribeiro SMTL, Lopes LR, Paula Costa G, Figueiredo VP, Shrestha D, Batista AP, Nicolato RLC, Oliveira FLP, Gomes JAS, Talvani A (2017) CXCL-16, IL-17, and bone morphogenetic protein 2 (BMP-2) are associated with overweight and obesity conditions in middle-aged and elderly women. *Immun Ageing* 14:6. <https://doi.org/10.1186/s12979-017-0089-0>
1329. Farup PG, Rootwelt H, Hestad K (2020) APOE - a genetic marker of comorbidity in subjects with morbid obesity. *BMC Med Genet* 21(1):146. <https://doi.org/10.1186/s12881-020-01082-2>
1330. Kimura H, Karasawa T, Usui F, Kawashima A, Endo Y, Kobayashi M, Sadatomo A, Nakamura J, Iwasaki Y, Yada T et al (2016) Caspase-1 deficiency promotes high-fat diet-induced adipose tissue inflammation and the development of obesity. *Am J Physiol Endocrinol Metab* 311(5):E881–E890. <https://doi.org/10.1152/ajpendo.00174.2016>
1331. Matheson J, Zhou XMM, Bourgault Z, Le Foll B (2021) Potential of fatty acid amide hydrolase (FAAH), monoacylglycerol lipase (MAGL), and diacylglycerol lipase (DAGL) enzymes as targets for obesity treatment: a narrative review. *Pharmaceuticals (Basel)* 14(12):1316. <https://doi.org/10.3390/ph14121316>
1332. Pérez-Díaz S, Koumaiaha Z, Borok MJ, Aurade F, Pini M, Periou B, Rouault C, Baba-Amer Y, Clément K, Derumeaux G et al (2022) Obesity impairs skeletal muscle repair through NID-1 mediated extracellular matrix remodeling by mesenchymal progenitors. *Matrix Biol* 112:90–115. <https://doi.org/10.1016/j.matbio.2022.08.006>
1333. Cheng ST, Wu S, Su CW, Teng MS, Hsu LA, Ko YL (2017) Association of ABCG2 rs2231142-A allele and serum uric acid levels in male and obese individuals in a Han Taiwanese population. *J Formos Med Assoc* 116(1):18–23. <https://doi.org/10.1016/j.jfma.2015.12.002>
1334. Khamlaoui W, Mehri S, Hammami S, Elosua R, Hammami M (2020) Association of angiotensin-converting enzyme insertion/deletion (ACE I/D) and angiotensinogen (AGT M235T) polymorphisms with the risk of obesity in a Tunisian population. *J Renin Angiotensin Aldosterone Syst* 21(2):1470320320907820. <https://doi.org/10.1177/1470320320907820>
1335. Maehle BO, Tretli S, Thorsen T (2004) The associations of obesity, lymph node status and prognosis in breast cancer patients: dependence on estrogen and progesterone receptor status. *APMIS* 112(6):349–357. <https://doi.org/10.1111/j.1600-0463.2004.apm1120605.x>
1336. Liu W, Li D, Yang M, Wang L, Xu Y, Chen N, Zhang Z, Shi J, Li W, Zhao S et al (2022) GREM2 is associated with human central obesity and inhibits visceral preadipocyte browning. *EBioMedicine* 78:103969. <https://doi.org/10.1016/j.ebiom.2022.103969>
1337. Lee S (2019) The genetic and epigenetic association of LDL Receptor Related Protein 1B (LRP1B) gene with childhood obesity. *Sci Rep* 9(1):1815. <https://doi.org/10.1038/s41598-019-38538-2>
1338. Aqeel Rashid F, Mahdi S, Abd-Alkader Mahdy S, Thamer SA (2021) Effect of obesity on plasma alkaline phosphatase activity in breast cancer. *Rep Biochem Mol Biol* 10(2):307–313. <https://doi.org/10.52547/rbmb.10.2.307>
1339. Wang Z, Fu Y (2022) Transient receptor potential cation channel 6 contributes to kidney injury induced by diabetes and hypertension. *Am J Physiol Renal Physiol* 322(1):F76–F88. <https://doi.org/10.1152/ajprenal.00296.2021>
1340. Landgraf K, Kühnapfel A, Schlanstein M, Biemann R, Isermann B, Kempf E, Kirsten H, Scholz M, Körner A (2022) Transcriptome analyses of adipose tissue samples identify EGFL6 as a candidate gene involved in obesity-related adipose tissue dysfunction in children. *Int J Mol Sci* 23(8):4349. <https://doi.org/10.3390/ijms23084349>
1341. Vassileva G, Hu W, Hoos L, Tetzloff G, Yang S, Liu L, Kang L, Davis HR, Hedrick JA, Lan H et al (2010) Gender-dependent effect of Gpbar1 genetic deletion on the metabolic profiles of diet-induced obese mice. *J Endocrinol* 205(3):225–232. <https://doi.org/10.1677/JOE-10-0009>
1342. Parikh D, Jayakumar S, Oliveira-Paula GH, Almonte V, Riascos-Bernal DF, Sibinga NES (2022) Allograft inflammatory factor-1-like is a situational regulator of leptin levels, hyperphagia, and obesity. *iScience* 25(10):105058. <https://doi.org/10.1016/j.isci.2022.105058>
1343. Cao J, Perez S, Goodwin B, Lin Q, Peng H, Qadri A, Zhou Y, Clark RW, Perreault M, Tobin JF et al (2014) Mice deleted for GPAT3 have reduced GPAT activity in white adipose tissue and altered energy and cholesterol homeostasis in diet-induced obesity. *Am J Physiol Endocrinol Metab* 306(10):E1176–E1187. <https://doi.org/10.1152/ajpendo.00666.2013>
1344. Kim JY, Tillison K, Zhou S, Wu Y, Smas CM (2007) The major facilitator superfamily member Slc37a2 is a novel macrophage-specific gene selectively expressed in obese white adipose tissue. *Am J Physiol Endocrinol Metab* 293(1):E110–E120. <https://doi.org/10.1152/ajpendo.00404.2006>
1345. Khamlaoui W, Mehri S, Hammami S, Hammouda S, Chraeif I, Elosua R, Hammami M (2020) Association between genetic variants in FADS1-FADS2 and ELOVL2 and obesity, lipid traits, and fatty acids in Tunisian population. *Clin Appl Thromb Hemost* 26:1076029620915286. <https://doi.org/10.1177/1076029620915286>
1346. Griffin JD, Zhu Y, Reeves A, Buhman KK, Greenberg AS (2024) Intestinal Acyl-CoA synthetase 5 (ACSL5) deficiency potentiates postprandial GLP-1 & PYY secretion, reduces food intake, and protects against diet-induced obesity. *Mol Metab*. <https://doi.org/10.1016/j.molmet.2024.101918>
1347. Lee S (2019) The association of genetically controlled CpG methylation (cg158269415) of protein tyrosine phosphatase, receptor type N2 (PTPRN2) with childhood obesity. *Sci Rep* 9(1):4855. <https://doi.org/10.1038/s41598-019-40486-w>
1348. Xi P, Zhu W, Zhang Y, Wang M, Liang H, Wang H, Tian D (2024) Upregulation of hypothalamic TRPV4 via S100a4/AMPA signaling pathway promotes the development of diet-induced obesity. *Biochim Biophys Acta Mol Basis Dis* 1870(1):166883. <https://doi.org/10.1016/j.bbadis.2023.166883>
1349. Bähr I, Jaeschke L, Nimptsch K, Janke J, Herrmann P, Kobelt D, Kielstein H, Pischon T, Stein U (2022) Obesity, colorectal cancer and MACC1 expression: a possible novel molecular association. *Int J Oncol* 60(2):17. <https://doi.org/10.3892/ijo.2022.5307>
1350. Villa-Osaba A, Gahete MD, Córdoba-Chacón J, de Lecea L, Pozo-Salas AI, Delgado-Lista FJ, Álvarez-Benito M, López-Miranda J, Luque RM, Castaño JP (2015) Obesity alters gene expression for GH/IGF-I axis in mouse mammary fat pads: differential role of cortistatin and somatostatin. *PLoS ONE* 10(3):e0120955. <https://doi.org/10.1371/journal.pone.0120955>
1351. Boumiza S, Bchir S, Ben Nasr H, Abbassi A, Jacob MP, Norel X, Tabka Z, Chahed K (2017) Role of MMP-1 (-519A/G, -1607 1G/2G), MMP-3 (Lys-45Glu), MMP-7 (-181A/G), and MMP-12 (-82A/G) variants and plasma MMP levels on obesity-related phenotypes and microvascular reactivity in a tunisian population. *Dis Markers* 2017:6198526. <https://doi.org/10.1155/2017/6198526>
1352. Mwangi SM, Nezami BG, Obukwelu B, Anitha M, Marri S, Fu P, Epperson MF, Le NA, Shanmugam M, Sitaraman SV et al (2014) Glial cell line-derived neurotrophic factor protects against high-fat diet-induced obesity. *Am J Physiol Gastrointest Liver Physiol* 306(6):G515–G525. <https://doi.org/10.1152/ajpgi.00364.2013>
1353. Wagner G, Fenzl A, Lindroos-Christensen J, Einwallner E, Husa J, Witzeneder N, Rauscher S, Gröger M, Derdak S, Mohr T et al (2021) LMO3 reprograms visceral adipocyte metabolism during obesity. *J Mol Med (Berl)* 99(8):1151–1171. <https://doi.org/10.1007/s00109-021-02089-9>
1354. Hetty S, Vranic M, Kamble PG, Lundqvist MH, Pereira MJ, Eriksson JW (2023) CABLES1 expression is reduced in human subcutaneous adipose tissue in obesity and type 2 diabetes but may not directly impact adipocyte glucose and lipid metabolism. *Adipocyte* 12(1):2242997. <https://doi.org/10.1080/21623945.2023.2242997>
1355. Dai JZ, Yang CC, Shueng PW, Wang YJ, Huang CS, Chao YC, Chen CH, Lin CW (2023) Obesity-mediated upregulation of the YAP/IL33 signaling axis promotes aggressiveness and induces an immunosuppressive tumor microenvironment in breast cancer. *J Cell Physiol* 238(5):992–1005. <https://doi.org/10.1002/jcp.30985>
1356. Bulló M, Peeraully MR, Trayhurn P, Folch J, Salas-Salvadó J (2007) Circulating nerve growth factor levels in relation to obesity and the metabolic syndrome in women. *Eur J Endocrinol* 157(3):303–310. <https://doi.org/10.1530/EJE-06-0716>
1357. Zhang J, Zhang Y, Sun T, Guo F, Huang S, Chandalia M, Abate N, Fan D, Xin HB, Chen YE et al (2013) Dietary obesity-induced Egr-1 in adipocytes facilitates energy storage via suppression of FOXO2. *Sci Rep* 3:1476. <https://doi.org/10.1038/srep01476>
1358. Gamu D, Juracic ES, Fajardo VA, Rietze BA, Tran K, Bombardier E, Tupling AR (2019) Phospholamban deficiency does not alter skeletal muscle SERCA pumping efficiency or predispose mice to diet-induced obesity. *Am J Physiol Endocrinol Metab* 316(3):E432–E442. <https://doi.org/10.1152/ajpendo.00288.2018>

1359. Cong L, Zhu Y, Pang H, Guanjun TU (2014) The interaction between aggrecan gene VNTR polymorphism and obesity in predicting incident symptomatic lumbar disc herniation. *Connect Tissue Res* 55(5–6):384–390. <https://doi.org/10.3109/03008207.2014.959117>
1360. Freitas-Alves DR, Vieira-Monteiro HA, Piranda DN, Sobral-Leite M, da Silva TSL, Bergmann A, Valença SS, Perini JA, Vianna-Jorge R (2018) PTGS2 polymorphism rs689466 favors breast cancer recurrence in obese patients. *Endocr Relat Cancer* 25(3):351–365. <https://doi.org/10.1530/ERC-17-0374>
1361. Yousof TR, Mejia-Benitez A, Morrison KM, Austin RC (2024) Reduced plasma GDF10 levels are positively associated with cholesterol impairment and childhood obesity. *Sci Rep* 14(1):1805. <https://doi.org/10.1038/s41598-024-51635-1>
1362. Wang KS, Zuo L, Pan Y, Xie C, Luo X (2015) Genetic variants in the CPNE5 gene are associated with alcohol dependence and obesity in Caucasian populations. *J Psychiatr Res* 71:1–7. <https://doi.org/10.1016/j.jpsychires.2015.09.008>
1363. Ning T, Zou Y, Yang M, Lu Q, Chen M, Liu W, Zhao S, Sun Y, Shi J, Ma Q et al (2017) Genetic interaction of DGAT2 and FAAH in the development of human obesity. *Endocrine* 56(2):366–378. <https://doi.org/10.1007/s12020-017-1261-1>
1364. Leung WK, Yau SY, Yang Y, Kwok AW, Wong EM, Cheung JK, Shum EW, Lam SC, Suen LK (2024) Effects of exercise interventions on brain-derived neurotrophic factor levels in overweight and obesity: a systematic review and meta-analysis. *J Exerc Sci Fit* 22(4):278–287. <https://doi.org/10.1016/j.jesf.2024.04.001>
1365. Michaelides M, Miller ML, Egervari G, Primeaux SD, Gomez JL, Ellis RJ, Landry JA, Szutorisz H, Hoffman AF, Lupica CR et al (2020) Striatal Rgs4 regulates feeding and susceptibility to diet-induced obesity. *Mol Psychiatry* 25(9):2058–2069. <https://doi.org/10.1038/s41380-018-0120-7>
1366. Zhang J, Chen Y, Yan L, Zhang X, Zheng X, Qi J, Yang F, Li J (2022) EphA3 deficiency in the hypothalamus promotes high-fat diet-induced obesity in mice. *J Biomed Res* 37(3):179–193. <https://doi.org/10.7555/JBR.36.20220168>
1367. Sato H, Taketomi Y, Ushida A, Isogai Y, Kojima T, Hirabayashi T, Miki Y, Yamamoto K, Nishito Y, Kobayashi T et al (2014) The adipocyte-inducible secreted phospholipases PLA2G5 and PLA2G2E play distinct roles in obesity. *Cell Metab* 20(1):119–132. <https://doi.org/10.1016/j.cmet.2014.05.0>
1368. Zhang G, Li R, Li W, Yang S, Sun Q, Yin H, Wang C, Hou B, Wang H, Yu L et al (2021) Toll-like receptor 3 ablation prevented high-fat diet-induced obesity and metabolic disorder. *J Nutr Biochem* 95:108761. <https://doi.org/10.1016/j.jnutbio.2021.108761>
1369. Schinzari F, Vizioli G, Campia U, Tesaro M, Cardillo C (2021) Variable changes of circulating ANGPTL3 and ANGPTL4 in different obese phenotypes: relationship with vasodilator dysfunction. *Biomedicines* 9(8):1037. <https://doi.org/10.3390/biomedicines9081037>
1370. Suzuki Y, Okabayashi K, Hasegawa H, Tsuruta M, Seishima R, Tokuda T, Kitagawa Y (2022) Role of EphB2/ephrin-B1 signalling in the development and progression of obesity-associated colorectal cancer. *Oncol Lett* 24(3):316. <https://doi.org/10.3892/ol.2022.13436>
1371. Li M, Liu L, Kang Y, Huang S, Xiao Y (2023) Circulating THBS1: a risk factor for nonalcoholic fatty liver disease in obese children. *Ann Nutr Metab* 79(1):16–28. <https://doi.org/10.1159/000527780>
1372. Doke M, Avecilla V, Felty Q (2018) Inhibitor of differentiation-3 and estrogenic endocrine disruptors: implications for susceptibility to obesity and metabolic disorders. *Biomed Res Int* 2018:6821601. <https://doi.org/10.1155/2018/6821601>
1373. Greatorex S, Kaur S, Xirouchaki CE, Goh PK, Wiede F, Genders AJ, Tran M, Jia Y, Raajendiran A, Brown WA et al (2023) Mitochondria- and NOX4-dependent antioxidant defense mitigates progression to nonalcoholic steatohepatitis in obesity. *J Clin Invest* 134(3):e162533. <https://doi.org/10.1172/JCI162533>
1374. Muhsin NIA, Bentley L, Bai Y, Goldsworthy M, Cox RD (2020) A novel mutation in the mouse Pcsk1 gene showing obesity and diabetes. *Mamm Genome* 31(1–2):17–29. <https://doi.org/10.1007/s00335-020-09826-4>
1375. Takahashi D, Mori T, Sohara E, Tanaka M, Chiga M, Inoue Y, Nomura N, Zeniya M, Ochi H, Takeda S et al (2017) WNK4 is an adipogenic factor and its deletion reduces diet-induced obesity in mice. *EBioMedicine* 18:118–127. <https://doi.org/10.1016/j.ebiom.2017.03.011>
1376. Kim OY, Lee SM, Chung JH, Do HJ, Moon J, Shin MJ (2012) Arginase I and the very low-density lipoprotein receptor are associated with phenotypic biomarkers for obesity. *Nutrition* 28(6):635–639. <https://doi.org/10.1016/j.nut.2011.09.012>
1377. Akter R, Afrose A, Sharmin S, Rezwan R, Rahman MR, Neelotpol S (2022) A comprehensive look into the association of vitamin D levels and vitamin D receptor gene polymorphism with obesity in children. *Biomed Pharmacother* 153:113285. <https://doi.org/10.1016/j.biopha.2022.113285>
1378. Zhang K, Zhu H, Wang L, Yang H, Pan H, Gong F (2021) Serum glypican4 and glycosylphosphatidylinositol-specific phospholipase D levels are associated with adipose tissue insulin resistance in obese subjects with different glucose metabolism status. *J Endocrinol Invest* 44(4):781–790. <https://doi.org/10.1007/s40618-020-01372-9>
1379. Pomar CA, Bonet ML, Ferre-Beltrán A, Fraile-Ribot PA, García-Gasalla M, Riera M, Picó C, Palou A (2022) Increased mRNA levels of ADAM17, IFITM3, and IFNE in peripheral blood cells are present in patients with obesity and may predict severe COVID-19 evolution. *Biomedicines* 10(8):2007. <https://doi.org/10.3390/biomedicines10082007>
1380. Klarin D, Emdin CA, Natarajan P, Conrad MF, Kathiresan S (2017) Genetic analysis of venous thromboembolism in UK biobank identifies the ZFPM2 locus and implicates obesity as a causal risk factor. *Circulat: Cardiovasc Gene* 10(2):e001643
1381. Cuesta N, Fernández-Veledo S, Punzón C, Moreno C, Barrocal B, Sreeramkumar V, Desco M, Fresno M (2022) Opposing actions of TLR2 and TLR4 in adipocyte differentiation and mature-onset obesity. *Int J Mol Sci* 23(24):15682. <https://doi.org/10.3390/ijms232415682>
1382. Gholami M, Sharifi F, Shahriari S, Khoshnevisan K, Larijani B, Amoli MM (2019) Association of interleukin-6 polymorphisms with obesity: a systematic review and meta-analysis. *Cytokine* 123:154769. <https://doi.org/10.1016/j.cyto.2019.154769>
1383. Abass MK, Al Shamsi A, Jan I, Masalawala MSY, Deeb A (2022) Combined SPINK1 mutations induce early-onset severe chronic pancreatitis in a child with severe obesity. *Endocrinol Diabetes Metab Case Rep*. <https://doi.org/10.1530/EDM-22-0273>
1384. Yao Z, Qi W, Zhang H, Zhang Z, Liu L, Shao Y, Zeng H, Yin J, Pan H, Guo X et al (2023) Down-regulated GAS6 impairs synovial macrophage efferocytosis and promotes obesity-associated osteoarthritis. *Elife* 12:e83069. <https://doi.org/10.7554/eLife.83069>
1385. Ong KL, Leung RY, Wong LY, Cherny SS, Sham PC, Lam TH, Lam KS, Cheung BM (2008) Association of F11 receptor gene polymorphisms with central obesity and blood pressure. *J Intern Med* 263(3):322–332. <https://doi.org/10.1111/j.1365-2796.2007.01886.x>
1386. Zhang C, Zhou L, Li S, Zhao J, Meng X, Ma L, Wang Y, Li C, Zheng L, Ming L (2022) Obesity accelerates immune evasion of non-small cell lung carcinoma via TFEB-dependent upregulation of Siglec-15 and glycolytic reprogramming. *Cancer Lett* 550:215918. <https://doi.org/10.1016/j.canlet.2022.215918>
1387. Masaki M, Kurisaki T, Shirakawa K, Sehara-Fujisawa A (2005) Role of meltrin alpha (ADAM12) in obesity induced by high-fat diet. *Endocrinology* 146(4):1752–1763. <https://doi.org/10.1210/en.2004-1082>
1388. Abdul Majeed S, Duzendorfer H, Weiner J, Heiker JT, Kiess W, Körner A, Landgraf K (2023) COBL, MKX and MYOC are potential regulators of brown adipose tissue development associated with obesity-related metabolic dysfunction in children. *Int J Mol Sci* 24(4):3085. <https://doi.org/10.3390/ijms24043085>
1389. Yang H, Seo SG, Shin SH, Min S, Kang MJ, Yoo R, Kwon JY, Yue S, Kim KH, Cheng JX et al (2017) 3'-Diindolylmethane suppresses high-fat diet-induced obesity through inhibiting adipogenesis of pre-adipocytes by targeting USP2 activity. *Mol Nutr Food Res*. <https://doi.org/10.1002/mnfr.201700119>
1390. Nixon M, Stewart-Fitzgibbon R, Fu J, Akhmedov D, Rajendran K, Mendoza-Rodriguez MG, Rivera-Molina YA, Gibson M, Berglund ED et al (2015) Skeletal muscle salt inducible kinase 1 promotes insulin resistance in obesity. *Mol Metab* 5(1):34–46. <https://doi.org/10.1016/j.molmet.2015.10.004>
1391. Tan JT, McLennan SV, Williams PF, Rezaeizadeh A, Lo LW, Bonner JG, Twigg SM (2013) Connective tissue growth factor/CCN-2 is upregulated in epididymal and subcutaneous fat depots in a dietary-induced obesity model. *Am J Physiol Endocrinol Metab* 304(12):E1291–E1302. <https://doi.org/10.1152/ajpendo.00654.2012>

1392. Al-Daghri NM, Manousopoulou A, Alokail MS, Yakout S, Alenad A, Garay-Baquero DJ, Fotopoulos M, Teng J, Al-Attas O, Al-Saleh Y et al (2018) Sex-specific correlation of IGFBP-2 and IGFBP-3 with vitamin D status in adults with obesity: a cross-sectional serum proteomics study. *Nutr Diabetes* 8(1):54. <https://doi.org/10.1038/s41387-018-0063-8>
1393. Michurina SV, Ishchenko IY, Arkhipov SA, Klimontov VV, Rachkovskaya LN, Kononov VI, Zavyalov EL (2016) Effects of melatonin, aluminum oxide, and polymethylsiloxane complex on the expression of LYVE-1 in the liver of mice with obesity and type 2 diabetes mellitus. *Bull Exp Biol Med* 162(2):269–272. <https://doi.org/10.1007/s10517-016-3592-y>
1394. Han CY, Kang I, Harten IA, Gebe JA, Chan CK, Omer M, Alonge KM, den Hartigh LJ, Gomes Kjerulf D, Goodspeed L et al (2020) Adipocyte-derived versican and macrophage-derived biglycan control adipose tissue inflammation in obesity. *Cell Rep* 31(13):107818. <https://doi.org/10.1016/j.celrep.2020.107818>
1395. Awazawa M, Gabel P, Tsaousidou E, Nolte H, Krüger M, Schmitz J, Ackermann PJ, Brandt C, Altmüller J, Motameny S et al (2017) A micro-RNA screen reveals that elevated hepatic ectodysplasin a expression contributes to obesity-induced insulin resistance in skeletal muscle. *Nat Med* 23(12):1466–1473. <https://doi.org/10.1038/nm.4420>
1396. Rask-Andersen M, Almén MS, Olausen HR, Olszewski PK, Eriksson J, Chavan RA, Levine AS, Fredriksson R, Schiöth HB (2011) Functional coupling analysis suggests link between the obesity gene FTO and the BDNF-NTRK2 signaling pathway. *BMC Neurosci* 12:117. <https://doi.org/10.1186/1471-2202-12-117>
1397. Hunyenyiwa T, Hendee K, Matus K, Kyi P, Mammoto T, Mammoto A (2021) Obesity inhibits angiogenesis through TWIST1-SLIT2 signaling. *Front Cell Dev Biol* 9:693410. <https://doi.org/10.3389/fcell.2021.693410>
1398. Liu JR, Deng ZH, Zhu XJ, Zeng YR, Guan XX, Li JH (2021) Roles of nicotinamide N-methyltransferase in obesity and type 2 diabetes. *Biomed Res Int* 2021:9924314. <https://doi.org/10.1155/2021/9924314>
1399. Teitsdottir UD, Arnardottir ES, Bjornsdottir E, Gislason T, Petersen PH (2018) Obesity modulates the association between sleep apnea treatment and CH13L1 levels but not CHIT1 activity in moderate to severe OSA: an observational study. *Sleep Breath* 22(4):1101–1109. <https://doi.org/10.1007/s11325-018-1731-6>
1400. Zhang X, Hou X, Xu C, Cheng S, Ni X, Shi Y, Yao Y, Chen L, Hu MG, Xia D (2023) Kaempferol regulates the thermogenic function of adipocytes in high-fat-diet-induced obesity via the CDK6/RUNX1/UCP1 signaling pathway. *Food Funct* 14(18):8201–8216. <https://doi.org/10.1039/d3fo00613a>
1401. De Los SS, Reyes-Castro LA, Coral-Vázquez RM, Mendez JP, Zambrano E, Canto P (2023) (-)-Epicatechin increases apelin/APLNR expression and modifies proteins involved in lipid metabolism of offspring descendants of maternal obesity. *J Nutr Biochem* 117:109350. <https://doi.org/10.1016/j.jnutbio.2023.109350>
1402. Pervanidou P, Chouliaras G, Akalestos A, Bastaki D, Apostolou F, Papassotiropoulos I, Chrousos GP (2014) Increased placental growth factor (PLGF) concentrations in children and adolescents with obesity and the metabolic syndrome. *Hormones (Athens)* 13(3):369–374. <https://doi.org/10.14310/horm.2002.1491>
1403. Abdelaziz HA, Abdelbaki TN, Dean YE, Assem S (2023) Is neuregulin-1 (NRG-1) a potential blood biomarker linking depression to obesity? A case-control study. *BMC Psychiatry* 23(1):670. <https://doi.org/10.1186/s12888-023-05160-6>
1404. Aerts E, Beckers S, Zegers D, Van Hoorenbeeck K, Massa G, Verrijken A, Verhulst SL, Van Gaal LF, Van Hul W (2016) CNV analysis and mutation screening indicate an important role for the NPY4R gene in human obesity. *Obesity (Silver Spring)* 24(4):970–976. <https://doi.org/10.1002/oby.21435>
1405. Osorio-Conles O, Guitart M, Moreno-Navarrete JM, Escoté X, Duran X, Fernandez-Real JM, Gomez-Foix AM, Fernández-Veledo S, Vendrell J (2017) Adipose tissue and serum CCDC80 in obesity and its association with related metabolic disease. *Mol Med* 23:225–234. <https://doi.org/10.2119/molmed.2017.00067>
1406. Abdel-Fadeil MR, Abd Allah ESH, Iraqi HM, Elgamal DA, Abdel-Ghani MA (2019) Experimental obesity and diabetes reduce male fertility: Potential involvement of hypothalamic Kiss-1, pituitary nitric oxide, serum vaspin and visfatin. *Pathophysiology* 26(3–4):181–189. <https://doi.org/10.1016/j.pathophys.2019.02.001>
1407. Drgonova J, Jacobsson JA, Han JC, Yanovski JA, Fredriksson R, Marcus C, Schiöth HB, Uhl GR (2013) Involvement of the neutral amino acid transporter SLC6A15 and leucine in obesity-related phenotypes. *PLoS ONE* 8(9):e68245. <https://doi.org/10.1371/journal.pone.0068245>
1408. Zhang Y, Guan Y, Pan S, Yan L, Wang P, Chen Z, Shen Q, Zhao F, Zhang X, Li J et al (2020) Hypothalamic extended synaptotagmin-3 contributes to the development of dietary obesity and related metabolic disorders. *Proc Natl Acad Sci USA* 117(33):20149–20158. <https://doi.org/10.1073/pnas.2004392117>
1409. Lin WH, Chiu KC, Chang HM, Lee KC, Tai TY, Chuang LM (2001) Molecular scanning of the human sorbin and SH3-domain-containing-1 (SORBS1) gene: positive association of the T228A polymorphism with obesity and type 2 diabetes. *Hum Mol Genet* 10(17):1753–1760. <https://doi.org/10.1093/hmg/10.17.1753>
1410. Bhutia YD, Mathew M, Sivaprakasam S, Ramachandran S, Ganapathy V (2022) Unconventional functions of amino acid transporters: role in macrophage-mediated insulin resistance and diet-induced obesity/metabolic syndrome (SLC38A5/SLC38A3) and diet-induced obesity/metabolic syndrome (SLC6A19/SLC6A14/SLC6A6). *Biomolecules* 12(2):235. <https://doi.org/10.3390/biom12020235>
1411. Kan S, Li R, Tan Y, Yang F, Xu S, Wang L, Zhang L, Sun X, Chen X, Yang Y et al (2022) Latexin deficiency attenuates adipocyte differentiation and protects mice against obesity and metabolic disorders induced by high-fat diet. *Cell Death Dis* 13(2):175. <https://doi.org/10.1038/s41419-022-04636>
1412. Rijkers J, Penders B, Dorenbos E, Straetemans S, Gerver WJ, Vreugdenhil A (2016) Pituitary response to thyrotropin releasing hormone in children with overweight and obesity. *Sci Rep* 6:31032. <https://doi.org/10.1038/srep31032>
1413. Siddiqui K, George TP, Nawaz SS, Joy SS (2019) VCAM-1, ICAM-1 and selectins in gestational diabetes mellitus and the risk for vascular disorders. *Future Cardiol* 15(5):339–346. <https://doi.org/10.2217/fca-2018-0042>
1414. Fruscalzo A, Viola L, Orsaria M, Marzinotto S, Bulfoni M, Driuli L, Londero AP, Mariuzzi L (2021) STRA6 and placental retinoid metabolism in gestational diabetes mellitus. *J Pers Med* 11(12):1301. <https://doi.org/10.3390/jpm11121301>
1415. Shan Y, Cui J, Kang X, Tang W, Lu Y, Gao Y, Chen L (2022) Aquaporin-8 overexpression is involved in vascular structure and function changes in placentas of gestational diabetes mellitus patients. *Open Life Sci* 17(1):1473–1486. <https://doi.org/10.1515/biol-2022-0522>
1416. Ron I, Mdah R, Zemet R, Ulman RY, Rathaus M, Brandt B, Mazaki-Tovi S, Hemi R, Barhod E, Tirosh A (2023) Adipose tissue-derived FABP4 mediates glucagon-stimulated hepatic glucose production in gestational diabetes. *Diabetes Obes Metab* 25(11):3192–3201. <https://doi.org/10.1111/dom.15214>
1417. Nuzzo AM, Giuffrida D, Moretti L, Re P, Grassi G, Menato G, Rolfo A (2021) Placental and maternal sFlt1/PlGF expression in gestational diabetes mellitus. *Sci Rep* 11(1):2312. <https://doi.org/10.1038/s41598-021-81785-5>
1418. Rassie K, Giri R, Joham AE, Mousa A, Teede H (2022) Prolactin in relation to gestational diabetes and metabolic risk in pregnancy and postpartum: a systematic review and meta-analysis. *Front Endocrinol (Lausanne)* 13:1069625. <https://doi.org/10.3389/fendo.2022.1069625>
1419. Artunc-Ulkumen B, Ulucay S, Pala HG, Cam S (2017) Maternal serum ADAMTS-9 levels in gestational diabetes: a pilot study. *J Matern Fetal Neonatal Med* 30(12):1442–1445. <https://doi.org/10.1080/14767058.2016.1219717>
1420. Bayman MG, Inal ZO, Hayiroglu F, Ozturk ENY, Gezginc K (2022) Foetal umbilical cord brain-derived neurotrophic factor (BDNF) levels in pregnancy with gestational diabetes mellitus. *J Obstet Gynaecol* 42(5):1097–1102. <https://doi.org/10.1080/01443615.2021.2006159>
1421. Cao X, Lu B, Gu Y, Li X, Guo D, Xia F (2022) miR-210-3p impairs pancreatic β-cell function by targeting Dtx1 in gestational diabetes mellitus. *J Environ Pathol Toxicol Oncol* 41(4):11–23. <https://doi.org/10.1615/JEnvironPatholToxicolOncol.2022041670>
1422. Sheu A, Chan Y, Ferguson A, Bakhtyari MB, Hawke W, White C, Chan YF, Bertolino PJ, Woon HG, Palendira U et al (2018) A proinflammatory CD4+ T cell phenotype in gestational diabetes mellitus. *Diabetologia* 61(7):1633–1643. <https://doi.org/10.1007/s00125-018-4615-1>
1423. Ozler S, Oztas E, Gumus Guler B, Erel O, Turhan Caglar A, Ergin M, Uygun D, Danisman N (2020) Are serum levels of ADAMTS5, TAS and TOS at

- 24–28 gestational weeks associated with adverse perinatal outcomes in gestational diabetic women? *J Obstet Gynaecol* 40(5):619–625. <https://doi.org/10.1080/01443615.2019.1634025>
1424. Chaparro A, Realini O, Hernández M, Albers D, Weber L, Ramírez V, Param F, Kusanovic JP, Sorsa T, Rice GE et al (2021) Early pregnancy levels of gingival crevicular fluid matrix metalloproteinases-8 and -9 are associated with the severity of periodontitis and the development of gestational diabetes mellitus. *J Periodontol* 92(2):205–215. <https://doi.org/10.1002/JPER.19-0743>
1425. Santos KD, Rosado EL, da Fonseca ACP, Belfort GP, da Silva LBG, Ribeiro-Alves M, Zembrzuski VM, Martínez JA, Saunders C (2022) FTO and ADRB2 genetic polymorphisms are risk factors for earlier excessive gestational weight gain in pregnant women with pregestational diabetes mellitus: results of a randomized nutrigenetic trial. *Nutrients* 14(5):1050. <https://doi.org/10.3390/nu14051050>
1426. Kang Y, Huang H, Li H, Sun W, Zhang C (2021) Functional genetic variants in the 3'UTR of PTPRD associated with the risk of gestational diabetes mellitus. *Exp Ther Med* 21(6):562. <https://doi.org/10.3892/etm.2021.9994>
1427. Li J, Qian G, Zhong X, Yu T (2018) Insulin treatment cannot promote lipogenesis in rat fetal lung in gestational diabetes mellitus because of failure to redress the imbalance among SREBP-1, SCAP, and INSIG-1. *DNA Cell Biol* 37(3):264–270. <https://doi.org/10.1089/dna.2017.3906>
1428. Mosavat M, Omar SZ, Tan PC, Razif MFM, Sthaneshwar P (2018) Leptin and soluble leptin receptor in association with gestational diabetes: a prospective case-control study. *Arch Gynecol Obstet* 297(3):797–803. <https://doi.org/10.1007/s00404-017-4617-0>
1429. Keckstein S, Pritz S, Amann N, Meister S, Beyer S, Jegen M, Kuhn C, Hutter S, Knabl J, Mahner S et al (2020) Sex specific expression of interleukin 7, 8 and 15 in placentas of women with gestational diabetes. *Int J Mol Sci* 21(21):8026. <https://doi.org/10.3390/ijms21218026>
1430. Ramanjaneya M, Butler AE, Bashir M, Bettahi I, Moin ASM, Ahmed L, Elrassy MA, Hunt SC, Atkin SL, Abou-Samra AB (2021) apoA2 correlates to gestational age with decreased apolipoproteins A2, C1, C3 and E in gestational diabetes. *BMJ Open Diabetes Res Care* 9(1):e001925. <https://doi.org/10.1136/bmjdr-2020-001925>
1431. Dmitrenko OP, Karpova NS, Nurbekov MK, Papyshva OV (2020) I/D polymorphism gene ACE and risk of preeclampsia in women with gestational diabetes mellitus. *Dis Markers* 2020:8875230. <https://doi.org/10.1155/2020/8875230>
1432. Houde AA, Ruchat SM, Allard C, Baillargeon JP, St-Pierre J, Perron P, Gaudet D, Brisson D, Hivert MF, Bouchard L (2015) LRP1B, BRD2 and CACNA1D: new candidate genes in fetal metabolic programming of newborns exposed to maternal hyperglycemia. *Epigenomics* 7(7):1111–1122. <https://doi.org/10.2217/epi.15.72>
1433. Xiong T, Zhong C, Sun G, Zhou X, Chen R, Li Q, Wu Y, Gao Q, Huang L, Hu X et al (2019) Early maternal circulating alkaline phosphatase with subsequent gestational diabetes mellitus and glucose regulation: a prospective cohort study in China. *Endocrine* 65(2):295–303. <https://doi.org/10.1007/s12020-019-01954-5>
1434. Tan YX, Hu SM, You YP, Yang GL, Wang W (2019) Replication of previous genome-wide association studies of HKDC1, BACE2, SLC16A11 and TMEM163 SNPs in a gestational diabetes mellitus case-control sample from Han Chinese population. *Diabetes Metab Syndr Obes* 12:983–989. <https://doi.org/10.2147/DMSO.S207019>
1435. Bo S, Gambino R, Menato G, Canil S, Ponzo V, Pinach S, Durazzo M, Ghigo E, Cassader M, Musso G (2015) Isoleucine-to-methionine substitution at residue 148 variant of PNPLA3 gene and metabolic outcomes in gestational diabetes. *Am J Clin Nutr* 101(2):310–318. <https://doi.org/10.3945/ajcn.114.095125>
1436. Tarnowski M, Malinowski D, Pawlak K, Dziedziczko V, Safranow K, Pawlik A (2017) GCK, GCKR, FADS1, DGKB/TMEM195 and CDKAL1 gene polymorphisms in women with gestational diabetes. *Can J Diabetes* 41(4):372–379. <https://doi.org/10.1016/j.cjcd.2016.11.009>
1437. Coban U, Celik ZB (2023) The promoter methylations of the autoimmune regulator (AIRE) gene and matrix metalloproteinase-3 (MMP-3) gene may have a role in gestational diabetes mellitus. *Eur Rev Med Pharmacol Sci* 27(3):1051–1057. https://doi.org/10.26355/eurrev_202302_31201
1438. Yilmaz F, Micili SC, Erbil G (2022) The role of FGF-4 and FGFR-2 on pre-implantation embryo development in experimental maternal diabetes. *Gynecol Endocrinol* 38(3):248–252. <https://doi.org/10.1080/09513590.2021.2005782>
1439. Fan W, Kang W, Li T, Luo D, Huang L, Yang Y, Sun Y (2021) Interleukin-33 and its receptor soluble suppression of tumorigenicity 2 in the diagnosis of gestational diabetes mellitus. *Int J Clin Pract* 75(12):e14944. <https://doi.org/10.1111/ijcp.14944>
1440. Yang J, Liu F, Li Y, Wu D, Zhang Z, Chen S, Deng M, Yang C, Yang J (2022) Forkhead box C2 is associated with insulin resistance in gestational diabetes mellitus. *Gynecol Endocrinol* 38(6):499–502. <https://doi.org/10.1080/09513590.2022.2072485>
1441. Näf S, Escote X, Ballesteros M, Yañez RE, Simón-Muela I, Gil P, Albaiges G, Vendrell J, Megia A (2014) Serum activin A and follistatin levels in gestational diabetes and the association of the Activin A-Follistatin system with anthropometric parameters in offspring. *PLoS ONE* 9(4):e92175. <https://doi.org/10.1371/journal.pone.0092175>
1442. Xu D, Gao C, Cao Y, Xiao B (2023) HOXC8 alleviates high glucose-triggered damage of trophoblast cells during gestational diabetes mellitus via activating TGFβ1-mediated Notch1 pathway. *Hum Cell* 36(1):195–208. <https://doi.org/10.1007/s13577-022-00816-z>
1443. Jaskolski MR, Diedrich AK, Odainic A, Schmidt SV, Schmitz MT, Strizek B, Gembruch U, Merz WM, Flöck A (2022) Brain-derived neurotrophic factor in gestational diabetes: analysis of maternal serum and cord blood pairs and comparison of dietary- and insulin-dependent GDM. *Metabolites* 12(6):482. <https://doi.org/10.3390/metabo12060482>
1444. Xu Y, Kang X, Liu H, Jiang H, Wang W (2023) LncRNA XIST promotes insulin resistance in gestational diabetes mellitus via the microRNA-181b-5p/NDRG2 axis. *Gen Physiol Biophys* 42(5):443–455. https://doi.org/10.4149/gpb_2023019
1445. Popova P, Vasilyeva L, Tkachuk A, Puzanov M, Golovkin A, Bolotko Y, Pustozero E, Vasilyeva E, Li O, Zazerskaya I et al (2018) A randomised, controlled study of different glycaemic targets during gestational diabetes treatment: effect on the level of adipokines in cord blood and ANGPTL4 expression in human umbilical vein endothelial cells. *Int J Endocrinol* 2018:6481658. <https://doi.org/10.1155/2018/6481658>
1446. Zhou H, Chen P, Dai F, Wang J (2022) Up-regulation of TGFBI and TGFBI2 in the plasma of gestational diabetes mellitus patients and its clinical significance. *Ir J Med Sci* 191(5):2029–2033. <https://doi.org/10.1007/s11845-021-02838-2>
1447. Liu J, Dai Q, Li W, Guo Y, Dai A, Wang Y, Deng M, Tang Z, She L, Chen X et al (2021) Association of vitamin D receptor gene polymorphisms with gestational diabetes mellitus—a case control study in Wuhan, China. *BMC Pregnancy Childbirth* 21(1):142. <https://doi.org/10.1186/s12884-021-03621-y>
1448. Deischinger C, Harreiter J, Leitner K, Wattar L, Baumgartner-Parzer S, Kautzky-Willer A (2021) Glypican-4 in pregnancy and its relation to glucose metabolism, insulin resistance and gestational diabetes mellitus status. *Sci Rep* 11(1):23898. <https://doi.org/10.1038/s41598-021-03454-x>
1449. Milan KL, Jayasuriya R, Harithpriya K, Anuradha M, Ramkumar KM (2024) MicroRNA-125b regulates vitamin D resistance by targeting CYP24A1 in the progression of gestational diabetes mellitus. *J Steroid Biochem Mol Biol* 239:106475. <https://doi.org/10.1016/j.jsbmb.2024.106475>
1450. Li Q, Pereira TJ, Moyce BL, Mahood TH, Doucette CA, Rempel J, Dolinsky VW (2016) In utero exposure to gestational diabetes mellitus conditions TLR4 and TLR2 activated IL-1beta responses in spleen cells from rat offspring. *Biochim Biophys Acta* 1862(11):2137–2146. <https://doi.org/10.1016/j.bbdis.2016.08.004>
1451. Zhang J, Chi H, Xiao H, Tian X, Wang Y, Yun X, Xu Y (2017) Interleukin 6 (IL-6) and tumor necrosis factor α (TNF-α) single nucleotide polymorphisms (SNPs), inflammation and metabolism in gestational diabetes mellitus in inner Mongolia. *Med Sci Monit* 23:4149–4157. <https://doi.org/10.12659/msm.903565>
1452. Zhang H, Chen Z, Wang X (2022) Differentiated serum levels of Krüppel-like factors 2 and 4, sP-selectin, and sE-selectin in patients with gestational diabetes mellitus. *Gynecol Endocrinol* 38(12):1121–1124. <https://doi.org/10.1080/09513590.2022.2164762>
1453. Gęca T, Kwaśniewska A (2020) The influence of gestational diabetes mellitus upon the selected parameters of the maternal and fetal system of insulin-like growth factors (IGF-1, IGF-2, IGFBP1-3)—a review and a clinical study. *J Clin Med* 9(10):3256. <https://doi.org/10.3390/jcm9103256>

1454. Wang Y, Zhao S, Peng W, Chen Y, Chi J, Che K, Wang Y (2022) The role of Slit-2 in gestational diabetes mellitus and its effect on pregnancy outcome. *Front Endocrinol (Lausanne)* 13:889505. <https://doi.org/10.3389/fendo.2022.889505>
1455. Guo YY, Li T, Liu H, Tang L, Li YC, Hu HT, Su YF, Lin Y, Wang YY, Li C et al (2020) Circulating levels of Elabela and Apelin in the second and third trimesters of pregnancies with gestational diabetes mellitus. *Gynecol Endocrinol* 36(10):890–894. <https://doi.org/10.1080/09513590.2020.1739264>
1456. Tarnowski M, Malinowski D, Safranow K, Dziedziczko V, Pawlik A (2018) HNF1B, TSPAN8 and NOTCH2 gene polymorphisms in women with gestational diabetes. *J Matern Fetal Neonatal Med* 31(7):837–842. <https://doi.org/10.1080/14767058.2017.1297793>
1457. Gorkem U, Togrul C, Arslan E (2020) Relationship between elevated serum level of placental growth factor and status of gestational diabetes mellitus. *J Matern Fetal Neonatal Med* 33(24):4159–4163. <https://doi.org/10.1080/14767058.2019.1598361>
1458. Zhang L, Lu B, Wang W, Miao S, Zhou S, Cheng X, Zhu J, Liu C (2021) Alteration of serum neuregulin 4 and neuregulin 1 in gestational diabetes mellitus. *Ther Adv Endocrinol Metab* 12:20420188211049616. <https://doi.org/10.1177/20420188211049616>
1459. Yanai S, Tokuhara D, Tachibana D, Saito M, Sakashita Y, Shintaku H, Koyama M (2016) Diabetic pregnancy activates the innate immune response through TLR5 or TLR1/2 on neonatal monocyte. *J Reprod Immunol* 117:17–23. <https://doi.org/10.1016/j.jri.2016.06.007>
1460. Liu L, Hu J, Wang N, Liu Y, Wei X, Gao M, Ma Y, Wen D (2020) A novel association of CCDC80 with gestational diabetes mellitus in pregnant women: a propensity score analysis from a case-control study. *BMC Pregnancy Childbirth* 20(1):53. <https://doi.org/10.1186/s12884-020-2743-3>
1461. Kapustin RV, Drobintseva AO, Alekseenkova EN, Onopriyuk AR, Arzhanova ON, Polyakova VO, Kvetnoy IM (2020) Placental protein expression of kisspeptin-1 (KISS1) and the kisspeptin-1 receptor (KISS1R) in pregnancy complicated by diabetes mellitus or preeclampsia. *Arch Gynecol Obstet* 301(2):437–445. <https://doi.org/10.1007/s00404-019-05408-1>
1462. Fadel MM, Abdel Ghaffar FR, Zwain SK, Ibrahim HM, Badr EA (2021) Serum netrin and VCAM-1 as biomarker for Egyptian patients with type II diabetes mellitus. *Biochem Biophys Rep* 27:101045. <https://doi.org/10.1016/j.bbrep.2021.101045>
1463. He QJ, Wang P, Liu QQ, Wu QG, Li YF, Wang J, Lee SC (2020) Secreted Wnt6 mediates diabetes-associated centrosome amplification via its receptor FZD4. *Am J Physiol Cell Physiol* 318(1):C48–C62. <https://doi.org/10.1152/ajpcell.00091.2019>
1464. Furuhashi M, Sakuma I, Morimoto T, Higashiura Y, Sakai A, Matsumoto M, Sakuma M, Shimabukuro M, Nomiya T, Arasaki O et al (2020) Independent and distinct associations of FABP4 and FABP5 With metabolic parameters in type 2 diabetes mellitus. *Front Endocrinol (Lausanne)* 11:575557. <https://doi.org/10.3389/fendo.2020.575557>
1465. Qi H, Yao L, Liu Q (2019) MicroRNA-96 regulates pancreatic β cell function under the pathological condition of diabetes mellitus through targeting Foxo1 and Sox6. *Biochem Biophys Res Commun* 519(2):294–301. <https://doi.org/10.1016/j.bbrc.2019.09.001>
1466. Wootton PT, Stephens JW, Hurel SJ, Durand H, Cooper J, Ninio E, Humphries SE, Talmud PJ (2006) Lp-PLA2 activity and PLA2G7 A379V genotype in patients with diabetes mellitus. *Atherosclerosis* 189(1):149–156. <https://doi.org/10.1016/j.atherosclerosis.2005.12.009>
1467. Goncalves I, Bengtsson E, Colhoun HM, Shore AC, Palombo C, Natali A, Edsfieldt A, Dunér P, Fredrikson GN, Björkbacka H et al (2015) Elevated plasma levels of MMP-12 are associated with atherosclerotic burden and symptomatic cardiovascular disease in subjects with type 2 diabetes. *Arterioscler Thromb Vasc Biol* 35(7):1723–1731. <https://doi.org/10.1161/ATVBAHA.115.305631>
1468. Melnik BC (2021) Synergistic effects of milk-derived exosomes and galactose on α -synuclein pathology in parkinson's disease and type 2 diabetes mellitus. *Int J Mol Sci* 22(3):1059. <https://doi.org/10.3390/ijms22031059>
1469. Overgaard M, Ravnsborg T, Lohse Z, Bytoft B, Clausen TD, Jensen RB, Damm P, Højlund K, Gravholt CH, Knorr S et al (2022) Apolipoprotein D and transthyretin are reduced in female adolescent offspring of women with type 1 diabetes: the EPICOM study. *Diabet Med* 39(7):e14776. <https://doi.org/10.1111/dme.14776>
1470. Mallardo D, Cortellini A, Capone M, Madonna G, Pinato DJ, Warren S, Simeone E, Ascierto PA (2022) Concomitant type 2 diabetes mellitus (T2DM) in metastatic melanoma patients could be related to lower level of LAG-3: a transcriptomic analysis of a retrospective cohort. *Ann Oncol* 33(4):445–447. <https://doi.org/10.1016/j.annonc.2022.01.007>
1471. Lewis JP, Palmer ND, Ellington JB, Divers J, Ng MC, Lu L, Langefeld CD, Freedman BI, Bowden DW (2010) Analysis of candidate genes on chromosome 20q12-13.1 reveals evidence for BMI mediated association of PREX1 with type 2 diabetes in European Americans. *Genomics* 96(4):211–219. <https://doi.org/10.1016/j.jygeno.2010.07.006>
1472. Bayat M, Chien S, Chehelcheraghi F (2021) Co-localization of Flt1 and tryptase of mast cells in skin wound of rats with type I diabetes: initial studies. *Acta Histochem* 123(2):151680. <https://doi.org/10.1016/j.acthis.2021.151680>
1473. Furuhashi M, Sakuma I, Morimoto T, Higashiura Y, Sakai A, Matsumoto M, Sakuma M, Shimabukuro M, Nomiya T, Arasaki O et al (2020) Independent and distinct associations of FABP4 and FABP5 with metabolic parameters in type 2 diabetes mellitus. *Front Endocrinol (Lausanne)* 11:575557. <https://doi.org/10.3389/fendo.2020.575557>
1474. Zbidi H, López JJ, Amor NB, Bartegi A, Salido GM, Rosado JA (2009) Enhanced expression of STIM1/Orai1 and TRPC3 in platelets from patients with type 2 diabetes mellitus. *Blood Cells Mol Dis* 43(2):211–213. <https://doi.org/10.1016/j.bcmd.2009.04.005>
1475. Reinhard L, Thomas C, Machalitz M, Lattwein E, Weiss LS, Vitu J, Wiech T, Stahl RAK, Hoxha E (2021) Characterization of THSD7A-antibodies not binding to glomerular THSD7A in a patient with diabetes mellitus but no membranous nephropathy. *Sci Rep* 11(1):16188. <https://doi.org/10.1038/s41598-021-94921-y>
1476. Freedman BI, Bowden DW, Murea M (2011) Protein kinase C- β gene variants and type 2 diabetes-associated kidney failure: what can we learn from gene association studies in diabetic nephropathy? *Am J Kidney Dis* 57(2):194–197. <https://doi.org/10.1053/j.ajkd.2010.10.042>
1477. Sano R, Miki T, Suzuki Y, Shimada F, Taira M, Kanatsuka A, Makino H, Hashimoto N, Saito Y (2001) Analysis of the insulin-sensitive phosphodiesterase 3B gene in type 2 diabetes. *Diabetes Res Clin Pract* 54(2):79–88. [https://doi.org/10.1016/s0168-8227\(01\)00287-x](https://doi.org/10.1016/s0168-8227(01)00287-x)
1478. Liang C, Sun R, Xu Y, Geng W, Li J (2020) Effect of the abnormal expression of BMP-4 in the blood of diabetic patients on the osteogenic differentiation potential of alveolar bmscs and the rescue effect of metformin: a bioinformatics-based study. *Biomed Res Int* 2020:7626215. <https://doi.org/10.1155/2020/7626215>
1479. Scairati R, Auriemma RS, Del Vecchio G, Di Meglio S, Pivonello R, Colao A (2024) Prolactin effects on the pathogenesis of diabetes mellitus. *Eur J Clin Invest*. <https://doi.org/10.1111/eci.14190>
1480. Korley FK, Goldstick J, Mastali M, Van Eyk JE, Barsan W, Meurer WJ, Sussman J, Falk H, Levine D (2019) Serum NfL (neurofilament light chain) levels and incident stroke in adults with diabetes mellitus. *Stroke* 50(7):1669–1675. <https://doi.org/10.1161/STROKEAHA.119.024941>
1481. Xu W, Sang YQ, Liu XK, Geng HF, Wang B, Shi L, Qiu QQ, Yu TP, Zhang Y, Zhang X et al (2023) Effect of glucagon-like peptide-1 receptor agonist on insulin secretion index and serum Wnt5a protein in patients with new-onset type 2 diabetes mellitus. *J Diabetes Metab Disord* 22(1):539–545. <https://doi.org/10.1007/s40200-022-01175-0>
1482. Alsters SI, Goldstone AP, Buxton JL, Zekavati A, Sosinsky A, Yiorkas AM, Holder S, Klaber RE, Bridges N, van Haelst MM et al (2015) Truncating homozygous mutation of carboxypeptidase E (CPE) in a morbidly obese female with type 2 diabetes mellitus, intellectual disability and hypogonadotropic hypogonadism. *PLoS ONE* 10(6):e0131417. <https://doi.org/10.1371/journal.pone.0131417>
1483. Boesgaard TW, Gjesing AP, Grarup N, Rutanen J, Jansson PA, Hribal ML, Sesti G, Fritsche A, Stefan N, Staiger H et al (2009) Variant near ADAMTS9 known to associate with type 2 diabetes is related to insulin resistance in offspring of type 2 diabetes patients—EUGENE2 study. *PLoS ONE* 4(9):e7236. <https://doi.org/10.1371/journal.pone.0007236>
1484. Moosaie F, Mohammadi S, Saghazadeh A, Dehghani Firouzabadi F, Rezaei N (2023) Brain-derived neurotrophic factor in diabetes mellitus: a systematic review and meta-analysis. *PLoS ONE* 18(2):e0268816. <https://doi.org/10.1371/journal.pone.0268816>

1485. Al-Daghri NM, Costa AS, Alokail MS, Zanzottera M, Alenad AM, Mohamed AK, Clerici M, Guerini FR (2016) Synaptosomal protein of 25 kDa (Snap25) polymorphisms associated with glycemic parameters in type 2 diabetes patients. *J Diabetes Res* 2016:8943092. <https://doi.org/10.1155/2016/8943092>
1486. Willecke F, Yuan C, Oka K, Chan L, Hu Y, Barnhart S, Bornfeldt KE, Goldberg IJ, Fisher EA (2015) Effects of high fat feeding and diabetes on regression of atherosclerosis induced by low-density lipoprotein receptor gene therapy in LDL receptor-deficient mice. *PLoS ONE* 10(6):e0128996. <https://doi.org/10.1371/journal.pone.0128996>
1487. Haskins K, Cooke A (2011) CD4 T cells and their antigens in the pathogenesis of autoimmune diabetes. *Curr Opin Immunol* 23(6):739–745. <https://doi.org/10.1016/j.coi.2011.08.004>
1488. Derbenev AV, Zsombok A (2016) Potential therapeutic value of TRPV1 and TRPA1 in diabetes mellitus and obesity. *Semin Immunopathol* 38(3):397–406. <https://doi.org/10.1007/s00281-015-0529-x>
1489. Shan TD, Yue H, Sun XG, Jiang YP, Chen L (2021) Rspo3 regulates the abnormal differentiation of small intestinal epithelial cells in diabetic state. *Stem Cell Res Ther* 12(1):330. <https://doi.org/10.1186/s13287-021-02385-8>
1490. Raza W, Guo J, Qadir MI, Bai B, Muhammad SA (2022) qPCR analysis reveals association of differential expression of SRR, NFKB1, and PDE4B genes with type 2 diabetes mellitus. *Front Endocrinol (Lausanne)* 12:774696. <https://doi.org/10.3389/fendo.2021.774696>
1491. Aliev G, Shahida K, Gan SH, Firoz C, Khan A, Abuzenadah AM, Kamal W, Kamal MA, Tan Y, Qu X et al (2014) Alzheimer disease and type 2 diabetes mellitus: the link to tyrosine hydroxylase and probable nutritional strategies. *CNS Neurol Disord Drug Targets* 13(3):467–477. <https://doi.org/10.2174/18715273113126660153>
1492. Reinbothe TM, Alkayali S, Ahlqvist E, Tuomi T, Isomaa B, Lyssenko V, Renström E (2013) The human L-type calcium channel Cav1.3 regulates insulin release and polymorphisms in CACNA1D associate with type 2 diabetes. *Diabetologia* 56(2):340–349. <https://doi.org/10.1007/s00125-012-2758-z>
1493. Merlo S, Starčević JN, Mankoč S, Šantl Letonja M, Cokan Vujkovic A, Zorc M, Petrovič D (2016) Vascular Endothelial growth factor polymorphism (rs2010963) and its receptor, kinase insert domain-containing receptor gene polymorphism (rs2071559), and markers of carotid atherosclerosis in patients with type 2 diabetes mellitus. *J Diabetes Res* 2016:1482194. <https://doi.org/10.1155/2016/1482194>
1494. Gu H, Jiang W, You N, Huang X, Li Y, Peng X, Dong R, Wang Z, Zhu Y, Wu K et al (2020) Soluble klotho improves hepatic glucose and lipid homeostasis in type 2 diabetes. *Mol Ther Methods Clin Dev* 18:811–823. <https://doi.org/10.1016/j.omtm.2020.08.002>
1495. Yaghoobkar H, Stancáková A, Freathy RM, Vangipurapu J, Weedon MN, Xie W, Wood AR, Ferrannini E, Mari A, Ring SM et al (2015) Association analysis of 29,956 individuals confirms that a low-frequency variant at CCND2 halves the risk of type 2 diabetes by enhancing insulin secretion. *Diabetes* 64(6):2279–2285. <https://doi.org/10.2337/db14-1456>
1496. Saif-Ali R, Al-Hamodi Z, Salem SD, Al-Habori M, Al-Dubai SA, Ismail IS (2024) Association of protein tyrosine phosphatase receptor type D and serine racemase genetic variants with type 2 diabetes in Malaysian Indians. *Indian J Endocrinol Metab* 28(1):55–59. https://doi.org/10.4103/ijem.ijem_209_23
1497. Guo M, Guo H, Zhu J, Wang F, Chen J, Wan C, Deng Y, Wang F, Xu L, Chen Y et al (2024) A novel subpopulation of monocytes with a strong interferon signature indicated by SIGLEC-1 is present in patients with recent-onset type 1 diabetes. *Diabetologia* 67(4):623–640. <https://doi.org/10.1007/s00125-024-06098-4>
1498. Habieb MS, Dawood AA, Emara MM, Elhelbawy MG, Elhelbawy NG (2020) The human genetic variants CYP2J2 rs2280275 and EPHX2 rs751141 and risk of diabetic nephropathy in Egyptian type 2 diabetic patients. *Appl Clin Genet* 13:165–178. <https://doi.org/10.2147/TACG.S281502>
1499. Wang XL, Greco M, Sim AS, Duarte N, Wang J, Wilcken DE (2002) Effect of CYP1A1 MspI polymorphism on cigarette smoking related coronary artery disease and diabetes. *Atherosclerosis* 162(2):391–397. [https://doi.org/10.1016/s0021-9150\(01\)00723-7](https://doi.org/10.1016/s0021-9150(01)00723-7)
1500. Adiga U, Banawali N, Mayur S, Bansal R, Ameera N, Rao S (2021) Association of insulin resistance and leptin receptor gene polymorphism in type 2 diabetes mellitus. *J Chin Med Assoc* 84(4):383–388. <https://doi.org/10.1097/JCMA.0000000000000507>
1501. Siewko K, Maciulewski R, Zielinska-Maciulewska A, Poplawska-Kita A, Szumowski P, Wawrusiewicz-Kurylonek N, Lipinska D, Milewski R, Gorska M, Kretowski A et al (2019) Interleukin-6 and interleukin-15 as possible biomarkers of the risk of autoimmune diabetes development. *Biomed Res Int* 2019:4734063. <https://doi.org/10.1155/2019/4734063>
1502. Zhang JM, Yu RQ, Wu FZ, Qiao L, Wu XR, Fu YJ, Liang YF, Pang Y, Xie CY (2021) BMP-2 alleviates heart failure with type 2 diabetes mellitus and doxorubicin-induced AC16 cell injury by inhibiting NLRP3 inflammasome-mediated pyroptosis. *Exp Ther Med* 22(2):897. <https://doi.org/10.3892/etm.2021.10329>
1503. El-Lebedy D, Raslan HM, Mohammed AM (2016) Apolipoprotein E gene polymorphism and risk of type 2 diabetes and cardiovascular disease. *Cardiovasc Diabetol* 15:12. <https://doi.org/10.1186/s12933-016-0329-1>
1504. Tsekmekidou X, Tsetsos F, Koufakis T, Karras SN, Georgitsi M, Papanas N, Papazoglou D, Roumeliotis A, Panagoutsos S, Thodis E et al (2020) Association between CUBN gene variants, type 2 diabetes and vitamin D concentrations in an elderly Greek population. *J Steroid Biochem Mol Biol* 198:105549. <https://doi.org/10.1016/j.jsbmb.2019.105549>
1505. Vincent JA, Mohr S (2007) Inhibition of caspase-1/interleukin-1beta signaling prevents degeneration of retinal capillaries in diabetes and galactosemia. *Diabetes* 56(1):224–230. <https://doi.org/10.2337/db06-0427>
1506. Taschler U, Radner FP, Heier C, Schreiber R, Schweiger M, Schoiswohl G, Preiss-Landl K, Jaeger D, Reiter B, Koefeler HC et al (2011) Monoglyceride lipase deficiency in mice impairs lipolysis and attenuates diet-induced insulin resistance. *J Biol Chem* 286(20):17467–17477. <https://doi.org/10.1074/jbc.M110.215434>
1507. Broquères-You D, Leré-Déan C, Merkulova-Rainon T, Mantsounga CS, Allanic D, Hainaud P, Contrères JO, Wang Y, Vilar J, Vially M et al (2012) Ephrin-B2-activated peripheral blood mononuclear cells from diabetic patients restore diabetes-induced impairment of postschismic neovascularization. *Diabetes* 61(10):2621–2632. <https://doi.org/10.2337/db11-1768>
1508. Khattab A, Torkamani A (2022) Nidogen-1 could play a role in diabetic kidney disease development in type 2 diabetes: a genome-wide association meta-analysis. *Hum Genom* 16(1):47. <https://doi.org/10.1186/s40246-022-00422-y>
1509. Pedersen-Bjergaard U, Agerholm-Larsen B, Pramming S, Hougaard P, Thorsteinsson B (2001) Activity of angiotensin-converting enzyme and risk of severe hypoglycaemia in type 1 diabetes mellitus. *Lancet* 357(9264):1248–1253. [https://doi.org/10.1016/S0140-6736\(00\)04405-6](https://doi.org/10.1016/S0140-6736(00)04405-6)
1510. Demir I, Yilmaz I, Horoz E, Bozkaya G, Bilgir O (2024) The relationship between stathmin-2 level and metabolic parameters in newly diagnosed type 2 diabetes mellitus patients. *Am J Med Sci*. <https://doi.org/10.1016/j.amjms.2024.03.023>
1511. Bonner SM, Pietropaolo SL, Fan Y, Chang Y, Sethupathy P, Morran MP, Beems M, Giannoukakis N, Trucco G, Palumbo MO et al (2012) Sequence variation in promoter of Ica1 gene, which encodes protein implicated in type 1 diabetes, causes transcription factor autoimmune regulator (AIRE) to increase its binding and down-regulate expression. *J Biol Chem* 287(21):17882–17893. <https://doi.org/10.1074/jbc.M111.319020>
1512. Graham S, Gorin Y, Abboud HE, Ding M, Lee DY, Shi H, Ding Y, Ma R (2011) Abundance of TRPC6 protein in glomerular mesangial cells is decreased by ROS and PKC in diabetes. *Am J Physiol Cell Physiol* 301(2):C304–C315. <https://doi.org/10.1152/ajpcell.00014.2011>
1513. Müssig K, Staiger H, Machicao F, Machann J, Schick F, Schäfer SA, Clausen CD, Holst JJ, Gallwitz B, Stefan N et al (2009) Preliminary report: genetic variation within the GPBAR1 gene is not associated with metabolic traits in white subjects at an increased risk for type 2 diabetes mellitus. *Metabolism* 58(12):1809–1811. <https://doi.org/10.1016/j.metabol.2009.06.012>
1514. Bayoumy NMK, El-Shabrawi MM, Leheta OF, Abo El-Ela AEM, Omar HH (2020) Association of ELMO1 gene polymorphism and diabetic nephropathy among Egyptian patients with type 2 diabetes mellitus. *Diabetes Metab Res Rev* 36(5):e3299. <https://doi.org/10.1002/dmrr.3299>
1515. Aslamy A, Oh E, Ahn M, Moin ASM, Chang M, Duncan M, Hacker-Stratton J, El-Shahawy M, Kandeel F, DiMeglio LA et al (2018) Exocytosis

- protein DOC2B as a biomarker of type 1 diabetes. *J Clin Endocrinol Metab* 103(5):1966–1976. <https://doi.org/10.1210/jc.2017-02492>
1516. Sun L, Zhang X, Wang T, Chen M, Qiao H (2017) Association of ANK1 variants with new-onset type 2 diabetes in a Han Chinese population from northeast China. *Exp Ther Med* 14(4):3184–3190. <https://doi.org/10.3892/etm.2017.4866>
 1517. Goodarzi MO, Lehman DM, Taylor KD, Guo X, Cui J, Quiñones MJ, Clee SM, Yandell BS, Blangero J, Hsueh WA et al (2007) SORCS1: a novel human type 2 diabetes susceptibility gene suggested by the mouse. *Diabetes* 56(7):1922–1929. <https://doi.org/10.2337/db06-1677>
 1518. Jiang H, Yao Q, An Y, Fan L, Wang J, Li H (2022) Baicalin suppresses the progression of Type 2 diabetes-induced liver tumor through regulating METTL3/m⁶A/HKDC1 axis and downstream p-JAK2/STAT1/cleaved Caspase3 pathway. *Phytomedicine* 94:153823. <https://doi.org/10.1016/j.phymed.2021.153823>
 1519. Moon S, Chung GE, Joo SK, Park JH, Chang MS, Yoon JW, Koo BK, Kim W (2022) A PNPLA3 polymorphism confers lower susceptibility to incident diabetes mellitus in subjects with nonalcoholic fatty liver disease. *Clin Gastroenterol Hepatol* 20(3):682–691.e8. <https://doi.org/10.1016/j.cgh.2021.04.038>
 1520. Li SW, Wang J, Yang Y, Liu ZJ, Cheng L, Liu HY, Ma P, Luo W, Liu SM (2016) Polymorphisms in FADS1 and FADS2 alter plasma fatty acids and desaturase levels in type 2 diabetic patients with coronary artery disease. *J Transl Med* 14:79. <https://doi.org/10.1186/s12967-016-0834-8>
 1521. Xia Q, Chesi A, Manduchi E, Johnston BT, Lu S, Leonard ME, Parlin UW, Rappaport EF, Huang P, Wells AD et al (2016) The type 2 diabetes presumed causal variant within TCF7L2 resides in an element that controls the expression of ACSL5. *Diabetologia* 59(11):2360–2368. <https://doi.org/10.1007/s00125-016-4077-2>
 1522. Smigoc Schweiger D, Mendez A, Kunilo Jamnik S, Bratanic N, Bratina N, Battelino T, Breclj J, Vidan-Jeras B (2014) Genetic risk for co-occurrence of type 1 diabetes and celiac disease is modified by HLA-C and killer immunoglobulin-like receptors. *Tissue Antigens* 84(5):471–478. <https://doi.org/10.1111/tan.12450>
 1523. Arner P, Petrus P, Esteve D, Boulomié A, Näslund E, Thorell A, Gao H, Dahlman I, Rydén M (2018) Screening of potential adipokines identifies S100A4 as a marker of pernicious adipose tissue and insulin resistance. *Int J Obes (Lond)* 42(12):2047–2056. <https://doi.org/10.1038/s41366-018-0018-0>
 1524. Sun T, Wang C, Huo L, Wang Y, Liu K, Wei C, Zhao H, Chen S, Ren L (2023) Serum cortistatin level in type 2 diabetes mellitus and its relationship with nonalcoholic fatty liver disease. *Int J Gen Med* 16:631–639. <https://doi.org/10.2147/IJGM.S396315>
 1525. Plesković A, Letonja MŠ, Vujković AC, Starčević JN, Caprnda M, Curilla E, Mozos I, Kruzliak P, Prosecky R, Petrović D (2017) Matrix metalloproteinase-3 gene polymorphism (rs3025058) affects markers atherosclerosis in type 2 diabetes mellitus. *Vasa* 46(5):363–369. <https://doi.org/10.1024/0301-1526/a000637>
 1526. Yang Y, Xie B, Ju C, Jin H, Ye X, Yao L, Jia M, Sun Z, Yuan Y (2019) The association of decreased serum gdnf level with hyperglycemia and depression in type 2 diabetes mellitus. *Endocr Pract* 25(9):951–965. <https://doi.org/10.4158/EP-2018-0492>
 1527. Shruthi S, Mohan V, Amutha A, Aravindhan V (2016) Increased serum levels of novel T cell cytokines IL-33, IL-9 and IL-17 in subjects with type-1 diabetes. *Cytokine* 86:6–9. <https://doi.org/10.1016/j.cyto.2016.07.007>
 1528. Hellweg R, Wöhrle M, Hartung HD, Stracke H, Hock C, Federlin K (1991) Diabetes mellitus-associated decrease in nerve growth factor levels is reversed by allogeneic pancreatic islet transplantation. *Neurosci Lett* 125(1):1–4. [https://doi.org/10.1016/0304-3940\(91\)90114-9](https://doi.org/10.1016/0304-3940(91)90114-9)
 1529. Häkansson J, Eliasson B, Smith U, Enerbäck S (2011) Adipocyte mitochondrial genes and the forkhead factor FOXO2 are decreased in type 2 diabetes patients and normalized in response to rosiglitazone. *Diabetol Metab Syndr* 3:32. <https://doi.org/10.1186/1758-5996-3-32>
 1530. Arellano Perez Vertti RD, Aguilar Muñoz LS, Morán Martínez J, González Galarza FF, Arguello AR (2019) Cartilage oligomeric matrix protein levels in type 2 diabetes associated with primary knee osteoarthritis patients. *Genet Test Mol Biomarkers* 23(1):16–22. <https://doi.org/10.1089/gtmb.2018.0184>
 1531. Motahari Rad M, Bijeh N, Attarzadeh Hosseini SR, Raouf SA (2023) The effect of two concurrent exercise modalities on serum concentrations of FGF21, irisin, follistatin, and myostatin in men with type 2 diabetes mellitus. *Arch Physiol Biochem* 129(2):424–433. <https://doi.org/10.1080/13813455.2020.1829649>
 1532. Sun X, Lee HC, Lu T (2024) Sorbs2 deficiency and vascular BK channelopathy in diabetes. *Circ Res* 134(7):858–871. <https://doi.org/10.1161/CIRCRESAHA.123.323538>
 1533. Yasuhara J, Manivannan SN, Majumdar U, Gordon DM, Lawrence PJ, Aljuhani M, Myers K, Stiver C, Bigelow AM, Galantowicz M et al (2024) Novel pathogenic GATA6 variant associated with congenital heart disease, diabetes mellitus and necrotizing enterocolitis. *Pediatr Res* 95(1):146–155. <https://doi.org/10.1038/s41390-023-02811-y>
 1534. Yu CY, Yang CY, Rui ZL (2019) MicroRNA-125b-5p improves pancreatic β -cell function through inhibiting JNK signaling pathway by targeting DACT1 in mice with type 2 diabetes mellitus. *Life Sci* 224:67–75. <https://doi.org/10.1016/j.lfs.2019.01.031>
 1535. Zhang Q, Huang Y, Li X, Liu H, He B, Wang B, Ma Y, Zhou X, Liu Y, Wu S (2019) Tangduqing granules attenuate insulin resistance and abnormal lipid metabolism through the coordinated regulation of PPAR γ and DGAT2 in type 2 diabetic rats. *J Diabetes Res* 2019:7403978. <https://doi.org/10.1155/2019/7403978>
 1536. Jiang T, Li Y, He S, Huang N, Du M, Zhai Q, Pu K, Wu M, Yan C, Ma Z et al (2023) Reprogramming astrocytic NDRG2/NF- κ B/C3 signaling restores the diabetes-associated cognitive dysfunction. *EbioMedicine* 95:104653. <https://doi.org/10.1016/j.ebiom.2023.104653>
 1537. Poetsch F, Henze LA, Estepa M, Moser B, Pieske B, Lang F, Eckardt KU, Alesutan I, Voelkl J (2020) Role of SGK1 in the osteogenic transdifferentiation and calcification of vascular smooth muscle cells promoted by hyperglycemic conditions. *Int J Mol Sci* 21(19):7207. <https://doi.org/10.3390/ijms21197207>
 1538. Gusarova V, O'Dushlaine C, Teslovich TM, Benotti PN, Mirshahi T, Gottesman O, Van Hout CV, Murray MF, Mahajan A, Nielsen JB et al (2018) Genetic inactivation of ANGPTL4 improves glucose homeostasis and is associated with reduced risk of diabetes. *Nat Commun* 9(1):2252. <https://doi.org/10.1038/s41467-018-04611-z>
 1539. Broquères-You D, Leré-Déan C, Merkulova-Rainon T, Mantsounga CS, Allanec D, Hainaud P, Contrères JO, Wang Y, Vilar J, Virally M et al (2012) Ephrin-B2-activated peripheral blood mononuclear cells from diabetic patients restore diabetes-induced impairment of postischemic neovascularization. *Diabetes* 61(10):2621–2632. <https://doi.org/10.2337/db11-1768>
 1540. Zhu ML, Fan JX, Guo YQ, Guo LJ, Que HD, Cui BY, Li YL, Guo S, Zhang MX, Yin YL et al (2024) Protective effect of alizarin on vascular endothelial dysfunction via inhibiting the type 2 diabetes-induced synthesis of THBS1 and activating the AMPK signaling pathway. *Phytomedicine*. <https://doi.org/10.1016/j.phymed.2024.155557>
 1541. Nam SM, Kwon HJ, Kim W, Kim JW, Hahn KR, Jung HY, Kim DW, Yoo DY, Seong JK, Hwang IK et al (2018) Changes of myelin basic protein in the hippocampus of an animal model of type 2 diabetes. *Lab Anim Res* 34(4):176–184. <https://doi.org/10.5625/lar.2018.34.4.176>
 1542. Papadimitriou A, Peixoto EB, Silva KC, Lopes de Faria JM, Lopes de Faria JB (2014) Increase in AMPK brought about by cocoa is renoprotective in experimental diabetes mellitus by reducing NOX4/TGF β -1 signaling. *J Nutr Biochem* 25(7):773–784. <https://doi.org/10.1016/j.jnutbio.2014.03.010>
 1543. Hope CM, Welch J, Mohandas A, Pederson S, Hill D, Gundsambuu B, Eastaff-Leung N, Grosse R, Bresatz S, Ang G et al (2019) Peptidase inhibitor 16 identifies a human regulatory T-cell subset with reduced FOXP3 expression over the first year of recent onset type 1 diabetes. *Eur J Immunol* 49(8):1235–1250. <https://doi.org/10.1002/eji.201948094>
 1544. Wu C, Liu W, Liu Y, Xu T, Li M, Li X, Wang Y, Meng G, Li L, Zheng R et al (2023) Human umbilical cord mesenchymal stem cell-derived TGFB1 attenuates streptozotocin-induced type 1 diabetes mellitus by inhibiting T-cell proliferation. *Hum Cell* 36(3):997–1010. <https://doi.org/10.1007/s13577-023-00868-9>
 1545. Rodriguez S, Eiriksdottir G, Gaunt TR, Harris TB, Launer LJ, Gudnason V, Day IN (2010) IGF2BP1, IGF2BP2 and IGF2BP3 genotype, haplotype and genetic model studies in metabolic syndrome traits and diabetes. *Growth Horm IGF Res* 20(4):310–318. <https://doi.org/10.1016/j.ghir.2010.04.002>
 1546. Ghodsian N, Ismail P, Ahmadloo S, Heidari F, Haghvirdizadeh P, Ataollahi Eshkoor S, Etemad A (2016) Novel association of WNK4 gene, ala589ser

- polymorphism in essential hypertension, and type 2 diabetes mellitus in Malaysia. *J Diabetes Res* 2016:8219543. <https://doi.org/10.1155/2016/8219543>
1547. Yuan G, Liu Y, Sun T, Xu Y, Zhang J, Yang Y, Zhang M, Cianflone K, Wang DW (2011) The therapeutic role of very low-density lipoprotein receptor gene in hyperlipidemia in type 2 diabetic rats. *Hum Gene Ther* 22(3):302–312. <https://doi.org/10.1089/hum.2010.038>
 1548. Fawzy MS, Beladi FIA (2018) Association of circulating vitamin D, VDBP, and vitamin D receptor expression with severity of diabetic nephropathy in a group of Saudi type 2 diabetes mellitus patients. *Clin Lab* 64(10):1623–1633. <https://doi.org/10.7754/Clin.Lab.2018.180401>
 1549. Garranzo-Asensio M, Solís-Fernández G, Montero-Calle A, García-Martínez JM, Fiuza MC, Pallares P, Palacios-García N, García-Jiménez C, Guzman-Aranguéz A, Barderas R et al (2022) Seroreactivity against tyrosine phosphatase PTPRN links type 2 diabetes and colorectal cancer and identifies a potential diagnostic and therapeutic target. *Diabetes* 71(3):497–510. <https://doi.org/10.2337/db20-1206>
 1550. Puri S, Akiyama H, Hebrok M (2013) VHL-mediated disruption of Sox9 activity compromises β -cell identity and results in diabetes mellitus. *Genes Dev* 27(23):2563–2575. <https://doi.org/10.1101/gad.227785.113>
 1551. Adel H, Fawzy O, Mahmoud E, Mohammed NS, Khidr EG (2023) Inactive matrix Gla protein in relation to diabetic retinopathy in type 2 diabetes. *J Diabetes Metab Disord* 22(1):603–610. <https://doi.org/10.1007/s40200-022-01180-3>
 1552. Tajiri M, Nakahashi O, Kagawa T, Masuda M, Ohminami H, Iwano M, Takeda E, Taketani Y, Yamamoto H (2020) Association of increased renal Cyp24a1 gene expression with low plasma 1,25-dihydroxyvitamin D levels in rats with streptozotocin-induced diabetes. *J Clin Biochem Nutr* 66(1):49–56. <https://doi.org/10.3164/jcfn.19-79>
 1553. Ermiş Karaali Z, Candan G, Aktuğlu MB, Velet M, Ergen A (2019) Toll-like receptor 2 (TLR-2) gene polymorphisms in type 2 diabetes mellitus. *Cell J* 20(4):559–563. <https://doi.org/10.22074/cellj.2019.5540>
 1554. Rehman K, Akash MSH, Liaqat A, Kamal S, Qadir MI, Rasul A (2017) Role of interleukin-6 in development of insulin resistance and type 2 diabetes mellitus. *Crit Rev Eukaryot Gene Expr* 27(3):229–236. <https://doi.org/10.1615/CritRevEukaryotGeneExpr.2017019712>
 1555. Everett BM, Cook NR, Chasman DI, Magnone MC, Bobadilla M, Rifai N, Ridker PM, Pradhan AD (2013) Prospective evaluation of B-type natriuretic peptide concentrations and the risk of type 2 diabetes in women. *Clin Chem* 59(3):557–565. <https://doi.org/10.1373/clinchem.2012.194167>
 1556. Schneider A, Suman A, Rossi L, Barmada MM, Beglinger C, Parvin S, Sattar S, Ali L, Khan AK, Gyr N et al (2002) SPINK1/PSTI mutations are associated with tropical pancreatitis and type II diabetes mellitus in Bangladesh. *Gastroenterology* 123(4):1026–1030. <https://doi.org/10.1053/gast.2002.36059>
 1557. Fan H, Han J, Chen L, Feng B, Sun X, Shi B (2022) Association between plasma growth arrest-specific protein 6 and carotid atherosclerosis in type 2 diabetes mellitus. *Nutr Metab Cardiovasc Dis* 32(8):1917–1923. <https://doi.org/10.1016/j.numecd.2022.05.007>
 1558. Adedayo A, Eluwole A, Tedla F, Kremer A, Khan M, Mastrogianni N, Rosenberg C, Dreizen P, La Rosa J, Saliccioli L et al (2022) Relationship between the soluble F11 receptor and annexin A5 in African Americans patients with type-2 diabetes mellitus. *Biomedicine* 10(8):1818. <https://doi.org/10.3390/biomedicine10081818>
 1559. Cheng W, Cai C, Xu Y, Xiao X, Shi T, Liao Y, Wang X, Chen S, Zhou M, Liao Z (2023) TRIM21-FOXO1-BCL-2 axis underlies hyperglycaemic cell death and diabetic tissue damage. *Cell Death Dis* 14(12):825. <https://doi.org/10.1038/s41419-023-06355-1>
 1560. Lamin V, Verry J, Eigner-Bybee I, Fuqua JD, Wong T, Lira VA, Dokun AO (2021) Modulation of miR-29a and ADAM12 reduces post-ischemic skeletal muscle injury and improves perfusion recovery and skeletal muscle function in a mouse model of type 2 diabetes and peripheral artery disease. *Int J Mol Sci* 23(1):429. <https://doi.org/10.3390/ijms23010429>
 1561. Wang FF, Zhang JL, Ji Y, Yan XJ, Sun L, Zhu Y, Jin H (2022) KLF2 mediates the suppressive effect of BDNF on diabetic intimal calcification by inhibiting HK1 induced endothelial-to-mesenchymal transition. *Cell Signal* 94:110324. <https://doi.org/10.1016/j.cellsig.2022.110324>
 1562. Durrani IA, Bhatti A, John P (2023) Integrated bioinformatics analyses identifying potential biomarkers for type 2 diabetes mellitus and breast cancer: in SIK1-ness and health. *PLoS ONE* 18(8):0289839
 1563. Alwahsh SM, Qutachi O, Starkey Lewis PJ, Bond A, Noble J, Burgoyne P, Morton N, Carter R, Mann J, Ferreira-Gonzalez S et al (2021) Fibroblast growth factor 7 releasing particles enhance islet engraftment and improve metabolic control following islet transplantation in mice with diabetes. *Am J Transplant* 21(9):2950–2963. <https://doi.org/10.1111/ajt.16488>
 1564. Kang YE, Choung S, Lee JH, Kim HJ, Ku BJ (2017) The role of circulating Slit2, the one of the newly batokines. *Human Diabetes Mellitus. Endocrinol Metab (Seoul)* 32(3):383–388. <https://doi.org/10.3803/EnM.2017.32.3.383>
 1565. Ragvin A, Moro E, Fredman D, Navratilova P, Drivenes Ø, Engström PG, Alonso ME, de la Calle ME, Gómez Skarmeta JL, Tavares MJ et al (2010) Long-range gene regulation links genomic type 2 diabetes and obesity risk regions to HHEX, SOX4, and IRX3. *Proc Natl Acad Sci USA* 107(2):775–780. <https://doi.org/10.1073/pnas.0911591107>
 1566. Zhang F, Han Y, Zheng L, Bao Z, Liu L, Li W (2024) Association between chitinase-3-like protein 1 and metabolic-associated fatty liver disease in patients with type 2 diabetes mellitus. *Ir J Med Sci*. <https://doi.org/10.1007/s11845-024-03671-z>
 1567. Zhong Y, Du G, Liu J, Li S, Lin J, Deng G, Wei J, Huang J (2022) RUNX1 and CCL3 in diabetes mellitus-related coronary artery disease: a bioinformatics analysis. *Int J Gen Med* 15:955–963. <https://doi.org/10.2147/IJGM.S350732>
 1568. Elsehrawy AAEW, El-Toukhy SE, Seliem NMA, Moustafa RS, Mohammed DS (2019) Apelin and chemerin as promising adipokines in children with type 1 diabetes mellitus. *Diabetes Metab Syndr Obes* 12:383–389. <https://doi.org/10.2147/DMSO.S189264>
 1569. Li Z, Sun Z, Chang D, Zhu L, Chen M, Zhao M (2022) Association between COL4A3 variant rs55703767 and susceptibility to diabetic kidney disease in patients with type 2 diabetes mellitus: results from the INDEED cohort study. *Chin Med J (Engl)* 135(9):1129–1130. <https://doi.org/10.1097/CM9.0000000000001955>
 1570. Al-Awaideh WJ, Hameed WS, Al-Hassany HJ, Al-Dabet MM, Al-Bawareed O, Hadi NR (2021) Evaluation of the Genetic association and expressions of Notch-2 /Jagged-1 in patients with Type 2 diabetes mellitus. *Med Arch* 75(2):101–108. <https://doi.org/10.5455/medarh.2021.75.101-108>
 1571. Vandekerckhove L, Vermeulen Z, Liu ZZ, Boimvaser S, Patzak A, Segers VF, De Keulenaer GW (2016) Neuregulin-1 attenuates development of nephropathy in a type 1 diabetes mouse model with high cardiovascular risk. *Am J Physiol Endocrinol Metab* 310(7):E495–E504. <https://doi.org/10.1152/ajpendo.00432.2015>
 1572. Tao J, Yu XL, Yuan YJ, Shen X, Liu J, Gu PP, Wang Z, Ma YT, Li GQ (2022) DMRT2 interacts with FXR and improves insulin resistance in adipocytes and a mouse model. *Front Endocrinol (Lausanne)* 12:723623. <https://doi.org/10.3389/fendo.2021.723623>
 1573. Choong YS, Lim YY, Soong JX, Savoo N, Guida C, Rhyman L, Ramracheya R, Ramasami P (2021) Theoretical study of the interactions between peptide tyrosine tyrosine [PYY (1–36)], a newly identified modulator in type 2 diabetes pathophysiology, with receptors NPY1R and NPY4R. *Hormones (Athens)* 20(3):557–569. <https://doi.org/10.1007/s42000-021-00278-2>
 1574. Pfeifer CW, Walsh JT, Santeford A, Lin JB, Beatty WL, Terao R, Liu YA, Hase K, Ruzycski PA, Apte RS (2023) Dysregulated CD200-CD200R signaling in early diabetes modulates microglia-mediated retinopathy. *Proc Natl Acad Sci USA* 120(45):e2308214120. <https://doi.org/10.1073/pnas.2308214120>
 1575. Zhou N, Liu W, Zhang W, Liu Y, Li X, Wang Y, Zheng R, Zhang Y (2021) Wip1 regulates the immunomodulatory effects of murine mesenchymal stem cells in type 1 diabetes mellitus via targeting IFN- α /BST2. *Cell Death Discov* 7(1):326. <https://doi.org/10.1038/s41420-021-00728-1>
 1576. Guo M, Xia Z, Hong Y et al (2023) The TFPI2-PPAR γ axis induces M2 polarization and inhibits fibroblast activation to promote recovery from post-myocardial infarction in diabetic mice. *J Inflamm (Lond)* 20(1):35. <https://doi.org/10.1186/s12950-023-00357-8>
 1577. Bhoj EJ, Romeo S, Baroni MG, Bartov G, Schultz RA, Zinn AR (2009) MODY-like diabetes associated with an apparently balanced translocation: possible involvement of MPP7 gene and cell polarity in the

- pathogenesis of diabetes. *Mol Cytogenet* 2:5. <https://doi.org/10.1186/1755-8166-2-5>
1578. Crocco P, Dato S, Montesanto A, Bonfigli AR, Testa R, Olivieri F, Passarino G, Rose G (2022) The genetic variability of members of the SLC38 family of amino acid transporters (SLC38A3, SLC38A7 and SLC38A9) affects susceptibility to type 2 diabetes and vascular complications. *Nutrients* 14(21):4440. <https://doi.org/10.3390/nu14214440>
 1579. Chen M, Lin WR, Lu CH, Chen CC, Huang YC, Liao WL, Tsai FJ (2014) Chimerin 2 genetic polymorphisms are associated with non-proliferative diabetic retinopathy in Taiwanese type 2 diabetic patients. *J Diabetes Complications* 28(4):460–463. <https://doi.org/10.1016/j.jdiacomp.2014.04.009>
 1580. Luo LG, Jackson I (2007) Thyrotropin releasing hormone (TRH) may preserve pancreatic islet cell function: potential role in the treatment of diabetes mellitus. *Acta Biomed* 78(Suppl 1):216–221
 1581. Barry DM, Liu XT, Liu B, Liu XY, Gao F, Zeng X, Liu J, Yang Q, Wilhelm S et al (2020) Exploration of sensory and spinal neurons expressing gastrin-releasing peptide in itch and pain related behaviors. *Nat Commun* 11(1):1397. <https://doi.org/10.1038/s41467-020-15230-y>
 1582. Powell-Roach KL, Yao Y, Cao X, Chamala S, Wallace MR, Cruz-Almeida Y, Molokie RE, Wang ZJ, Wilkie DJ (2023) Analysis of AVPR1A, thermal and pressure pain thresholds, and stress in sickle cell disease. *Front Pain Res (Lausanne)* 3:1060245. <https://doi.org/10.3389/fpain.2022.1060245>
 1583. Wang Y, Hu X, Huang H, Jin Z, Gao J, Guo Y, Zhong Y, Li Z, Zong X, Wang K et al (2022) Optimization of 4-arylthiophene-3-carboxylic acid derivatives as inhibitors of ANO1: lead optimization studies toward their analgesic efficacy for inflammatory pain. *Eur J Med Chem* 237:114413. <https://doi.org/10.1016/j.ejmech.2022.114413>
 1584. Echaniz-Laguna A, Altuzarra C, Verloes A, De La Banda MGG, Quijano-Roy S, Tudorache RA, Jaxybayeva A, Myrzaliyeva B, Tazir M, Vallat JM et al (2021) NTRK1 gene-related congenital insensitivity to pain with anhidrosis: a nationwide multicenter retrospective study. *Neurogenetics* 22(4):333–341. <https://doi.org/10.1007/s10048-021-00668-z>
 1585. Bian J, Zhang B, Zhang Y, Tian Y, Yin L, Zou W (2022) FGF 10 inhibited spinal microglial activation in neuropathic pain via PPAR- γ /NF- κ B signaling. *Neuroscience* 500:52–62. <https://doi.org/10.1016/j.neuroscience.2022.07.033>
 1586. Husain SF, Lam RWM, Hu T, Ng MWF, Liao ZQ, Nagata K, Khanna S, Lam Y, Bhakoo K, Ho RCM et al (2019) Locating the site of neuropathic pain in vivo using MMP-12-targeted magnetic nanoparticles. *Pain Res Manag* 2019:9394715. <https://doi.org/10.1155/2019/9394715>
 1587. Niederberger E, Möller M, Mungo E, Hass M, Wilken-Schmitz A, Mander-scheid C, Möser CV, Geisslinger G (2023) Distinct molecular mechanisms contribute to the reduction of melanoma growth and tumor pain after systemic and local depletion of alpha-Synuclein in mice. *FASEB J* 37(12):e23287. <https://doi.org/10.1096/fj.202301489R>
 1588. Pawlik K, Ciapała K, Ciechanowska A, Kwiatkowski K, Mika J (2022) Pharmacological evidence of the important roles of CCR1 and CCR3 and their endogenous ligands CCL2/7/8 in hypersensitivity based on a murine model of neuropathic pain. *Cells* 12(1):98. <https://doi.org/10.3390/cells12010098>
 1589. Das V, Kc R, Li X et al (2018) Blockade of vascular endothelial growth factor receptor-1 (Flt-1), reveals a novel analgesic for osteoarthritis-induced joint pain. *Gene Rep* 11:94–100. <https://doi.org/10.1016/j.genrep.2018.03.008>
 1590. Gordon C, Trainor J, Shah RJ, Studholme K, Gelman A, Doswell F, Sadar F, Giovannetti A, Gershenson J, Khan A et al (2024) Fatty acid binding protein 5 inhibition attenuates pronociceptive cytokine/chemokine expression and suppresses osteoarthritis pain: a comparative human and rat study. *Osteoarthritis Cartilage* 32(3):266–280. <https://doi.org/10.1016/j.joca.2023.11.010>
 1591. Aoki Y, Nishizawa D, Ohka S, Kasai S, Arita H, Hanaoka K, Yajima C, Iseki M, Kato J, Ogawa S et al (2023) Rs11726196 single-nucleotide polymorphism of the transient receptor potential canonical 3 (TRPC3) gene is associated with chronic pain. *Int J Mol Sci* 24(2):1028. <https://doi.org/10.3390/ijms24021028>
 1592. Liu C, Sun Q, Xu J, Shen W, Li H, Yang L (2022) The role of bone morphogenetic protein 4 in microglial polarization in the process of neuropathic pain. *J Inflamm Res* 15:2803–2817. <https://doi.org/10.2147/JIR.S356531>
 1593. Mecklenburg J, Wangzhou A, Hovhannisyian AH, Barba-Escobedo P, Shein SA, Zou Y, Weldon K, Lai Z, Goffin V, Dussor G et al (2022) Sex-dependent pain trajectories induced by prolactin require an inflammatory response for pain resolution. *Brain Behav Immun* 101:246–263. <https://doi.org/10.1016/j.bbi.2022.01.016>
 1594. Liu X, Bae C, Gelman BB, Chung JM, Tang SJ (2022) A neuron-to-astrocyte Wnt5a signal governs astrogliosis during HIV-associated pain pathogenesis. *Brain* 145(11):4108–4123. <https://doi.org/10.1093/brain/awac015>
 1595. He M, Pang J, Sun H, Zheng G, Lin Y, Ge W (2020) Overexpression of TIMP3 inhibits discogenic pain by suppressing angiogenesis and the expression of substance P in nucleus pulposus. *Mol Med Rep* 21(3):1163–1171. <https://doi.org/10.3892/mmr.2020.10922>
 1596. Lien WC, Zhou XR, Liang YJ, Ching CT, Wang CY, Lu FI, Chang HC, Lin FH, Wang HD (2022) Therapeutic potential of nanoceria pretreatment in preventing the development of urological chronic pelvic pain syndrome: Immunomodulation via reactive oxygen species scavenging and SerpinB2 downregulation. *Bioeng Transl Med* 8(1):e10346. <https://doi.org/10.1002/btm2.10346>
 1597. Verma V, Khoury S, Parisien M, Cho C, Maixner W, Martin LJ, Diatchenko L (2020) The dichotomous role of epiregulin in pain. *Pain* 161(5):1052–1064. <https://doi.org/10.1097/j.pain.0000000000001792>
 1598. Sikandar S, Minett MS, Millet Q, Santana-Varela S, Lau J, Wood JN, Zhao J (2018) Brain-derived neurotrophic factor derived from sensory neurons plays a critical role in chronic pain. *Brain* 141(4):1028–1039. <https://doi.org/10.1093/brain/awy009>
 1599. Wan J, Nan S, Liu J, Ding M, Zhu H, Suo C, Wang Z, Hu M, Wang D, Ding Y (2020) Synaptotagmin 1 is involved in neuropathic pain and electroacupuncture-mediated analgesic effect. *Int J Mol Sci* 21(3):968. <https://doi.org/10.3390/ijms21030968>
 1600. Zahr NM, Sullivan EV, Pohl KM, Pfefferbaum A, Saranathan M (2020) Sensitivity of ventrolateral posterior thalamic nucleus to back pain in alcoholism and CD4 nadir in HIV. *Hum Brain Mapp* 41(5):1351–1361. <https://doi.org/10.1002/hbm.24880>
 1601. Braden K, Giancotti LA, Chen Z, DeLeon C, Latzo N, Boehn T, D’Cunha N, Thompson BM, Doyle TM, McDonald JG et al (2020) GPR183-oxysterol axis in spinal cord contributes to neuropathic pain. *J Pharmacol Exp Ther* 375(2):367–375. <https://doi.org/10.1124/jpet.120.000105>
 1602. Monteiro S, de Araujo D, Nassini R, Geppetti P, De Logu F (2020) TRPA1 as a therapeutic target for nociceptive pain. *Expert Opin Ther Targets* 24(10):997–1008. <https://doi.org/10.1080/14728222.2020.1815191>
 1603. Zhang X, Xia L, Xie A, Liao O, Ju F, Zhou Y (2021) Low concentration of Bupivacaine ameliorates painful diabetic neuropathy by mediating miR-23a/PDE4B axis in microglia. *Eur J Pharmacol* 891:173719. <https://doi.org/10.1016/j.ejphar.2020.173719>
 1604. Ebbinghaus M, Jenei-Lanzl Z, Segond von Banchet G, Stangl H, Gajda M, Straub RH, Schaible HG (2018) A promising new approach for the treatment of inflammatory pain: transfer of stem cell-derived tyrosine hydroxylase-positive cells. *NeuroImmunoModulation* 25(4):225–237. <https://doi.org/10.1159/000495349>
 1605. Tajerian M, Clark JD (2019) Spinal matrix metalloproteinase 8 regulates pain after peripheral trauma. *J Pain Res* 12:1133–1138. <https://doi.org/10.2147/JPR.S197761>
 1606. Chen N, Ge MM, Li DY, Wang XM, Liu DQ, Ye DW, Tian YK, Zhou YQ, Chen JP (2021) β 2-adrenoreceptor agonist ameliorates mechanical allodynia in paclitaxel-induced neuropathic pain via induction of mitochondrial biogenesis. *Biomed Pharmacother* 144:112331. <https://doi.org/10.1016/j.biopha.2021.112331>
 1607. Zareba P, Gryzlo B, Malawska K, Sałat K, Höfner GC, Nowaczyk A, Fijałkowski Ł, Rapacz A, Podkowa A, Furgala A et al (2020) Novel mouse GABA uptake inhibitors with enhanced inhibitory activity toward mGAT3/4 and their effect on pain threshold in mice. *Eur J Med Chem* 188:111920. <https://doi.org/10.1016/j.ejmech.2019.111920>
 1608. Montera MA, Goins AE, Alles SRA, Westlund KN (2023) Urokinase-type plasminogen activator-induced mouse back pain model. *J Vis Exp*. <https://doi.org/10.3791/63997>. doi:10.3791/63997
 1609. Inoue R, Nishizawa D, Hasegawa J, Nakayama K, Fukuda KI, Ichinohe T, Mieda T, Tsujita M, Nakagawa H, Kitamura A et al (2021) Effects of rs958804 and rs7858836 single-nucleotide polymorphisms of the ASTN2 gene on pain-related phenotypes in patients who underwent laparoscopic colectomy and mandibular sagittal split ramus osteotomy.

- Neuropsychopharmacol Rep 41(1):82–90. <https://doi.org/10.1002/npr.12159>
1610. Sun JM, Sun LZ, Liu J, Su BH, Shi L (2013) Serum interleukin-15 levels are associated with severity of pain in patients with knee osteoarthritis. *Dis Markers* 35(3):203–206. <https://doi.org/10.1155/2013/176278>
1611. Yang L, Liu S, Wang Y (2019) Role of bone morphogenetic protein-2/4 in astrocyte activation in neuropathic pain. *Mol Pain* 15:1744806919892100. <https://doi.org/10.1177/1744806919892100>
1612. Liu S, Yang S, Zhu X, Li X, Zhang X, Zhou X, Cheng H, Huo FQ, Mao Q, Liang L (2023) Spinal apolipoprotein E is involved in inflammatory pain via regulating lipid metabolism and glial activation in the spinal dorsal horn. *Biol Direct* 18(1):85. <https://doi.org/10.1186/s13062-023-00444-z>
1613. Kruse LS, Møller M, Tibaek M, Gammeltoft S, Olesen J, Kruuse C (2009) PDE9A, PDE10A, and PDE11A expression in rat trigeminovascular pain signalling system. *Brain Res* 1281:25–34. <https://doi.org/10.1016/j.brainres.2009.05.012>
1614. Fang Y, Cui H, Liu F, Su S, Wang T, Yuan B, Xie Y, Ma C (2022) Astrocytic phosphatase and tensin homolog deleted on chromosome 10 regulates neuropathic pain by facilitating 3-hydroxy-3-methylglutaryl-CoA reductase-dependent cholesterol biosynthesis. *Pain* 163(12):e1192–e1206. <https://doi.org/10.1097/j.pain.0000000000002682>
1615. Hühne K, Leis S, Schmelz M, Rautenstrauss B, Birklein F (2004) A polymorphic locus in the intron 16 of the human angiotensin-converting enzyme (ACE) gene is not correlated with complex regional pain syndrome I (CRPS I). *Eur J Pain* 8(3):221–225. <https://doi.org/10.1016/j.ejpain.2003.08.004>
1616. Zhang M, Yang K, Wang QH, Xie L, Liu Q, Wei R, Tao Y, Zheng HL, Lin N, Xu H et al (2023) The cytidine N-acetyltransferase NAT10 participates in peripheral nerve injury-induced neuropathic pain by stabilizing SYT9 expression in primary sensory neurons. *J Neurosci* 43(17):3009–3027. <https://doi.org/10.1523/JNEUROSCI.2321-22.2023>
1617. Wang J, Zhao M, Jia P, Liu FF, Chen K, Meng FY, Hong JH, Zhang T, Jin XH, Shi J (2020) The analgesic action of laryxyl acetate, a potent TRPC6 inhibitor, in rat neuropathic pain model induced by spared nerve injury. *J Neuroinflamm* 17(1):118. <https://doi.org/10.1186/s12974-020-01767-8>
1618. Ciecchanowska A, Rojewska E, Piotrowska A, Barut J, Pawlik K, Ciapała K, Kreiner G, Mika J (2022) New insights into the analgesic properties of the XCL1/XCR1 and XCL1/ITGA9 axes modulation under neuropathic pain conditions - evidence from animal studies. *Front Immunol* 13:1058204. <https://doi.org/10.3389/fimmu.2022.1058204>
1619. Wack G, Metzner K, Kuth MS, Wang E, Bresnick A, Brandes RP, Schröder K, Wittig I, Schmidtko A, Kallenborn-Gerhardt W (2021) Nox4-dependent upregulation of S100A4 after peripheral nerve injury modulates neuropathic pain processing. *Free Radic Biol Med* 168:155–167. <https://doi.org/10.1016/j.freeradbiomed.2021.03.021>
1620. Singh S, Kartha S, Bulka BA, Stiansen NS, Winkelstein BA (2019) Physiologic facet capsule stretch can induce pain & upregulate matrix metalloproteinase-3 in the dorsal root ganglia when preceded by a physiological mechanical or nonpainful chemical exposure. *Clin Biomech (Bristol, Avon)* 64:122–130. <https://doi.org/10.1016/j.clinbiomech.2018.01.009>
1621. Merighi A (2016) Targeting the glial-derived neurotrophic factor and related molecules for controlling normal and pathologic pain. *Expert Opin Ther Targets* 20(2):193–208. <https://doi.org/10.1517/14728222.2016.1085972>
1622. Fattori V, Staurengo-Ferrari L, Zaninelli TH, Casagrande R, Oliveira RD, Louzada-Junior P, Cunha TM, Alves-Filho JC, Teixeira MM, Cunha FQ et al (2020) IL-33 enhances macrophage release of IL-1 β and promotes pain and inflammation in gouty arthritis. *Inflamm Res* 69(12):1271–1282. <https://doi.org/10.1007/s00011-020-01399-x>
1623. Barker PA, Mantyh P, Arendt-Nielsen L, Viktrup L, Tive L (2020) Nerve growth factor signaling and its contribution to pain. *J Pain Res* 13:1223–1241. <https://doi.org/10.2147/JPR.S247472>
1624. Kluzek S, Bay-Jensen AC, Judge A, Karsdal MA, Shorthose M, Spector T, Hart D, Newton JL, Arden NK (2015) Serum cartilage oligomeric matrix protein and development of radiographic and painful knee osteoarthritis. A community-based cohort of middle-aged women. *Biomarkers* 20(8):557–564. <https://doi.org/10.3109/1354750X.2015.1105498>
1625. Perera RS, Dissanayake PH, Senarath U, Wijayaratne LS, Karunanayake AL, Dissanayake VHW (2017) Variants of ACAN are associated with severity of lumbar disc herniation in patients with chronic low back pain. *PLoS ONE* 12(7):e0181580. <https://doi.org/10.1371/journal.pone.0181580>
1626. Rausch SM, Gonzalez BD, Clark MM, Patten C, Felten S, Liu H, Li Y, Sloan J, Yang P (2012) SNPs in PTGS2 and LTA predict pain and quality of life in long term lung cancer survivors. *Lung Cancer* 77(1):217–223. <https://doi.org/10.1016/j.lungcan.2012.02.01>
1627. Jiang L, Tan B, Li S, Wang L, Zheng L, Liu Y, Long Z, Wu Y (2017) Decrease of growth and differentiation factor 10 contributes to neuropathic pain through N-methyl-D-aspartate receptor activation. *NeuroReport* 28(8):444–450. <https://doi.org/10.1097/WNR.0000000000000785>
1628. Ding S, Zhu T, Tian Y, Xu P, Chen Z, Huang X, Zhang X (2018) Role of brain-derived neurotrophic factor in endometriosis pain. *Reprod Sci* 25(7):1045–1057. <https://doi.org/10.1177/1933719117732161>
1629. Ma LT, Bai Y, Li J, Qiao Y, Liu Y, Zheng J (2021) Elemene emulsion injection administration reduces neuropathic pain by inhibiting astrocytic NDRG2 expression within spinal dorsal horn. *Chin J Integr Med* 27(12):912–918. <https://doi.org/10.1007/s11655-021-3438-3>
1630. Liu B, Li N, He Z, Zhang X, Duan G (2021) Emerging role of serum glucocorticoid-regulated kinase 1 in pathological pain. *Front Mol Neurosci* 14:683527. <https://doi.org/10.3389/fnmol.2021.683527>
1631. Zhang X, Gao R, Zhang C, Teng Y, Chen H, Li Q, Liu C, Wu J, Wei L, Deng L et al (2023) Extracellular RNAs-TLR3 signaling contributes to cognitive impairment after chronic neuropathic pain in mice. *Signal Transduct Target Ther* 8(1):292. <https://doi.org/10.1038/s41392-023-01543-z>
1632. Zhao J, Yuan G, Cendan CM, Nassar MA, Lagerström MC, Kullander K, Gavazzi I, Wood JN (2010) Nociceptor-expressed ephrin-B2 regulates inflammatory and neuropathic pain. *Mol Pain* 6:77. <https://doi.org/10.1186/1744-8069-6-77>
1633. Shubayev VI, Strongin AY, Yaksh TL (2018) Structural homology of myelin basic protein and muscarinic acetylcholine receptor: significance in the pathogenesis of complex regional pain syndrome. *Mol Pain* 14:1744806918815005. <https://doi.org/10.1177/1744806918815005>
1634. Yoon SY, Kwon SG, Kim YH, Yeo JH, Ko HG, Roh DH, Kaang BK, Beitz AJ, Lee JH, Oh SB (2017) A critical role of spinal Shank2 proteins in NMDA-induced pain hypersensitivity. *Mol Pain* 13:1744806916688902. <https://doi.org/10.1177/1744806916688902>
1635. Garrity R, Arora N, Haque MA, Weis D, Trinh RT, Neerukonda SV, Kumari S, Cortez I, Ubogu EE, Mahalingam R et al (2023) Fibroblast-derived PI16 sustains inflammatory pain via regulation of CD206+ myeloid cells. *Brain Behav Immun* 112:220–234. <https://doi.org/10.1016/j.bbi.2023.06.011>
1636. Zhang G, Zhou Y, Su M, Yang X, Zeng B (2020) Inhibition of microRNA-27b-3p relieves osteoarthritis pain via regulation of KDM4B-dependent DLX5. *BioFactors* 46(5):788–802. <https://doi.org/10.1002/biof.1670>
1637. Celikbilek A (2020) Possible associations of vitamin D, vitamin D-binding protein, and vitamin D receptor with diabetic neuropathic pain and balance [Letter]. *J Pain Res* 13:465–466. <https://doi.org/10.2147/JPR.S249871>
1638. Habib AM, Matsuyama A, Okorokov AL, Santana-Varela S, Bras JT, Aloisi AM, Emery EC, Bogdanov YD, Follenfant M, Gossage SJ et al (2018) A novel human pain insensitivity disorder caused by a point mutation in ZFH2. *Brain* 141(2):365–376. <https://doi.org/10.1093/brain/awx326>
1639. Kim MJ, Son JY, Ju JS, Ahn DK (2020) Early blockade of EphA4 pathway reduces trigeminal neuropathic pain. *J Pain Res* 13:1173–1183. <https://doi.org/10.2147/JPR.S249185>
1640. Trivedi MK, Mondal S, Gangwar M, Jana S (2023) Effects of cannabidiol interactions with CYP2R1, CYP27B1, CYP24A1, and vitamin D3 receptors on spatial memory, pain, inflammation, and aging in vitamin D3 deficiency diet-induced rats. *Cannabis Cannabinoid Res* 8(6):1019–1029. <https://doi.org/10.1089/can.2021.0240>
1641. Jiang L, Wu Q, Yang T (2016) Silencing of Id2 alleviates chronic neuropathic pain following chronic constriction injury. *J Mol Neurosci* 59(1):99–105. <https://doi.org/10.1007/s12031-016-0713-z>
1642. Gao X, Gao LF, Zhang YN, Kong XQ, Jia S, Meng CY (2023) Huc-MSCs-derived exosomes attenuate neuropathic pain by inhibiting activation of the TLR2/MyD88/NF- κ B signaling pathway in the spinal microglia by targeting Rsad2. *Int Immunopharmacol* 114:109505. <https://doi.org/10.1016/j.intimp.2022.109505>
1643. Atzeni F, Nucera V, Masala IF, Sarzi-Puttini P, Bonitta G (2019) IL-6 Involvement in pain, fatigue and mood disorders in rheumatoid arthritis

- and the effects of IL-6 inhibitor sarilumab. *Pharmacol Res* 149:104402. <https://doi.org/10.1016/j.phrs.2019.104402>
1644. Tremblay K, Dubois-Bouchard C, Brisson D, Gaudet D (2014) Association of CTRC and SPINK1 gene variants with recurrent hospitalizations for pancreatitis or acute abdominal pain in lipoprotein lipase deficiency. *Front Genet* 5:90. <https://doi.org/10.3389/fgene.2014.00090>
1645. Fan W, Liu C, Chen D, Xu C, Qi X, Zhang A, Zhu X, Liu Y, Wang L, Hao L et al (2023) Ozone alleviates MSU-induced acute gout pain via upregulating AMPK/GAS6/MerTK/SOCS3 signaling pathway. *J Transl Med* 21(1):890. <https://doi.org/10.1186/s12967-023-04769-1>
1646. Zhang J, Zhao H, Zhang A, Zhao C, Mei Z, Yao H, Fan Z, Liang D (2022) Identifying a novel KLF2/lncRNA SNHG12/miR-494-3p/RAD23B axis in Spare Nerve Injury-induced neuropathic pain. *Cell Death Discov* 8(1):272. <https://doi.org/10.1038/s41420-022-01060-y>
1647. Zhang J, Mei Z, Yao W, Zhao C, Wu S, Ouyang J (2023) SIX1 induced HULC modulates neuropathic pain and Schwann cell oxidative stress after sciatic nerve injury. *Gene* 882:147655. <https://doi.org/10.1016/j.gene.2023.147655>
1648. Yamanaka H, Kobayashi K, Okubo M, Fukuoka T, Noguchi K (2011) Increase of close homolog of cell adhesion molecule L1 in primary afferent by nerve injury and the contribution to neuropathic pain. *J Comp Neurol* 519(8):1597–1615. <https://doi.org/10.1002/cne.22588>
1649. Ke C, Gao F, Tian X, Li C, Shi D, He W, Tian Y (2017) Slit2/Robo1 mediation of synaptic plasticity contributes to bone cancer pain. *Mol Neurobiol* 54(1):295–307. <https://doi.org/10.1007/s12035-015-9564-9>
1650. Yuan ZL, Liu XD, Zhang ZX, Li S, Tian Y, Xi K, Cai J, Yang XM, Liu M, Xing GG (2022) Activation of GDNF-ERK-Runx1 signaling contributes to P2X3R gene transcription and bone cancer pain. *iScience* 25(9):104936. <https://doi.org/10.1016/j.isci.2022.104936>
1651. Deng J, Liang J, Cao Y, Tong X, Li H (2024) CCL2- and Notch2-mediated central sensitization in a rat chronic pelvic pain model. *In Vivo* 38(1):205–212. <https://doi.org/10.121873/invivo.13427>
1652. Heesch C, Dimmeler S, Fichtlscherer S, Hamm CW, Berger J, Simoons ML, Zeiher AM (2004) Prognostic value of placental growth factor in patients with acute chest pain. *JAMA* 291(4):435–441. <https://doi.org/10.1001/jama.291.4.435>
1653. Wan C, Xu Y, Cen B, Xia Y, Yao L, Zheng Y, Zhao J, He S, Chen Y (2021) Neuregulin1-ErbB4 signaling in spinal cord participates in electroacupuncture analgesia in inflammatory pain. *Front Neurosci* 15:636348. <https://doi.org/10.3389/fnins.2021.636348>
1654. Zhang D, Zhao W, Liu J, Ou M, Liang P, Li J, Chen Y, Liao D, Bai S, Shen J et al (2021) Sodium leak channel contributes to neuronal sensitization in neuropathic pain. *Prog Neurobiol* 202:102041. <https://doi.org/10.1016/j.pneurobio.2021.102041>
1655. Kühlein HN, Tegeder I, Möser C, Lim HY, Häussler A, Spieth K, Jennes I, Marschalek R, Beckhaus T, Karas M et al (2011) Nerve injury evoked loss of latexin expression in spinal cord neurons contributes to the development of neuropathic pain. *PLoS ONE* 6(4):e19270. <https://doi.org/10.1371/journal.pone.0019270>
1656. Behbehani MM, Zemlan FP (1990) Bulbosplinal and intraspinal thyrotropin releasing hormone systems: modulation of spinal cord pain transmission. *Neuropeptides* 15(3):161–168. [https://doi.org/10.1016/0143-4179\(90\)90149-s](https://doi.org/10.1016/0143-4179(90)90149-s)
1657. Kleemann M, Schneider H, Unger K, Sander P, Schneider EM, Fischer-Posovszky P, Handrick R, Otte K (2018) MiR-744-5p inducing cell death by directly targeting HNRNPC and NFIX in ovarian cancer cells. *Sci Rep* 8(1):9020. <https://doi.org/10.1038/s41598-018-27438-6>
1658. Huang L, Schauer IG, Zhang J, Mercado-Urbe I, Deavers MT, Huang J, Liu J (2011) The oncogenic gene fusion TMPRSS2: ERG is not a diagnostic or prognostic marker for ovarian cancer. *Int J Clin Exp Pathol* 4(7):644–650
1659. Zhang Z, Chen F, Li S, Guo H, Xi H, Deng J, Han Q, Zhang W (2020) ERG the modulates Warburg effect and tumor progression in cervical cancer. *Biochem Biophys Res Commun* 522(1):191–197. <https://doi.org/10.1016/j.bbrc.2019.11.079>
1660. Landi S, Giannetti F, Benzoni P, Camprostrini G, Rossi G, Piantoni C, Bertoli G, Bonfanti C, Carnevali L, Bucchi A et al (2023) Lack of the transcription factor Nfix causes tachycardia in mice sinus node and rats neonatal cardiomyocytes. *Acta Physiol (Oxf)* 239(2):e13981. <https://doi.org/10.1111/apha.13981>
1661. Sperone A, Dryden NH, Birdsey GM, Madden L, Johns M, Evans PC, Mason JC, Haskard DO, Boyle JJ, Paleolog EM et al (2011) The transcription factor Erg inhibits vascular inflammation by repressing NF-kappaB activation and proinflammatory gene expression in endothelial cells. *Arterioscler Thromb Vasc Biol* 31(1):142–150. <https://doi.org/10.1161/ATVBAHA.110.216473>
1662. Yang L, Yang Z, Yao R, Li Y, Liu Z, Chen X, Zhang G (2018) miR-210 promotes progression of endometrial carcinoma by regulating the expression of NFIX. *Int J Clin Exp Pathol* 11(1):5213–5222
1663. Chen L, Lai L, Zheng L, Wang Y, Lu H, Chen Y (2024) Construction of an exosome-associated miRNA-mRNA regulatory network and validation of FYCO1 and miR-17-5p as potential biomarkers associated with ovarian cancer. *Transl Cancer Res* 13(2):1052–1067. <https://doi.org/10.21037/tcr-23-940>
1664. Aman P, Pejovic T, Wennborg A, Heim S, Mitelman F (1993) Mapping of the 19p13 breakpoint in an ovarian carcinoma between the INSR and TCF3 loci. *Genes Chromosomes Cancer* 8(2):134–136. <https://doi.org/10.1002/gcc.2870080212>
1665. Chen Y, Cao XY, Li YN, Qiu YY, Li YN, Li W, Wang H (2018) Reversal of cisplatin resistance by microRNA-139-5p-independent RNF2 downregulation and MAPK inhibition in ovarian cancer. *Am J Physiol Cell Physiol* 315(2):C225–C235. <https://doi.org/10.1152/ajpcell.00283.2017>
1666. Sun Y, Jin L, Sui YX, Han LL, Liu JH (2017) Circadian gene CLOCK affects drug-resistant gene expression and cell proliferation in ovarian cancer SKOV3/DDP cell lines through autophagy. *Cancer Biother Radiopharm* 32(4):139–146. <https://doi.org/10.1089/cbr.2016.2153>
1667. Herold N, Schmollinger J, Ernst C, Ataseven B, Blümcke B, Schömig-Markiefka B, Heikaus S, Göhring UJ, Engel C, Lampe B et al (2023) Pathogenic germline variants in SMARCA4 and further cancer predisposition genes in early onset ovarian cancer. *Cancer Med* 12(14):15256–15260. <https://doi.org/10.1002/cam4.6214>
1668. Zhang F, Zhu T, Wu C, Shen D, Liu L, Chen X, Guan Y, Ding H, Tong X (2023) TRIM28 recruits E2F1 to regulate CBX8-mediated cell proliferation and tumor metastasis of ovarian cancer. *Hum Cell* 36(6):2113–2128. <https://doi.org/10.1007/s13577-023-00983-7>
1669. Xu Y, Zhang Q, Lin F, Zhu L, Huang F, Zhao L, Ou R (2019) Casiopeina II-gly acts on lncRNA MALAT1 by miR-17-5p to inhibit FZD2 expression via the Wnt signaling pathway during the treatment of cervical carcinoma. *Oncol Rep* 42(4):1365–1379. <https://doi.org/10.3892/or.2019.7268>
1670. Yu X, Li Z, Bai R, Tang F (2022) Transcriptional factor 3 binds to siRTUIN1 to activate the Wnt/ β -catenin signaling in cervical cancer. *Bioengineered* 13(5):12516–12531. <https://doi.org/10.1080/21655979.2022.2076481>
1671. Li F, Wang Z, Lu G (2018) TRIM28 promotes cervical cancer growth through the mTOR signaling pathway. *Oncol Rep* 39(4):1860–1866. <https://doi.org/10.3892/or.2018.6235>
1672. Chan GC, Than WH, Kwan BC, Lai KB, Chan RC, Ng JK, Chow KM, Cheng PM, Law MC et al (2022) Adipose expression of miR-130b and miR-17-5p with wasting, cardiovascular event and mortality in advanced chronic kidney disease patients. *Nephrol Dial Transplant* 37(10):1935–1943. <https://doi.org/10.1093/ndt/gfab287>
1673. Liang J, Bai S, Su L, Li C, Wu J, Xia Z, Xu D (2015) A subset of circulating microRNAs is expressed differently in patients with myocardial infarction. *Mol Med Rep* 12(1):243–247. <https://doi.org/10.3892/mmr.2015.3422>
1674. Su D, Ju Y, Han W, Yang Y, Wang F, Wang T, Tang J (2020) Tcf3-activated lncRNA Gas5 regulates newborn mouse cardiomyocyte apoptosis in diabetic cardiomyopathy. *J Cell Biochem* 121(11):4337–4346. <https://doi.org/10.1002/jcb.29630>
1675. Liu L, Li C, Yu L, Wang Y, Pan X, Huang J (2024) Deciphering the role of SMARCA4 in cardiac disorders: insights from single-cell studies on dilated cardiomyopathy and coronary heart disease. *Cell Signal*. <https://doi.org/10.1016/j.cellsig.2024.111150>
1676. D'Antona S, Porro D, Gallivanone F, Bertoli G (2024) Characterization of cell cycle, inflammation, and oxidative stress signaling role in non-communicable diseases: insights into genetic variants, microRNAs and pathways. *Comput Biol Med* 174:108346. <https://doi.org/10.1016/j.complbiomed.2024.108346>
1677. Ma Z, Lian H, Lin X, Li Y (2021) lncRNA MIAT promotes allergic inflammation and symptoms by targeting MiR-10b-5p in allergic rhinitis mice.

- Am J Rhinol Allergy 35(6):781–789. <https://doi.org/10.1177/1945892421998143>
1678. Li Y, Li R, Li Y, Li G, Zhao Y, Mou H, Chen Y, Xiao L, Gong K (2023) Transcription factor TCF3 promotes macrophage-mediated inflammation and mmp secretion in abdominal aortic aneurysm by regulating miR-143-5p/CCL20. *J Cardiovasc Pharmacol* 82(6):458–469. <https://doi.org/10.1097/FJC.0000000000001484>
1679. Sun MY, Lin JN (2017) Relationship between NR112 polymorphisms and inflammatory bowel disease risk: a systematic review and meta-analysis. *Clin Res Hepatol Gastroenterol* 41(2):230–239. <https://doi.org/10.1016/j.clinre.2016.10.006>
1680. Qin Y, Li Q, Liang W, Yan R, Tong L, Jia M, Zhao C, Zhao W (2021) TRIM28 SUMOylates and stabilizes NLRP3 to facilitate inflammasome activation. *Nat Commun* 12(1):4794. <https://doi.org/10.1038/s41467-021-25033-4>
1681. Santos AS, Ferreira LRP, da Silva AC, Alves LI, Damasceno JG, Kulikowski L, Cunha-Neto E, da Silva MER (2022) Progression of type 1 diabetes: circulating MicroRNA expression profiles changes from preclinical to overt disease. *J Immunol Res* 2022:2734490. <https://doi.org/10.1155/2022/2734490>
1682. Zogg H, Singh R, Ha SE, Wang Z, Jin B, Ha M, Dafinone M, Batalon T, Hoberg N, Poudrier S et al (2023) miR-10b-5p rescues leaky gut linked with gastrointestinal dysmotility and diabetes. *United Eur Gastroenterol J* 11(8):750–766. <https://doi.org/10.1002/ueg2.12463>
1683. Jia SZ, Yang Y, Lang J, Sun P, Leng J (2013) Plasma miR-17-5p, miR-20a and miR-22 are down-regulated in women with endometriosis. *Hum Reprod* 28(2):322–330. <https://doi.org/10.1093/humrep/des413>
1684. Rooda I, Hasan MM, Roos K, Viil J, Andronowska A, Smolander OP, Jaakma Ü, Salumets A, Fazeli A, Velthut-Meikas A (2020) Cellular, extracellular and extracellular vesicular miRNA profiles of pre-ovulatory follicles indicate signaling disturbances in polycystic ovaries. *Int J Mol Sci* 21(24):9550. <https://doi.org/10.3390/ijms21249550>
1685. Brennan E, Butler AE, Drage DS, Sathyapalan T, Atkin SL (2023) Serum polychlorinated biphenyl levels and circulating miRNAs in non-obese women with and without polycystic ovary syndrome. *Front Endocrinol (Lausanne)* 14:1233484. <https://doi.org/10.3389/fendo.2023.1233484>
1686. Chen R, Xin G, Zhang X (2019) Long non-coding RNA HCP5 serves as a ceRNA sponging miR-17-5p and miR-27a/b to regulate the pathogenesis of childhood obesity via the MAPK signaling pathway. *J Pediatr Endocrinol Metab* 32(12):1327–1339. <https://doi.org/10.1515/jpem-2018-0432>
1687. Russo P, Lauria F, Sirangelo I, Siani A, Iacomino G (2023) Association between urinary AGEs and circulating miRNAs in children and adolescents with overweight and obesity from the Italian I family cohort: a pilot study. *J Clin Med* 12(16):5362. <https://doi.org/10.3390/jcm12165362>
1688. Bond ST, King EJ, Henstridge DC, Tran A, Moody SC, Yang C, Liu Y, Mellett NA, Nath AP, Inouye M et al (2021) Deletion of Trim28 in committed adipocytes promotes obesity but preserves glucose tolerance. *Nat Commun* 12(1):74. <https://doi.org/10.1038/s41467-020-20434-3>
1689. Li J, Gan B, Lu L, Chen L, Yan J (2023) Expression of microRNAs in patients with gestational diabetes mellitus: a systematic review and meta-analysis. *Acta Diabetol* 60(4):461–469. <https://doi.org/10.1007/s00592-022-02005-8>
1690. Gui T, Liu M, Yao B, Jiang H, Yang D, Li Q, Zeng X, Wang Y, Cao J, Deng Y et al (2021) TCF3 is epigenetically silenced by EZH2 and DNMT3B and functions as a tumor suppressor in endometrial cancer. *Cell Death Differ* 28(12):3316–3328. <https://doi.org/10.1038/s41418-021-00824-w>
1691. Kaur R, Mehta J, Borges AM (2021) Role of SMARCA4 (BRG1) and SMARCB1 (INI1) in dedifferentiated endometrial carcinoma with paradoxical aberrant expression of mmr in the well-differentiated component: a case report and review of the literature. *Int J Surg Pathol* 29(5):571–577. <https://doi.org/10.1177/1066896920959453>
1692. Chen Y, Cheng H, Long H (2021) Tripartite motif containing 28 (TRIM28) promotes the growth and migration of endometrial carcinoma cells by regulating the AKT/mTOR signaling pathway. *Gen Physiol Biophys* 40(3):245–252. https://doi.org/10.4149/gpb_2021009
1693. Zhong Y, Zhu F, Ding Y (2019) Differential microRNA expression profile in the plasma of preeclampsia and normal pregnancies. *Exp Ther Med* 18(1):826–832. <https://doi.org/10.3892/etm.2019.7637>
1694. Ma H, He Y, Bai M, Zhu L, He X, Wang L, Jin T (2019) The genetic polymorphisms of ZC3HC1 and SMARCA4 are associated with hypertension risk. *Mol Genet Genomic Med* 7(11):e942. <https://doi.org/10.1002/mgg3.942>
1695. Chen M, Shi P, Wang P, Zhang T, Zhao J, Zhao L (2024) Up-regulation of Trim28 in pregnancy-induced hypertension is involved in the injury of human umbilical vein endothelial cells through the p38 signaling pathway. *Histol Histopathol* 39(5):603–610. <https://doi.org/10.14670/HH-18-651>
1696. Liu B, Liu L, Sulaiman Z, Wang C, Wang L, Zhu J, Liu S, Cheng Z (2024) Comprehensive analysis of lncRNA-miRNA-mRNA ceRNA network and key genes in granulosa cells of patients with biochemical primary ovarian insufficiency. *J Assist Reprod Genet* 41(1):15–29. <https://doi.org/10.1007/s10815-023-02937-2>
1697. Abedini A, Landry DA, Macaulay AD, Vaishnav H, Parbhakar A, Ibrahim D, Salehi R, Maranda V, Macdonald E, Vanderhyden BC (2023) SWI/SNF chromatin remodeling subunit Smarca4/BRG1 is essential for female fertility. *Biol Reprod* 108(2):279–291. <https://doi.org/10.1093/biolre/iocac209>
1698. Tang J, Chen Q, Xiang L, Tu T, Zhang Y, Ou C (2024) TRIM28 fosters microglia ferroptosis via autophagy modulation to enhance neuropathic pain and neuroinflammation. *Mol Neurobiol*. <https://doi.org/10.1007/s12035-024-04133-4>

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.